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# Effect of Technique of Disinfection by Ultrasonic Nebulization on Accuracy of Vinyl Polysiloxane Impressions

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## ABSTRACT

**Introduction:** To prevent cross-contamination, impressions should be properly disinfected after removing from the mouth. To be efficacious, a successful disinfection must maintain the physico-chemical properties of the impression materials and should not interfere negatively in the fabrication of stone casts.

**Aim:** To evaluate the effect of different disinfection techniques on the accuracy of dental impressions made of vinyl polysiloxane.

**Materials and Methods:** 40 test samples made of vinyl polysiloxane were obtained using a pattern cylinder. The samples were randomly divided into 5 experimental conditions: I-Control, impressions without any disinfection, II-Immersion in 2% glutaraldehyde for 10 minutes, III-Immersion in 0.2% peracetic acid for 10 minutes, IV-ultrasonic nebulization for 10 minutes in 2% glutaraldehyde and V-ultrasonic nebulization for 10 minutes in 0.2% peracetic acid. The impressions obtained were poured in type IV gypsum and both the height and diameter of the stone casts were measured. For this purpose a profile projector joined to a digital measurement system was used. The data were submitted to statistical analysis using Bioestat 5.3 software. The Shapiro-Wilk test was applied to assess normal

data distribution. Data were analysed by one-way ANOVA and Tukey's test for comparisons of the means of the different groups, p-value less than 0.05 was considered as statistically significant.

**Results:** Groups I, II and III did not differ statistically among themselves, both in diameter and height. Group IV presented statistically different results from the others for diameter. Whereas for height, the results were shown to be similar among groups I, II, III and IV. For Group V, the results obtained were statistically different for both height and diameter from those of the other groups.

**Conclusion:** The immersion technique did not interfere in the accuracy of the stone casts however the ultrasonic nebulization with 2% glutaraldehyde solution did not show significant differences for height and presented better values of dimensional accuracy in diameter, when compared to the control group. The ultrasonic nebulization method associated with 2% peracetic acid solution presented the worst dimensional accuracy values for height and better values for the diameter compared to the other groups.

**Keywords:** Dental impression materials, Dimensional accuracy, Infection control, Siloxanes

## INTRODUCTION

Dental impressions contribute to an important step to get a perfect cast, as the aim of an impression is to produce a dimensionally stable "negative" to serve as the cast mold. The impression materials should reproduce the static and oral structures accurately [1]. Among the options of these materials, dentists have tended to use vinyl polysiloxane because of their improved physical and mechanical properties and good patient acceptance.

To prevent cross-contamination, impressions should be properly disinfected after removing from the mouth, since they are always contaminated with saliva, frequently with blood and bacterial plaque [2]. Manipulation of these contaminated impressions may contribute to the dissemination of causative microorganisms of infectious and contagious diseases [3,4]. At present, there is a variety of chemical products sold as agents suitable for disinfecting dental impressions. To be efficacious, a successful disinfection must maintain the physico-chemical properties of the impression materials and also not interfere negatively in the obtainment of stone casts [5].

Among the chemical disinfectants available, glutaraldehyde has been widely used because it presents compatibility with many impression materials and has also demonstrated a bactericidal and fungicidal effect and virucidal action [6,7]. Nevertheless, glutaraldehyde releases toxic vapors, irritants and allergens that cause irritation to the eyes, nose and throat, allergy, contact dermatitis, asthma and rhinitis [8].

A safe alternative to glutaraldehyde, as an effective disinfectant, is the peracetic acid [9], which is a powerful microbicidal agent used for high-level disinfection in hospitals. According to the FDA's Classification of Sterilising and Disinfecting Chemical Liquids published in the Federal Register, the peracetic acid is a mild, non-toxic and non-allergenic irritant, being recommended for high-level disinfection processes [10]. However, its use to disinfect dental impressions is still scarcely mentioned in the literature.

In addition to the impression materials and the chemical agents used during the disinfection process, professionals should consider the technique of choice as well. Therefore, in order to obtain decontaminated stone casts, these three factors should be interdependent factors. As already mentioned, the immersion impression technique has been the widely applied; however, studies have described a new method of disinfection by ultrasonic nebulization for dental impressions [11-14].

The nebulization process has been used for medical treatment purposes. Nebulization is characterised by dispersion of liquid into the air [12]. Ultrasonic nebulizers use a piezoelectric crystal that emits ultrasonic waves to produce aerosol. This method requires only around 10 mL of the disinfection solution for each cycle [12]. When used for disinfection, the interaction of the effect of ultrasonic nebulization and the chemical agent may potentiate its action; ultrasonic nebulization with 2% glutaraldehyde has shown higher microbicidal activity when compared to the immersion method using the same chemical agent [13].

Many studies have evaluated comparatively different mold disinfection techniques [15-18]. However, despite microbiological evaluation of the ultrasonic nebulization effect with peracetic acid and glutaraldehyde on disinfection methods of dental impressions, literature information about the effect of the nebulization method on the dimensional precision of dental impressions is scarce [13]. In addition, there were no studies found evaluating the accuracy of VPS immersed in peracetic acid solution.

Based on the author's earlier study [13], which followed a similar methodology and evaluated dental impressions microbiologically, disinfected by nebulization and immersion methods using peracetic acid and glutaraldehyde, the present study aimed to investigate the effect of these methods on the accuracy of impressions made with polyvinyl siloxane. The null hypothesis of the present study was that both disinfection methods would not affect the dimensional precision of polyvinyl siloxane impressions when compared to control.

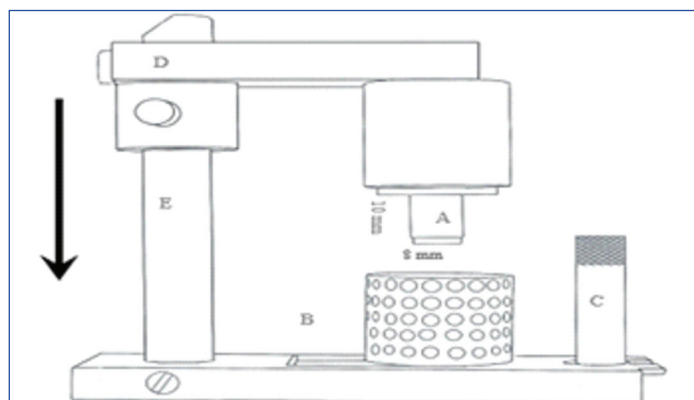
## MATERIALS AND METHODS

The study was conducted between January and June 2010 in Cascavel Campus of State University of Western Paraná (UNIOESTE), Paraná, Brazil. The sample was calculated using a family *F* probability, with a repeated families design, with interaction within and among the factors. The effect size of 0.18, type 1 ( $\alpha$ ) error of 0.05, and analysis power of 0.90 chosen resulted in 40 sample units, with 8 samples per experimental group ( $n=8$ ). G Power software (version 3.1.9.2, University of Düsseldorf, Germany) was used for sample calculation. Since this was an in-vitro study, ethical clearance was not found necessary to be obtained. Thus ethical clearance was not obtained. The materials used in this study are shown in [Table/Fig-1].

Material	Commercial brand-manufacturer
Vinyl polysiloxane	Aquasil Easy Mix Putty, Dentsply Caulk, Milford, USA.
	Aquasil Ultra LV, Dentsply Caulk, Milford, USA.
0.2% peracetic acid	Sterilife, Lifemed Ind. Equip. Art. Med. Hosp. S.A., Pelotas, RS, Brazil.
2% glutaraldehyde	Glutaron II, Ind. Farm. Rioquímica Ltda., Sao Jose do Rio Preto, SP, Brazil.
Type IV gypsum	Durone IV, Dentsply Ind. e Com. Ltda, Petropolis, RJ, Brazil.

[Table/Fig-1]: Materials used and manufacturers.

The accuracy of impressions submitted to different disinfection techniques was evaluated by means of stone casts dimensions obtained from a stainless steel pattern model [14-18]. This pattern model was used to obtaining cylindrical samples, with 8 mm in diameter and 10 mm in height [Table/Fig-2]. Joined to it, there was a mobile device that ran along a vertical bar, allowing the cylinder to meet with a tray containing the impression material in a standardised thickness. The trays were prepared with 20-mm-height and 20-mm-internal diameter. This mobile device allowed standardisation of pressure used during the act of taking the impression [Table/Fig-2].



[Table/Fig-2]: Diagram illustrating the stainless steel pattern apparatus used for the impression. A- pattern model, B- impression tray, C- pin for fixing tray, D- mobile device, E- fixed vertical bar.

In addition to the technique for disinfection by immersion for 10 minutes, the technique for disinfection by ultrasonic nebulization was also evaluated. This technique was performed using an ultrasonic nebulizer (Pulmonosc Star II, Soniclear, São Paulo, São Paulo, Brazil), set at an ultrasonic frequency of 2.4 MHz, nebulization rate of 1.25 cc/min, and the disinfectant solution mist was guided into a transparent plastic box (20×20×25 cm). The samples in the box were disinfected by means of ultrasonic nebulization and kept for 10 minutes until the fog in the box reached saturation [10]. The solutions used were 0.2% peracetic acid (Sterilife, Lifemed Ind. Equip. Art. Med. Hosp. S.A.) and 2% glutaraldehyde (Glutaron II, Ind. Farm. Rioquímica Ltda).

The impressions were made with vinyl polysiloxane (Aquasil, Dentsply Caulk, Milford, USA.) by means of the one-step impression technique, made with the putty and light-body materials simultaneously, because this technique is one of the most used in clinical dentistry. The impressions were allowed to polymerize on the stainless steel pattern model for 12 minutes [19] and remained in a controlled environmental condition at 23±1°C. The putty consistency VPS material was proportioned and manipulated based on the manufacturer's instructions. The light-body material was used with an automatic mixing syringe. The 40 impressions obtained were washed under running water for 10 seconds and then randomly submitted to one of the 5 experimental conditions ( $n=8$ ), according to [Table/Fig-3].

Groups	Disinfection technique
I	Control-impressions without any disinfection.
II	Immersion in 2% glutaraldehyde for 10 minutes.
III	Immersion in 0.2% peracetic acid for 10 minutes.
IV	Nebulization with 2% glutaraldehyde for 10 minutes.
V	Nebulization with 0.2% peracetic acid for 10 minutes.

[Table/Fig-3]: Experimental groups.

After disinfection period, the impressions were again washed under filtered running water for 10 seconds and dried with air jets. After the period of 1 hour, in order to favor the release of hydrogen, the impressions were filled with type IV gypsum (Durone IV, Dentsply Ind. e Com. Ltda, Petropolis, RJ, Brazil.) to obtain the stone casts. The gypsum was manipulated in accordance with the manufacturer's instructions. To mix the gypsum, a mechanical vacuum mixer (A 300, Polidental, Cotia, São Paulo, Brazil) was used for 60 seconds. The gypsum was poured into the impression under vibration, in small quantities, until it was completely filled. After the gypsum had set (1 hour), the stone cast was separated from the impression, identified, and remained in a controlled environmental condition at 23±1°C, relative humidity between 40 and 60%, for 24±1 hour. After this period, the measurements of the stone casts were taken.

The stone casts were measured in a profile projector (VB300/P, Starrett, Athol, Massachusetts, USA) coupled to a digital measurement system (Quadra Check 200, Metronics, Mentor, Ohio, USA), with a accuracy of 0.001 mm, taking the measurements of the height (*h*) and diameter (*d*) of each specimen. Each stone cast was measured six times and an arithmetic mean was obtained for each sample. That way, the evaluation of the test specimen dimensional alteration (%) was verified by the formula:

$$\Delta L = 100 \times \{(L1 - L2) / L1\}$$

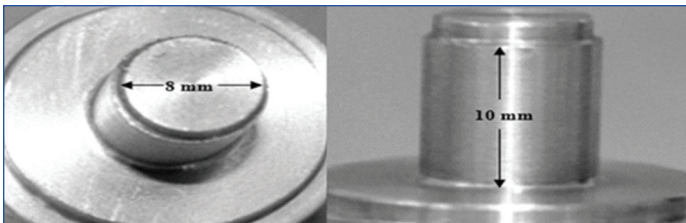
$\Delta L$ -dimensional alteration.

L1-stone cast dimension.

L2-dimension of the stainless steel pattern [Table/Fig-4].

## STATISTICAL ANALYSIS

The data were submitted to statistical analysis using Bioestat 5.3 software (Mamirauá Institute, AM, Brazil, 2007). The Shapiro-Wilk



**[Table/Fig-4]:** Stainless steel pattern model used for obtaining the specimens.

test was applied to assess normal data distribution. Data were analysed by one-way ANOVA and Tukey's test for comparisons of the means of the different groups,  $p < 0.05$  was considered statistically significant.

## RESULTS

The mean values for dimensional alteration obtained from the measurement of the test specimen diameters are shown in [Table/Fig-5]. Statistical analysis showed that groups I, II, and III had the highest means of dimensional alteration and did not differ among them. The group IV showed intermediate mean and differed statistically from the others, the group V differed statistically from the other groups and presented the lowest mean of dimensional alteration. Statistically significant differences were obtained on inter-group comparison using ANOVA ( $p = 0.025$ ).

Group	Mean (SD)
I- Control-impresions without any disinfection	-1.63 (0.48) a
II- Immersion in 2% glutaraldehyde	-1.61 (0.41) a
III- Immersion in 0.2% peracetic acid	-1.59 (0.58) a
IV- Nebulization with 2% glutaraldehyde	-0.89 (0.29) b
V- Nebulization with 0.2% peracetic acid	-0.27 (0.08) c

**[Table/Fig-5]:** Mean values and standard deviation (SD) of dimensional alteration (%) in diameter.  
Different letters in the same row mean statistically significant differences,  $p < 0.05$

The mean values for dimensional alteration verified from measurement of test specimen heights are shown in [Table/Fig-6]. Statistical analysis of these results demonstrated that groups I, II, III, and IV did not differ among them, while the group V showed the highest mean dimensional alteration. Statistically significant differences were obtained on inter-group comparison using ANOVA ( $p = 0.022$ ).

Group	Mean (SD)
I- Control-impresions without any disinfection	-1.39 (0.33) a
II- Immersion in 2% glutaraldehyde	-1.47 (0.24) a
III- Immersion in 0.2% peracetic acid	-1.48 (0.29) a
IV- Nebulization with 2% glutaraldehyde	-1.58 (0.26) a
V- Nebulization with 0.2% peracetic acid	-2.09 (0.54) b

**[Table/Fig-6]:** Mean values and standard deviation (SD) of dimensional alteration (%) in height.  
Different letters in the same row mean statistically significant differences,  $p < 0.05$

## DISCUSSION

Evaluation of the dimensional precision of stone casts obtained from impressions submitted to greatly differing processes of disinfection have been reported in scientific literature. The majority concluded that disinfection methods do not significantly alter the dimensional precision of stone casts [20]. However, no studies were found, which evaluated the dimensional precision of stone casts obtained from impressions disinfected with peracetic acid and the ultrasonic nebulization technique.

In this study, the null hypothesis was rejected, since the groups that were disinfected by nebulization presented different dimensional alteration when compared to control group. The results of the present study demonstrated that the impressions submitted to immersion, both in glutaraldehyde and in peracetic acid, as well

as the control group, showed no statistically significant difference in their accuracy. Previous studies have found similarity among the experimental groups, immediately after using different disinfectant solutions with VPS, by immersion technique [20,21].

In the study carried out by Chen SY et al., a pattern model and a metal tray, similar to the ones used in this current study, were used for evaluation [14]. The accuracy differences, measured in stone casts from impressions made from several impression materials, were evaluated, however, not undergoing disinfection processes. In the above-mentioned study, the dimensional alteration values for the contraction percentage (%) of polyvinyl siloxane varied between  $1.00 \pm 0.79$ – $1.35 \pm 0.77$ .

The results found in this study showed that there was a contraction in all the evaluated stone casts compared to the stainless steel standards, which is a common finding in the literature [14,22]. Such results were observed as, during the polymerization reaction, the impression material retracted from the trays.

In the absence of a tray adhesive, there would be unrestricted polymerization shrinkage of the impression material, resulting in a cast that is smaller in diameter and height [21]. The tray adhesive was not used in this study because the trays had mechanically retentive features [Table/Fig-2]. In addition to this, the mechanical spatulation of the gypsum can be contributed to a lower setting expansion and, consequently, lower dimensional alteration of specimens.

The accuracy evaluation of dental VPS impressions submitted to ultrasonic nebulization techniques, using glutaraldehyde or peracetic acid solution, has not been reported in the literature yet. In this study, the analysis of the dimensional alterations of the test specimen's diameter has shown that the ultrasonic nebulization, using glutaraldehyde solution, has shown statistically different values in relation to all the other experimental groups, presenting smaller dimensional alteration than the control group (I) and the immersion groups (II and III). On the other hand, the group of ultrasonic nebulization, using Sterilife™, has presented smaller dimensional alterations than all the groups. However, considering the height dimensional alteration of the samples, greater dimensional alterations than all the groups was found. Thus, ultrasonic nebulization, using peracetic acid, has shown the worst (2.09% for height) and the best results (0.27% for the diameter) for the dimensional changes in relation to the other groups. Therefore, it can be said that the interaction between the peracetic acid solution and ultrasonic nebulization processes interfered negatively in the accuracy of the models, causing shrinkage.

During ultrasonic nebulization, the concentration of the nebulized solution increases [11]. This is caused by water evaporation during the aerosol output [23]. Corroborating with these statements, Steckel H et al., have found that changes in the droplet size, the surface tension, the viscosity and the saturated vapor pressure, during the ultrasonic nebulization process, can be caused by the temperature and the solution concentration in the nebulizer reservoir [24]. Such changes generally lead to an increase in the solutions' concentration, which may additionally increase the molecule of the pharmacological solutions, which could, potentially, increase the adverse reactions to the nebulized liquid. Considering that the Sterilife® solution was used with ultrasonic nebulization, the droplet size may be changed, and the saturated vapor pressure increased, increasing the absorption effect of this solution in the polymer matrix of the VPS, causing a greater dimensional change to the stone mold. This situation, when transferred to the clinical practice, could act negatively in obtaining the stone mold used in the preparation of prosthetic crowns.

The selection of a disinfection technique should be established based on several criteria, besides the accuracy of the obtained stone casts, including the microbicidal capability of the disinfectant solution. In a study carried out by Mendonça MJ et al., the authors have evaluated the microbicidal capacity of 2% glutaraldehyde



and Sterilife® solutions, applying disinfection methods: ultrasonic nebulization and immersion [13]. In this research study, the peracetic acid has shown microbicidal efficacy for both methods, and the glutaraldehyde has shown total microbicidal activity only for the impressions submitted to ultrasonic nebulization. Considering the results of the current laboratory study and the work by Mendonça MJ et al., it may be suggested that, in dental offices, the application of ultrasonic nebulization should be used, preferably, with glutaraldehyde solutions, whereas peracetic acid should be used with immersion techniques [13].

## LIMITATION

The present study was an in vitro study; therefore it is necessary that future clinical studies should evaluate the difference of dimensional alteration when subjected to different disinfection techniques that influence the adaptation and indirect restorations.

## CONCLUSION

It can be concluded that the immersion technique both in the glutaraldehyde and peracetic acid solutions did not interfere in the accuracy of the stone casts obtained, in comparison to the control group. The ultrasonic nebulization method associated with 2% glutaraldehyde solution did not differ significantly for height and even presented better values of dimensional accuracy of diameter compared to the control group. The ultrasonic nebulization method associated with 2% peracetic acid solution presented the worst values for height and better dimensional accuracy values for the diameter.

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## REFERENCES

- [1] Matys J, Swider K, Flieger R. Laser instant implant impression method: A case presentation. *Dent Med Probl.* 2017;54:101-06.
- [2] Ud Din S, Parker S, Braden M, Tomlins P, Patel M. Experimental hydrophilic vinyl polysiloxane (VPS) impression materials incorporating a novel surfactant compared with commercial VPS. *Dent Mater* 2017;33:e301-09.
- [3] Centers for Disease Control and Prevention: Guidelines for Infection Control in Dental-Health-Care Settings MMWR. Centers for Disease Control 2003;52:1-76.
- [4] Celebi H, Buyukerkmen EB, Torlak E. Disinfection of polyvinyl siloxane impression material by gaseous ozone. *J Prosthet Dent.* 2018;120:138-43.
- [5] Sinobad T, Obradovic-Djuricic K, Nikolic Z, Dodic S, Lazic V, Sinobad V. The effect of disinfectants on dimensional stability of addition and condensation silicone impressions. *Vojnosanit Pregl* 2014;71:251-58.
- [6] Rabenau HF, Steinmann J, Rapp I, Schwebke I, Eggers M. Evaluation of a virucidal quantitative carrier test for surface disinfectants. *PLoS One.* 2014;27:e86128.
- [7] Jayapalan V, Krishnan CS, Ramasubramanian H, Sampathkumar J, Azhagarasan NS, Krishnan M. Comparative evaluation of the antimicrobial efficacy of three immersion chemical disinfectants on clinically derived poly (vinyl siloxane) impressions. *J prosthodont.* 2018;27:469-75.
- [8] Chidambaranathan AS, Balasubramaniam M. Comprehensive review and comparison of the disinfection techniques currently available in the literature. *J Prosthodont.* 2017;1-8.doi: 10.1111/jopr.12597. [Epub ah ad of print].
- [9] Costa AS, de Paula OFF, Silva CRG, Leão MVP, Santos SSF. Stability of antimicrobial activity of peracetic acid solutions used in the final disinfection process. *Braz Oral Res.* 2015;29:01-06.
- [10] Food and Drug Administration. General hospital and personal use devices: proposed classification of liquid chemical sterilants and general purpose disinfectants-FDA. Proposed rule. *Fed Regist.* 1998;63:59917-21.
- [11] Wu G, Yu X, Gu Z. Ultrasonically nebulised electrolysed oxidising water: a promising new infection control programme for impressions, metals and gypsum casts used in dental hospitals. *J Hosp Infect.* 2008;68:348-54.
- [12] Wanner A, Rao A. Clinical indications for and effects of bland, mucolytic, and antimicrobial aerosols. *Am Rev Respir Dis.* 1980;122:79-87.
- [13] Mendonca MJ, Rafael RS, Camilotti V, Menolli RA, Sicoli EA, Teixeira N, et al. Microbiological evaluation of the ultrasonic nebulization effect with peracetic acid and glutaraldehyde on disinfection methods of dental impressions. *Gen Dent.* 2013;61:10-13.
- [14] Chen SY, Liang WM, Chen FN. Factors affecting the accuracy of elastometric impression materials. *J Dent.* 2004;32:603-09.
- [15] Nandini Y, Vinitha KB, Manvi S, Smitha MJ. Comparison of dimensional accuracy of four different die materials before and after disinfection of the impression: an in vitro study. *Contemp Dent Pract.* 2013;14:668-74.
- [16] Nassar U, Chow AK. Surface detail reproduction and effect of disinfectant and long-term storage on the dimensional stability of a novel vinyl polyether silicone impression material. *J Prosthodont.* 2015;24:494-98.
- [17] Nassar U, Flores-Mir C, Heo G, Torrealba Y. The effect of prolonged storage and disinfection on the dimensional stability of 5 vinyl polyether silicone impression materials. *J Adv Prosthodont.* 2017;9:182-87.
- [18] Samra RK, Bhide SV. Comparative evaluation of dimensional stability of impression materials from developing countries and developed countries after disinfection with different immersion disinfectant systems and ultraviolet chamber. *Saudi Dent J.* 2018;30:125-41.
- [19] Caputi S, Varvara G. Dimensional accuracy of resultant casts made by a monophasic, one-step and two-step, and a novel two-step putty/light-body impression technique: an in vitro study. *J Prosthet Dent.* 2008;99:274-81.
- [20] Martin N, Martin MV, Jedynekiewicz NM. The dimensional stability of dental impression materials following immersion in disinfecting solutions. *Dent Mater.* 2007;23:760-68.
- [21] Guiraldo RD, Berger SB, Siqueira RM, Grandi VH, Lopes MB, Gonini-Júnior A, et al. Surface detail reproduction and dimensional accuracy of molds: influence of disinfectant solutions and elastomeric impression materials. *Acta Odontol Latinoam.* 2017;30:13-18.
- [22] Ceyhan JA, Johnson GH, Lepe X. The effect of tray selection, viscosity of impression material, and sequence of pour on the accuracy of dies made from dual-arch impressions. *J Prosthet Dent.* 2003;90:143-49.
- [23] Ip AY, Niven RW. Prediction and experimental determination of solute output from a Collision nebulizer. *J Pharm Sci.* 1994;83:1047-51.
- [24] Steckel H, Eskandar F. Factors affecting aerosol performance during nebulization with jet and ultrasonic nebulizers. *Eur J Pharm Sci.* 2003;19:443-55.

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