



UNIVERSIDADE ESTADUAL DE CAMPINAS  
FACULDADE DE CIÊNCIAS MÉDICAS

DANIELE BRAZ TORRES

**Análise dos efeitos da restrição proteica *in utero* no BNST e na  
Amígdala de ratos: Estudo da estrutura dendrítica neural, de  
parâmetros funcionais e moleculares.**

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Tese apresentada à Faculdade de Ciências Médicas da Universidade Estadual de Campinas como parte dos requisitos exigidos para a obtenção do título de Doutora em Ciências

**Orientadora:** Profa. Dra. Patricia Aline Boer

**Coorientador:** Prof. Dr. Jose Antonio Rocha Gontijo

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*Dedico este trabalho com todo meu amor e gratidão aos meus pais, José Messias  
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*“Tudo posso N a quele que me fortalece.”*

*Filipenses 4:13*

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## RESUMO

O núcleo da estria terminal (BNST) e a amígdala têm sido associados à modulação do comportamento ansioso e do medo. Devido à sua plasticidade, tanto a morfologia quanto a composição de neurotransmissores destas regiões podem ser influenciadas por eventos adversos durante o período perinatal. A restrição proteica *in utero*, com consequente exposição do feto a concentração elevada de glicocorticóides (GC), está associada à desregulação do eixo HPA e a resposta exacerbada ao estresse na vida adulta. Recentemente, um estudo conduzido em nosso laboratório demonstrou que a restrição proteica gestacional promove alterações neuroquímicas e morfológicas no BNST que se associam ao comportamento de ansiedade em ratos machos adultos. Entretanto, neste estudo não foi possível afirmar se as alterações encontradas são de fato decorrentes da programação gestacional ou secundárias a outros fatores desconhecidos que poderiam determinar sua ocorrência na vida adulta. Assim, o objetivo do presente estudo foi investigar, em animais com idade precoce, os possíveis efeitos da restrição proteica gestacional durante o desenvolvimento do BNST e da amígdala. Nosso estudo mostrou que a prole submetida à restrição proteica gestacional apresenta diminuição significativa do peso corporal ao nascer, que persiste reduzido até o 14º dia de vida. Além disso, a quantificação das células do BNST e os estudos esteriológicos mostram redução significativa do número total de células e do volume da região antero-dorsal do BNST em animais do grupo LP, no 14º dia de vida em comparação ao grupo NP de mesma idade. Houve também redução no comprimento e na arborização dendrítica dos neurônios do BNST na prole LP. Demonstramos ainda redução na expressão de receptores de glicocorticóides (GR) e mineralocorticóides (MR) nos animais do grupo LP com 7 e 14 dias de vida, associada a redução na expressão do BDNF e aumento na expressão do receptor CRF1 no BNST de animais do grupo LP com 7 dias de vida, apesar de não ter havido alteração nos níveis plasmáticos de corticosterona. Nos animais do grupo LP com 14 dias observamos redução na expressão dos receptores 5HT1A e aumento na expressão dos receptores 5HT2A nos animais do grupo LP com 7 e 14 dias de vida. Nossos resultados também demonstraram aumento significativo

da concentração de norepinefrina e 5HIAA no BNST e redução do *turnover* de dopamina e seu precursor DOPAC no BNST, respectivamente, nos animais do grupo LP com 7 e 14 dias. Quanto à amígdala nós não observamos alterações significativas no comprimento e no número de ramificações dos dendritos de neurônios localizados na amígdala basolateral dos animais do grupo LP com 14 dias comparativamente aos animais do grupo NP de mesma idade. Da mesma forma, o número de células na amígdala foi similar nos animais dos grupos LP e NP no 14º dia. Houve redução significativa na concentração de norepinefrina, epinefrina e dopamina bem como na expressão de CRF e BDNF na amígdala de animais do grupo LP no 7º dia, assim como houve redução na expressão de GR, MR e CRF na amígdala dos animais do grupo LP no 14º dia. Em conclusão, esta é a primeira descrição na literatura da ocorrência de modulação da plasticidade, tanto com relação ao número de neurônios e outros tipos celulares quanto da neuroquímica do BNST precocemente, decorrente da restrição proteica gestacional. Além disso, as alterações neuroquímicas observadas na amígdala e no BNST podem contribuir para as alterações comportamentais induzidas pela restrição proteica gestacional bem como alterações em outras regiões cerebrais. Estas diferenças podem representar a adaptação que ocorre durante o desenvolvimento embrionário às altas concentrações de glicocorticóides maternos e pode estar relacionada à hiperatividade do eixo HPA e ao comportamento de ansiedade observado na idade adulta em indivíduos submetidos à restrição proteica gestacional.

**Palavras chaves:** BNST, amígdala, programação fetal, receptores esteróides, CRF, BDNF, serotonina, dopamina, catecolamina, análise dendrítica.

## ABSTRACT

The bed nucleus of the stria terminalis (BNST) and amygdala have been associated with the modulation of anxiety-like behavior and fear. Due to their plasticity, its morphology and neurotransmitter compounds may be affected by perinatal adverse events such as protein restriction *in utero*, when the fetus is exposed to high maternal glucocorticoids (GC) levels, leading to a deregulation of HPA axis and exacerbated stress answer in adult life. We have recently demonstrated that gestational protein-restricted intake causes neurochemical and morphological changes in the BNST associated with anxiety-like behavior in male offspring adulthood. However, our study did not allow asserting the full mode whether these disorders are from gestational programming or secondary to other factors that could determine these changes in adulthood. Thus, our objective was investigating the possible effects of gestational protein restriction on development of BNST and amygdala in early age. The current study shows a significant decrease in body birth weight that remains up to 14 day of age, in gestational protein-restricted offspring. Otherwise, BNST cell quantification and stereology studies show a significant reduction of the total cells number associated with reduced volume of the anterodorsal BNST division in 14 d-old LP offspring compared to age-matched NP group. Also, we found reduction of the dendritic length and arborizations in the BNST neurons of the LP offspring. The present work demonstrates a decreased expression of gluco- and mineralocorticoid receptors (GR and MR) in 7 and 14-d old accompanied, by fall in BDNF and enhanced CRF1 receptor expression in the BNST of the 7-d old LP offspring, despite of unchanged corticosterone plasma level. Additionally, the 14 d-old LP offspring presents a reduced 5HT<sub>1A</sub> receptor subtype levels, reciprocally accompanied, by increased 5HT<sub>2A</sub> receptors in the 7 and 14-d old LP. Our findings also show a significant increase in the BNST norepinephrine and 5HIAA levels and, reduced dopamine turnover and 3,4-DOPAC BNST levels, respectively in 7-d old LP and 14 d-old LP. In complementary study we did not observe any significant changes in the basolateral amygdala neuronal dendrites length and ramifications in 14 d-old NP compared with age-matched gestational protein-restricted offspring.

Also, the amygdala cells number was similar in 14 day-old LP and NP offspring. We demonstrate a significant decrease in the amygdala content of norepinephrine, epinephrine, dopamine, CRF and BDNF in 7 day-old rats, as well as reduction in GR, MR and CRF expression in 14 day-old LP offspring. In conclusion, as far as we know is the first description of the modulation of neuron and non-neuron plasticity and neurochemistry of the BNST in the early life, by gestational protein-restricted intake. Also, the BNST and amygdala neurochemical changes observed in the current study may contribute to behavioral alterations induced by gestational protein restriction and these may be a primer for alterations in other brain regions. These findings may represent the adaptation during embryonic development to elevate maternal corticosteroids exposure and could be related to HPA axis hyperactivity and anxiety-like behavior observed in maternal protein-restricted offspring adulthood.

**Keywords:** BNST, amygdala, fetal programming, steroid receptors, CRF, BDNF, serotonin, dopamine, catecholamine, dendritic analysis

## **LISTA DE SIGLAS E ABREVIATURAS**

**OMS** Organização Mundial da Saúde

**BPN** baixo peso ao nascer

**RCIU** retardo do crescimento fetal intra-utero

**SNC** sistema nervoso central

**11 $\beta$ -HSD<sub>2</sub>** enzima 11 $\beta$ -hidroxiesteróide desidrogenase tipo 2 placentária

**HPHA** eixo hipocampo-hipotálamo-pituitária-adrenal

**GC** glicocorticoides

**MR** Receptores de mineralocorticóides

**ACTH** Hormônio adrenocorticotrófico

**PVN** núcleo paraventricular

**BNST** núcleo da estria terminal

**CRF** hormônio liberador de corticotrofina

**VTA** núcleo ventral tegmentar

**CeA** núcleos central da amígdala

**CeM** núcleo medial da amígdala medial

**BNSTL** divisão lateral do BNST

**MeA** amígdala medial

**BNSTM** divisão medial do BNST

**BLA** núcleo basolateral da amígdala

**BLAC** parte caudal do BNST

**TDH** déficit de atenção e hiperatividade

**CRF1** fator liberador de corticotropina receptor 1

**5-HT** serotonina

**5-HT1-7** família de receptores de serotonina dos tipos 1 a 7

**5-HT1A** receptor de serotonina tipo 1A

**BDNF** fator neurotrófico derivado do cérebro

**ICVS** Instituto de Ciências da Vida

**DGAV** Direção Geral de Alimentação e Veterinário

**NP** *Normal Protein*

**LP** *Low Protein*

**DAG** distância ano-genital

**µm** micrometros

**NP7D** *Normal Protein* sete dias

**NP14D** *Normal Protein* quatorze dias

**LP7D** *Low Protein* sete dias

**LP14D** *Low protein* quatorze dias

**PFA** paraformolaldeído

**WB** *western blotting*

**RIPA** Radio Immuno Precipitation Assay Buffer

**rpm** rotações por minuto

**HPLC / CE** cromatografia líquida de alto desempenho, combinada com detecção eletroquímica

**COBEA** Colégio brasileiro de experimentação animal

**CEEA** comissão de ética na experimentação animal

**CEMIB** Unicamp Centro Multidisciplinar para Investigações Biológicas da Unicamp

**RIA** radioimunoensaio

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## 1. INTRODUÇÃO

Embora o número de pessoas que passam fome no mundo tenha caído ele ainda é inaceitável, abrangendo 795 milhões de pessoas. A prevalência da desnutrição em regiões em desenvolvimento também vem diminuindo, mas ainda afeta 12,9% da população. No Brasil são 3,4 milhões de pessoas subalimentadas apesar da queda de 82% entre 2002 e 2014 (FAO, 2015).

Tanto em países desenvolvidos quanto naqueles em desenvolvimento a desnutrição materno-infantil tem repercussões evidentes sobre a saúde da população. A desnutrição não é causada apenas pela falta de alimentos, mas também pela dieta ocidental que prioriza gordura e carboidratos em detrimento de proteínas e vitaminas.

De acordo com dados de 2013 da Organização Mundial da Saúde (OMS), a desnutrição materna continua sendo um problema de saúde pública mundial e está relacionada ao baixo peso ao nascer (BPN). Anualmente nascem cerca de 20 milhões de crianças com BPN em todo mundo, representando cerca de 15,5% de todos nascimentos. O baixo peso ao nascer foi incluído no último relatório da OMS que definiu uma meta global de reduzir o BPN em 30% até 2025.

O período pré-natal constitui uma convergência crítica dos fatores de curto e longo prazo que afetam a saúde ao longo da vida (Procter & Campbell, 2014). A quantidade inadequada de nutrientes essenciais durante os períodos cruciais de desenvolvimento fetal pode levar a reprogramação dos tecidos fetais, predispondo o indivíduo a doenças crônicas na vida adulta fundamentando o conceito da programação fetal (Ashton, 2000). O pesquisador inglês David Barker et al (1989) foi pioneiro em afirmar que a nutrição durante o período intrauterino e a infância estava associada à risco aumentado para o aparecimento de doença cardíaca no adulto e, alguns anos mais tarde também afirmou que crianças que nascem com baixo peso tem maior risco de desenvolver doença cardiovascular quando adultas, independente da exposição a outros fatores de risco clássicos como obesidade,

sedentarismo e tabagismo. Estas constatações levaram Barker, em 1989, a interpretar o ambiente fetal como um novo componente na etiologia das doenças cardiovasculares e fundamentaram aquela que ficou conhecida como hipótese de Barker. Posteriormente, Barker (1995; 1997; 1998) obteve novas evidências da associação entre o baixo peso ao nascer e o desenvolvimento de doenças cardiovasculares no adulto, fortalecendo a hipótese de que o retardo do crescimento intra-útero (RCIU) aumenta o risco de desenvolver doenças cardiovasculares na fase adulta.

Entretanto, os primeiros estudos são da década de 60, quando Rose (1964) mostrou que na família de pacientes com doença isquêmica cardíaca, havia o dobro de registros de irmãos natimortos e de mortes prematuras na infância em relação aos demais pacientes. Forsdahl, em 1967 demonstrou que os índices de mortalidade infantil em anos anteriores correlacionavam-se à prevalência de doença cardíaca aterosclerótica daquele ano na Noruega.

Roseboom e colaboradores, (2006) fizeram um levantamento importante de indivíduos que foram gestados durante o período da “Fome Holandesa”. Este foi um período (1944-45) de escassez intensa de alimentos imposta pelo exército alemão durante a II Guerra Mundial. O suprimento foi cortado inclusive para crianças, lactantes e mulheres grávidas, que normalmente tinham direito a uma quantidade extra. Eles observaram que estes indivíduos apresentaram baixo peso ao nascer e redução da circunferência do crânio e quando adultos os mesmos apresentaram desequilíbrio metabólico. Mais tarde foram estabelecidas associações entre o surgimento de doenças em adultos e o período de exposição à fome durante a gestação e ficou evidente que aqueles expostos a fome logo no início da gestação apresentavam complicações maiores quando adultos (Painter et al, 2005; Roseboom et al, 2001).

Porém, apenas no ano de 1991, o conceito de programação fetal foi definido por Alan Lucas, como sendo a “*resposta permanente do organismo a um estímulo ou insulto durante um período crítico do desenvolvimento embrionário e fetal*”. Portanto, o ambiente materno atua como uma previsão das condições que o

feto irá encontrar após o nascimento. O feto responde e adapta-se a esta previsão usando estratégias a fim de maximizar sua chance de sobrevivência (Hales, 2013).

Desde então vários modelos experimentais de subnutrição gestacional foram desenvolvidos, nos quais o baixo peso da prole ao nascer foi associado à elevação pressórica na idade adulta (Persson & Jansson, 1992; Woodall et al, 1996; Prentice 1991; Godfrey et al, 1996). Dentre os modelos experimentais têm sido bastante utilizada variações no conteúdo de proteína da dieta gestacional para produzir restrição leve (12% de caseína), moderada (9%) e severa (6%) (Langley-Evans et al, 1996). Esses experimentos resultaram em alterações variáveis no peso dos recém-natos e no tamanho das placentas. Os animais submetidos à restrição desenvolveram hipertensão arterial a partir da quarta semana de vida a qual foi mantida até a idade adulta (Langley-Evans et al, 1994; 1995).

As proteínas são formadas por agrupamentos de aminoácidos resultantes da ingestão de dietas e são necessárias para diversas funções do sistema nervoso central (SNC), dentre outros. Assim o déficit de proteína pode afetar diretamente o desenvolvimento cerebral (Lima et al, 1993; Morgane et al, 2002).

Diversos trabalhos demonstram que o baixo peso no nascimento é um fator de risco para o desenvolvimento de desordens psicológicas relacionadas ao desenvolvimento neural, tais como o autismo (Burd et al, 1999) e depressão podendo levar ao suicídio (Barker et al, 1995). Entretanto, Welberg & Seckl (2001) demonstraram que o baixo peso no nascimento não é um fator crucial *per se* servindo, provavelmente, como um marcador de possíveis efeitos deletérios durante o desenvolvimento do sistema nervoso.

Existem diversos fatores que podem estar envolvidos na gênese da programação fetal, no entanto, o mais estudado é a diminuição na concentração e atividade da enzima 11 $\beta$ -hidroxiesteróide desidrogenase tipo 2 (11 $\beta$ -HSD<sub>2</sub>) placentária (Benediktsson et al, 1993; Stewart et al, 1995; Langley-Evans et al,

1996; Langley-Evans, 1997). A diminuição na  $11\beta$ -HSD<sub>2</sub> placentária, devida à restrição proteica gestacional, resulta no aumento da exposição fetal aos corticosteróides endógenos maternos (Langley-Evans et al., 1996). Normalmente o feto é exposto a 10%-20% de cortisol e este cortisol presente na circulação fetal é uma combinação do cortisol produzido pelo feto mais o cortisol materno derivado da placenta materna (Seth et al, 2015).

O cérebro é extremamente sensível à programação pré-natal e a exposição intra-uterina a glicocorticoides tem influência ampla na estrutura nervosa e formação de sinapses (Ding et al, 2016). Diversos agentes como fatores de crescimento, de transcrição e nutrientes podem afetar permanentemente o desenvolvimento neural (Welberg & Seckl, 2001). Particularmente os esteroides têm propriedades poderosas na programação neural (Matsumoto & Arai, 1997).

O fato da resposta ao estresse ocasionar liberação de grande quantidade de corticoides torna-o um candidato óbvio como fator programador no paradigma do estresse pré-natal (Welberg & Seckl, 2001).

Nos ratos o período que consiste no pico do desenvolvimento do SNC se dá entre o 7º e 14º dia de vida, perdurando até 35º (Morgane et al, 2002). Durante esse período estão ocorrendo fenômenos como neurogênese, gliogênese e intensa mielinização em diferentes regiões do cérebro (Levitisky & Barnes, 1972; Morgane et al, 2002)

Existem evidências de que a função pós-natal do eixo hipocampo-hipotálamo-pituitária-adrenal (HHPA) pode ser alterada por eventos pré-natais. Tais alterações podem ocasionar, no adulto, exposição cronicamente aumentada a glicocorticoides (GC), bem como resposta exacerbada a estímulos estressantes. Essas alterações são geralmente atribuídas a modificações na capacidade de retroalimentação hipotálamo-hipofisária de esteroides, decorrente de modulações na expressão de genes codificantes de receptores de GC (incluindo receptores glicocorticóides – GR - e mineralocorticóide – MR - no hipocampo) que levam a redução destes receptores. Paralelamente, ocorrem alterações em diversos

neurotransmissores de varias outras regiões do encéfalo (para revisão ver Welberg & Seckl, 2001).

Alterações na expressão destes receptores podem levar, em longo prazo, à disfunção na regulação da concentração plasmática de adrenocorticotrofina (ACTH) e de GC. Em fetos de porcos, foi determinada a presença de RNAm para GR por todo o cérebro, sendo a maior concentração encontrada nos núcleos paraventriculares (PVN) hipotalâmicos. Já o RNAm para MR está presente exclusivamente no sistema límbico (hipocampo, amígdala, e giro dentado) ocorrendo maior concentração no hipocampo (Lingas, et al, 1999). Estes autores demonstraram, em porcos, que a restrição nutricional materna altera a expressão destes receptores em períodos gestacionais relacionados ao maior desenvolvimento neural.

Como os sistemas de neurotransmissores e GC neurais interagem para modular tanto o comportamento quanto a atividade do eixo HHPA (McEwen, 1987) é possível que os efeitos do estresse pré-natal sejam mediados por alterações permanentes nestes sistemas (Welberg & Seckl, 2001). Devemos ainda considerar que a regulação do eixo HHPA pelo hipocampo e outras estruturas límbicas é mediada, em parte, por reles sinápticos no núcleo da estria terminal (BNST) (Cullinan et al, 1993; Feldman et al, 1990; Gray et al, 1993; Herman et al, 1994; Onaka & Yagi, 1998; Prewitt & Herman 1998; Zhu et al., 2001). O BNST está situado numa posição chave para regular não somente o comportamento de ansiedade, mas também as respostas ao estresse, implicadas em neuropatologias e distúrbios neuropsicológicos. Além disso, evidências sugerem que o BNST tem alta plasticidade podendo esta ser influenciada pelo estresse (Walker et al, 2003).

Estudos desenvolvidos em nossos laboratórios, avaliando ratos machos adultos submetidos a restrição proteica gestacional, revelaram redução da arborização dendritica de neurônios do BNST paralelamente ao aumento significativo da concentração plasmática de corticosterona e da ansiedade. Desta forma, demostramos pela primeira vez que o BNST é programado pela restrição

proteica gestacional resultando em alterações na neuroquímica e na morfologia levando a alterações comportamentais (artigo submetido).

Sabe-se ainda que o comportamento de ansiedade é mediado pela amígdala, provavelmente via hormônio liberador de corticotrofina (CRF) (Schulkin et al, 1994) cuja transcrição é facilitada pelos corticosteróides (Makino et al, 1994; Hsu et al, 1998; Hatalski et al, 1998). A prole de ratas submetidas à carboxolano (que inativa a  $11\beta$ -HSD) apresenta expressão aumentada de GR na amígdala, podendo aumentar a sensibilidade a concentrações elevadas de corticosteróides (Welberg et al, 2000) induzindo aumento do CRF.

Além disso, estudos demonstram que a exposição gestacional a glicocorticoides induz um fenótipo hiperemocional na vida adulta (Oliveira et al, 2006).

Como já está bem estabelecido o papel da amígdala e do BNST nos comportamentos de medo e ansiedade (Davis 1992, 2010) ambos relacionados à resposta ao estresse, estas estruturas constituem alvos de estudo importantes nos modelos de programação fetal.

### ***1.1 Plasticidade do BNST e da amígdala***

O núcleo da estria terminal (BNST) foi definido originalmente por Johnston, em 1923, como uma estrutura de massa cinzenta que envolve a estria terminal e se expande em suas extremidades caudal e rostral. A extremidade caudal descrita por Johnston (1923) é agora considerada como parte da amígdala. A região rostral, encontrando-se na zona ventral do septo lateral e dorsal para a área pré-óptica do hipotálamo, é denominada BNST. Johnston (1923) também observou que o BNST forma uma ligação contínua com estruturas da amígdala fato que originou o termo “amígdala estendida”.

O BNST é uma estrutura em posição privilegiada para integrar informações de estresse e regular tanto o estresse quanto os sistemas de recompensa. Tem sido demonstrado que tanto a exposição crônica a estressores como tratamentos farmacológicos (ex: corticosterona) influenciam na plasticidade do BNST (revisado em Hammack et al, 2010). Diversos estudos demonstram que a expressão de CRF no BNST foi aumentada após situação onde o indivíduo é subjugado socialmente por período prolongado (estresse social crônico), exposto a estresse crônico moderado ou ao tratamento crônico com corticosterona (Makino et al, 1994; Watts & Sanchez-Watts 1995; Schulkin et al, 1998). Adicionalmente, tem sido demonstrado que estes tratamentos aumentam os sinais de neuroplasticidade no BNST. A exposição ao modelo de estresse crônico inesperado foi associada ao aumento no volume do BNST e no comprimento dos dendritos neuronais (Pêgo et al, 2008).

A imobilização crônica aumentou o número de pontos de ramificações da arborização dendrítica de neurônios do BNST (Vyas et al, 2002; 2003) enquanto o modelo de estresse variado por uma semana aumentou o comprimento dendrítico de neurônios deste núcleo.

Têm sido observadas correlações funcionais relacionadas à neuroplasticidade do BNST já que a exposição crônica a drogas de abuso tem sido relacionada ao aumento das correntes excitatórias pós-sinápticas originadas no núcleo ventral tegmental (VTA) que se associam a neurônios do BNST, bem como aumento na expressão de transportadores de norepinefrina no BNST (Macey et al, 2003).

Elevações na concentração de CRF e da neuroplasticidade do BNST tem sido associadas ao aumento do comportamento de ansiedade e anedonia (um sintoma de depressão, Stout et al, 2000). Conseqüentemente, a exposição crônica a estressores podem alterar a neuroquímica, a morfologia e a função do BNST levando ao aumento do medo e da ansiedade. Recentemente, Oliveira et al (2012) observaram que a administração de dexametasona em ratas prenhes leva a programação do BNST e da amígdala da prole paralelamente ao fenótipo de



hiperansiedade e alteração no comportamento de medo. Eles encontraram aumento no volume e no comprimento dendrítico no BNST e redução do volume e do comprimento dendrítico na amígdala.

Assim, podemos concluir que exposição crônica a estressores leva a modulações estruturais e funcionais no BNST, já que este apresenta grande plasticidade, que medeiam desordens de ansiedade.

### **1.2 Conexões entre a amígdala e o BNST**

Alheid et al (1998) e Alheid & Heimer (1988) demonstraram que os núcleos central (CeA) e medial (CeM) da amígdala e o BNST são conectados por colunas de células localizadas por toda a estria terminal, pelas fibras do trato que conectam estes núcleos da amígdala ao BNST e também pela parte localizada ventralmente, do tronco cerebral basal. Estes autores também mostraram que o CeA emite projeções principalmente para a divisão lateral do BNST (BNSTL) e a amígdala medial (MeA) emite projeções principalmente para a divisão medial do BNST (BNSTM). Estas projeções foram denominadas como células de amígdala estendida. Além disso, o CeA e o BNSTL são muito similares anatomicamente em termos de “inputs”, “outputs”, tipos celulares, e conteúdo neuroquímico, especialmente no que diz respeito aos níveis elevados de peptídeos encontrados em ambas as estruturas (Alheid et al, 1995). O núcleo basolateral da amígdala (BLA) também emite projeções não só para o CeA, mas também para o BNSTL, especialmente partindo da região caudal do BLA (Dong et al, 2001; McDonald, 1983; Weller & Smith, 1982).

Duas áreas ricas em CRF que medeiam a resposta a estressores incluem o BNST e o CeA. Tem sido sugerido que a ativação do BNST ou do CeA coordenam a rede de respostas comportamentais apropriadas para a luta frente as ameaças percebidas e, ao mesmo tempo, engajando sistemas catabólicos periféricos que suportem estas alterações comportamentais. Conseqüentemente, tanto o CeA

quanto o BNST tem sido implicados na mediação da resposta ao estresse bem como tem sido considerados críticos na modulação de comportamentos afetivos (medo e ansiedade) (Walker et al, 2003, 2009).

### ***1.3 Mediadores do medo e da ansiedade***

É amplamente conhecido que as catecolaminas desempenham papel importante no sistema neuroquímico do cérebro, e estão envolvidas em uma série de funções cerebrais, dentre elas a resposta ao medo e a ansiedade, bem como distúrbios neuropsiquiátricos como esquizofrenia, dependência de drogas, transtorno de déficit de atenção e hiperatividade (TDAH) e doença de Parkinson (Genro et al. 2010 ; Howes & Kapur 2009 ; Melis et al. 2005 ; Oades et al. 2005 ; Piazza & Le Moal 1996). Além disso, neurotransmissores clássicos como dopamina e noradrenalina são responsáveis por controlar uma variedade de funções, incluindo a locomoção, função autonômica, secreção hormonal e os comportamentos complexos que estão associados com afeto, emoção e recompensa (Torres et al, 2003).

Em resposta ao estresse, o CRF regula a atividade do eixo HHPA através da ativação do receptor CRF1 (Davis, 1992; Merali et al., 2004; Muller , 2003). Adicionalmente à liberação de CRF no sangue, os neurônios do PVN apresentam projeções para outros locais do sistema nervoso central como BNST, CeA e VTA, resultando em uma variedade de respostas nestas regiões cerebrais (para revisão ver Corominas et al., 2010).

Além de regular a atividade do eixo HHPA, em resposta a estressores, a liberação de CRF desencadeia mudanças em outros sistemas de neurotransmissores, tais como a serotonina (5-HT) (Millan, 2005; Holsboer, 2003; Nestler et al., 2002; Leonard, 2005; Holmes et al, 2003).

Um substancial corpo de evidências na literatura tem implicado o sistema 5-HT na modulação do comportamento de medo e ansiedade (Graeff et al, 1996;

Handley et al 1993; Handley, 1995; Lowry et al, 2005; 2008). Os receptores de 5-HT têm sido classificados em sete famílias distintas (5-HT 1-7) que medeiam ações excitatórias e inibitórias quando ativados por 5-HT. Hammack et al, 2010 injetaram antagonistas seletivos do receptor 5-HT<sub>1A</sub> no BNST e observaram aumento dose-dependente no comportamento de ansiedade, isto é, um comportamento ansiolítico. Estes mesmos autores verificaram que a ativação de 5-HT<sub>2A</sub>, 5-HT<sub>2c</sub> e ou 5-HT<sub>7</sub> é ansiogênica.

O sistema 5-HT é suscetível ao estresse e ao cortisol. O estresse agudo causa liberação de 5-HT pelas células do núcleo da rafe, mas o estresse em longo prazo pode depletar estes estoques (Fontenot et al., 1995). Esta depleção pode ser permanente: em macacos que foram estressados por 14 meses e mantidos em recuperação por mais 14 meses as concentrações de 5-HT no córtex pré-frontal ventral nunca retornou aos valores observados antes do estresse (Fontenot et al., 1995). O estresse pode afetar as células da rafe através dos receptores de glicocorticóides, os quais estão presentes em células serotoninérgicas (Laaris et al., 1995) e afetam o tipo e a quantidade de proteínas produzidas por estas células. No BNST GR (Kream et al, 1983) e MR (para revisão ver Gomez-Sanchez, 2011) também estão implicados no comportamento ansiogênico e ansiolítico, respectivamente.

Além disso, o fator neurotrófico derivado do cérebro (BDNF), uma neurotrofina extremamente responsiva ao estresse, a qual exerce papel central no desenvolvimento, na plasticidade, na fisiologia e nas doenças do sistema nervoso podendo estar implicado nos comportamentos de medo e ansiedade (Hammack et al, 2010).

## 2. JUSTIFICATIVA E OBJETIVOS

Tendo em vista a fundamentação apresentada acima o desenvolvimento do presente projeto se **JUSTIFICA** diante dos seguintes fatores:

- A prevalente desnutrição materno-infantil, com evidentes repercussões sobre a saúde e a necessidade de desenvolvimento de políticas de saúde pública que minimizem seus efeitos;
- A extrema sensibilidade do SNC à programação pré-natal;
- A programação fetal como fator de risco para o desenvolvimento de desordens psicológicas;
- A comprovada implicação da programação fetal em alteração do eixo HHPA podendo ocasionar, no adulto, exposição aumentada cronicamente a GC e/ou exacerbação na resposta ao estresse;
- O aumento da exposição fetal a corticosteroides em modelos de programação fetal;
- A já estabelecida ação moduladora neural dos corticosteroides podendo ocasionar alterações estruturais tanto no volume quanto na estrutura fina (por exemplo: número de sinapses e morfologia dendrítica) de neurônios;
- O BNST e a Amígdala medeiam respostas a estressores e o comportamento de ansiedade e medo estando implicados em distúrbios psicológicos;
- A exposição materna a restrição proteica gestacional leva, na idade adulta, a importantes mudanças neuroquímicas e morfológicas no BNST tornando estes animais mais ansiosos.
- A inexistência de trabalhos avaliando no início da vida a amígdala e o BNST neste modelo;

A ocorrência de desenvolvimento neural intenso entre o 7º e 14º dia de vida em ratos;

Foram **OBJETIVOS** do presente projeto estudar em ratos machos submetidos à restrição proteica *in útero*, comparativamente aos seus controles, no 7º e 14º dia de vida, a concentração de corticosterona basal e na amígdala e no BNST:

- O volume e número de neurônios e outros tipos celulares.
- A estrutura dendrítica de neurônios;

- A concentração de MR, GR, CRF, CRF1, receptores de 5-OH-Triptamina (5HT1A e 5HT2A) e BDNF;
- A concentração de catecolaminas e serotonina;

### 3. MATERIAL E MÉTODOS

A primeira parte do estudo foi realizada na Universidade do Minho, Portugal durante o estágio de doutorado sanduiche.

#### 3.1 Primeira parte

Os experimentos foram realizados nos laboratórios do Instituto de Ciências da Vida (ICVS), na Escola de Ciências da Saúde da Universidade do Minho, Braga, Portugal. Todo o estudo foi aprovado pela Direção Geral de Alimentação e Veterinária (DGAV, número 023432/2013-08-30).

Ratos Wistar Hannover machos e fêmeas foram obtidos do *Charles Rivers Laboratories* (Barcelona, Espanha) com nove semanas de vida. Após período de quarentena foram realocados no biotério do ICVS.

Os animais permaneceram com água e ração padrão para roedores *ad libitum*. Os mesmos foram mantidos no biotério com temperatura e umidade controlada, com sistema de luz 12h/12h.

Após o período de adaptação às condições ambientais de biotério, os animais foram submetidos ao acasalamento em sistema de harém durante todo o período noturno a partir da décima segunda semana de vida e, após constatação da presença de espermatozoides no lavado vaginal foram consideradas prenhas.

Então, a partir deste momento foram separadas individualmente de forma aleatoria em dois grupos experimentais. Um grupo passou a ser alimentado com ração normoproteica contendo 17% de caseína (n=20) sendo denominado grupo NP (*normal protein*). O outro grupo recebeu ração hipoproteica com 6% de

caseína (n=20) denominado grupo LP (*low protein*). Ambos os grupos receberam água e ração *ad libitum*. A ração (Tabela 1) foi produzida pela Ultragene (Equipamento para Experimentação e Investigação Científica Laboratorial de Portugal).

**Tabela 1:** Formulação da ração utilizada (g/kg).

INGREDIENTES	NORMOPROTEICA	HIPOPROTEICA
	17% (NP)	6% (LP)
Amido de milho	410,10g	484,80g
Caseína	188,90g	66,70g
Amido dextrinizado	130,50g	159,00g
Sacarose	100,00g	121,00g
Óleo de soja	70,00g	70,00g
Celulose microcristalina	50,00g	50,00g
Mix mineral AIN 93	35,00g	35,00g
Mix vit AIN 93	10,00g	10,00g
L cistina	3,0g	3,0g
Bitartarato de colina	2,5g	2,5g
BHT	0,014g	0,014g

Semanalmente foi verificado o peso dos animais bem como seu consumo alimentar.

Ao nascimento as dietas foram retiradas e retornou-se a dieta padrão para roedores e água *ad libitum*.

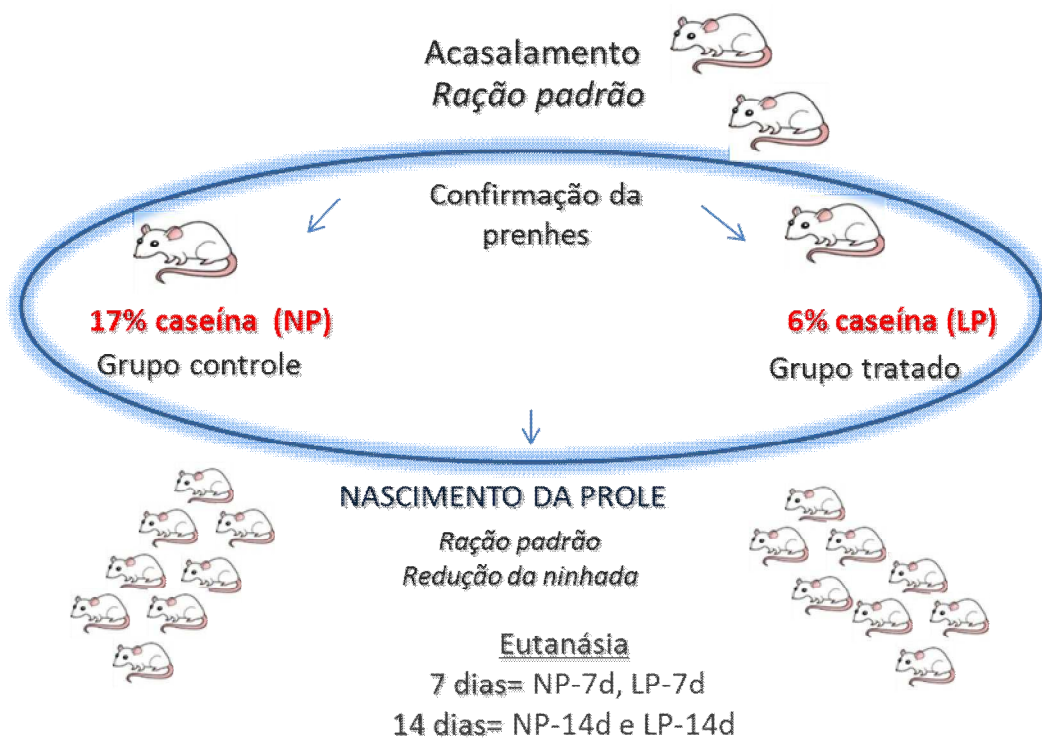
Neste momento também foi mensurada a distância ano-genital (DAG) para verificar o sexo dos filhotes. Os animais com DAG inferior a 2 mm foram

consideradas fêmeas e aqueles com distancia superior a 2 mm machos. O peso ao nascer de cada filhote também foi verificado.

A ninhada foi reduzida a, no máximo, 8 filhotes por progenitora a fim de que todos os animais tivessem a mesma oferta alimentar e cuidado materno.

No sétimo e décimo quarto dia de vida pós-natal os grupos foram divididos da seguinte forma:

- ✓ *Normal Protein* sete dias (NP7D)
- ✓ *Normal Protein* quatorze dias (NP14D)
- ✓ *Low Protein* sete dias (LP7D)
- ✓ *Low protein* quatorze dias (LP14D)



**Figura 1.** Desenho experimental

### **3.1.1 Reconstrução neuronal 3D**

No 7<sup>o</sup> e 14<sup>o</sup> dia pós-natal, os animais (n= 4 NP e 4 LP de 4 diferentes progenitoras) foram perfundidos com solução salina 0,9%, sob anestesia profunda, com ketamina (75mg/kg) e xilasina (10mg/kg), e processada de acordo com o protocolo descrito por Gibb & Kolb (1998). Os cérebros foram removidos e imersos em solução de Golgi-Cox (solução 1:1 de dicromato de potássio 5% e cloreto de mercúrio 5% diluído 4: 10 com cromato de potássio 5% - Glaser & Van Der Loos, 1981) durante 14 dias; os cérebros foram então transferidos para uma solução de sacarose 30% (mínimo 3 dias), antes de serem cortados em um vibratomo. Secções coronais (200 µm de espessura) foram coletadas em sacarose 6% e secas em lâminas. Posteriormente alcalinizados em amônia 18,7%, em Dektol (Kodak), fixados em Kodak Rapid Fix (preparada conforme instruções da embalagem com omissão da solução B), desidratados em serie crescente de álcool, diafanizadas em xilol e montadas.

Para cada neurônio selecionado, todos os ramos da árvore dendrítica foram reconstruídos em ampliação de 600x usando um microscópio motorizado (Axioplan 2, Carl Zeiss, Alemanha), com objetivas de imersão, e ligado a uma câmara (DXC-390, Sony Corporation, Tóquio, Japão) e *software* Neurolucida (Microbrightfield, VT). A análise 3D dos neurônios reconstruídos foi realizada utilizando *software* Neurolucida (Microbrightfield). Foram comparadas as mudanças globais, o comprimento total das árvores e número de ramificações dendríticas entre os grupos, (Uylings & Van Pelt 2002). Para avaliar as diferenças na organização dendrítica, uma versão em 3D da análise de Sholl (Sholl 1956; Uylings & Van Pelt 2002) foi realizada. Para isso, contamos o número de intersecções com os dendritos de esferas concêntricas posicionados em intervalos radiais de 10 µm para o BNST e 20 µm para Amígdala, também foi avaliado o comprimento da árvore dendrítica localizada entre duas esferas consecutivas.

Neurônios completos do BNST e da Amígdala foram selecionados para análise de sholl.



### **3.1.2 Estereologia**

Para estimativa do volume e número total de células utilizamos a estereologia. Então, no 7<sup>o</sup> e 14<sup>o</sup> dia pós-natal, os animais (n= 4 NP e 4 LP de 4 diferentes progenitoras) foram anestesiados com ketamina (75mg/kg) e xilasina (10mg/kg), e então perfundidos com solução salina 0,9% e paraformolaldeído (PFA) a 4%. Os cérebros foram removidos e imersos em PFA por mais 2 semanas e desidratados em uma bateria com concentração crescente de álcool, permanecendo em álcool 100% por 4 dias, sendo este trocado 2 vezes por dia. Após este período o material foi incluído em Tecnovit 7100 (Heraeus Kulzer, GmbH). Secções de 30µm foram obtidas utilizando navalha de tungstênio, coletando apenas os cortes pares que foram corados com Giensa 20%. As estimativas de volume e número de células foram obtidas utilizando *software* Estereo Investigator ® (MicroBrightField, Williston, VT, EUA) e um microscópio motorizado (Axioplan 2, Carl Zeiss, Hamburgo, Alemanha) acoplado a uma câmara (DXC- 390, Sony Corporation, Tóquio, Japão). O princípio de Cavalieri foi usado para avaliar o volume de cada região. O número médio de células foi estimado utilizando método óptico. Coeficientes de erro foram calculados de acordo com as fórmulas publicadas anteriormente para números celulares e para volume estimado (Oliveira et al, 2012).

Foram analisados o volume e número médio de células do BNST anterodorsal e anteroventral.

### **3.1.3 Western Blotting**

A expressão proteica foi avaliada pela técnica de *western blotting* (WB). Animais (n= 10 NP e 10 LP de 5 diferentes progenitoras) com 7 e 14 dias de vida pós-natal foram decapitados e seus crânios foram rapidamente congelados em nitrogênio líquido a fim de evitar degradação durante a macrodissecção. O BNST

e Amígdala foram cuidadosamente dissecados em uma placa de gelo e armazenados em *eppendorfs* em freezer  $-80^{\circ}\text{C}$ .

*Extração de proteínas:* O tecido foi homogeneizado em tampão de extração RIPA (Radio Immuno Precipitation Assay Buffer) com seringa 23G ou homogeneizador e, em seguida, adicionamos 10% de triton x 100 e 10% de SDS e incubamos em gelo por 1h.

As amostras foram centrifugadas por 10 minutos a 13.000 rpm (rotações por minuto) a  $4^{\circ}\text{C}$  e foi coletado o sobrenadante.

*Quantificação de proteínas:* As proteínas totais foram quantificadas utilizando-se o método de Bradford e a leitura foi feita a 595nm.

As amostras foram diluídas em tampão de Laemmli, aquecido a  $95^{\circ}\text{C}$  por 5 minutos e a quantidade correspondente de proteína foi aplicada em gel SDS-PAGE e colocado em aparelho de eletroforese Bio-Rad (Mini-Protean, Bio-Rad). A eletroforese das proteínas no gel foi feita a 120V. Depois da separação eletroforética as proteínas foram transferidas para membrana de nitrocelulose e incubadas com anticorpo primário a  $4^{\circ}\text{C}$  durante a noite.

Posteriormente, as membranas foram lavadas com solução basal e incubadas com anticorpo secundário específico por 2 horas em temperatura ambiente. As bandas imunoreativas foram detectadas utilizando-se solução quimioluminescente. As imagens foram obtidas em fotodocumetador ChemiDoc XRS system (Biorad; 170870) e a intensidade das bandas quantificada por densidade ótica no *software* TINA. Os valores da densidade obtidos foram utilizados para análise estatística.

### *ANTICORPOS UTILIZADOS:*

- ✓ GR (H-300- Santa Cruz 8992) anti-rabbit → 1:300
- ✓ MR (H-300- Santa Cruz) anti-rabbit → 1:300
- ✓ BDNF (Abcam; ab46176) anti-rabbit → 1:4.000
- ✓ 5HT1A (ab101914) anti-goat → 1:1.000
- ✓ 5HT2A (ab160228) anti-rabbit → 1:1.000
- ✓ CRF (S-19- Santa Cruz 1761) anti-goat → 1:500
- ✓ CRF1 (ab 59023) anti-goat → 1:500
- ✓ Alfa tubulina (DSHB; AA4.3) anti-mouse → 1:200
- ✓ Anti-goat (Santa Cruz biotechnology; sc-2020) → 1:7.500
- ✓ Anti-rabbit (BioRad; 170-6515) → 1:10.000
- ✓ Anti-mouse (BioRad 1721011) → 1:10.000

#### ***3.1.4 Dosagem de neurotransmissores por HPLC***

A concentração de catecolaminas e serotonina foram analisadas por cromatografia líquida de alto desempenho, combinada com detecção eletroquímica (HPLC / CE) usando um instrumento Gilson (Gilson, Middleton, WI, EUA), equipado com uma coluna analítica (Supelco 'Supelcosil LC-18, 3 mM, Bellefonte, PA, EUA; taxa de fluxo: 1,0 ml • min<sup>-1</sup>) (n= 6 NP e 6 LP de 5 diferentes progenitoras).

#### ✓ ***PREPARO DAS AMOSTRAS***

Foi adicionado 200ul de ácido perclórico ( 0,2) em cada amostra e, após 30 minutos de repouso em gelo, foram sonicadas e centrifugadas por 10 minutos a 13.000rpm a 4°C.

O sobrenadante foi recolhido para um eppendorf com filtro e novamente centrifugado por 8 minutos a 10.000rpm a 4°C e o pellet armazenado.

O sobrenadante resultante foi filtrado através de uma coluna de HPLC Spin-X (Costar, Lowell, MA, EUA) para remover os detritos e alíquotas de 150µl foram injetadas no sistema de HPLC, usando uma fase móvel de fosfato de potássio 0,7 M aquoso (pH 3,0) em 10% de metanol, ácido 1-heptano (222mg/l) e Na-EDTA (40mg/l). Uma curva padrão utilizando concentrações conhecidas de todas as catecolaminas foi corrida a cada dia.

A dosagem de proteína total foi feita a partir do pellet armazenado utilizando o método de Bradford a fim de normalizar a leitura realizada em HPLC.

**Tabela 2:** Padrões utilizados

NEUROTRANSMISSORES	REFERENCIAS
Serotonina	Sigma H-7752 1g 4°C
5-HIAA	Sigma H-8876 500MG -20°C
Dopamina	Sigma H-8502 5G 25°C
DOPAC	Sigma D-9128 1G 25°C
HVA	Sigma H-1252 250MG 25°C
Epinefrina	Sigma E-4375 1g 4°C
Norepinefrina	Sigma 74460 25°C

### **3.2 Segunda parte.**

*Realizada na Universidade Estadual de Campinas (Unicamp)*

Todo o estudo foi feito de acordo com os princípios éticos na Experimentação animal adotado pelo Colégio brasileiro de Experimentação Animal (COBEA) e foi aprovado pela Comissão de Ética na Experimentação Animal (CEEA) protocolo 3908-1 da Universidade Estadual de Campinas. Os estudos foram realizados no Laboratório de Metabolismo Hidrossalino no Núcleo de Medicina e Cirurgia Experimental da Unicamp.

Os animais foram obtidos no Centro Multidisciplinar para Investigações Biológicas da Unicamp (CEMIB – Unicamp). Foi utilizando o mesmo desenho experimental descrito anteriormente.

### **3.2.1 Corticosterona basal**

Amostras de sangue foram coletadas de animais de 7 e 14 dias de vida (n= 14 NP e 15 LP animais de 5 progenitoras diferentes), no momento da eutanásia por decapitação, entre 8 e 9h. Após a coleta o sangue foi centrifugado 10 minutos a 13.000 rpm, o sobrenadante foi retirado e rapidamente estocado em freezer - 80°C.

A dosagem foi realizada pelo método de radioimunoensaio (RIA) usando um kit comercial de corticosterona da R&D Systems™ a biotechne brand seguindo instruções do fabricante:

#### ✓ *PRÉ-TRATAMENTO DA AMOSTRA*

O pré-tratamento remove potenciais interferências de proteínas ligadas à proteína e de corticosterona ligada à proteína. As amostras foram analisadas dentro de 8h após o pré-tratamento.

Foi então adicionado 150µl de amostra e 150µl de pré-tratamento, agitado cuidadosamente e deixado em repouso por 15 minutos em temperatura ambiente. Após este período o material foi centrifugado por 4 minutos em centrifuga

previamente resfriada a 4°C a 12.000 rpm e coletado o sobrenadante. O fator de diluição da curva padrão foi 2.

#### ✓ *DOSAGEM EM MICROPLACA*

Em seguida ao pré-tratamento foi adicionado 50µl de anticorpo 1º anti corticosterona à microplaca excluindo o controle negativo. A placa foi coberta com película adesiva e incubada por 1h em temperatura ambiente sob agitação.

A seguir, após 4 lavagens consecutivas dos pocinhos foi adicionado 100µl de tratamento F, 50µl de amostra ou amostra padrão em duplicata e 50µl do conjugado de corticosterona em todos os pocinhos. A placa foi novamente coberta com película adesiva e incubada em temperatura ambiente por 2h sob agitação. Após mais 4 lavagens foi adicionado 200µl solução substrato a todos os pocinho e a placa foi incubada por 30 minutos em temperatura ambiente na bancada protegida da luz.

Adicionamos 100µl de solução *stop* em todos os pocinhos e determinamos a densidade ótica a 450nm e a 540nm.

#### ✓ *ANALISE*

O Valor da leitura em 540nm foi descontado da leitura em 450nm para corrigir imperfeições ópticas da placa. A seguir foi feita a média de todas as medidas e descontado o valor do branco. A partir das leituras das amostras padrão foi construída a curva padrão e equação da curva que foi utilizada para calcular a concentração de corticosterona corrigida pelo fator de diluição.

A dose mínima detectada pelo kit foi de 0,028ng/ml.

### 3.2.2 Fracionamento isotrópico

Para estimar o número total de células e de neurônios aplicamos a técnica de fracionamento isotrópico (Herculano-Housel & Lent, 2005).

Os animais foram anestesiados no 7<sup>o</sup> ou 14<sup>o</sup> dia de vida (n= 7 NP e 7 LP animais de 3 progenitores diferentes) com ketamina (75mg/kg) e xilasina (10mg/kg) e perfundidos com solução salina 0,9% e paraformaldeído (PFA) a 4%. Depois os cérebros foram removidos e o BNST e a amígdala foram dissecados, pesados e imersos em PFA por 48 horas, em agitação e desidratados em uma bateria crescente de álcool, permanecendo em álcool 100%.

Cada amostra foi homogeneizada em um vidro homogeneizador com 3ml de solução de dissociação (40 Mm de citato de sódio e 1% de Triton X-100) e transferidos com pipetas de vidro para tubos de centrifugação graduados.

Após esta etapa homogeneizou-se manualmente 20 vezes a solução por tombamento, imediatamente uma alíquota de 1ml foi retirada e centrifugada durante 5' a 6000 rpm. O sobrenadante foi descartado e o pellet foi suspenso para 1ml com PBS sendo que esta lavagem (processo de centrifugação e re-suspensão do pellet) foi realizada 3x.

Após a última lavagem o pellet foi suspenso e incubado a temperatura ambiente por 24h com anti-NeuN igG (1:200 EM pbs; Chemicon, Temeluca, CA). No dia seguinte a alíquota foi lavada 3x e os núcleos foram incubados com anticorpo secundário, Goat Anti-Mouse conjugado a CY3 (1:200 em 40% PBS, 10g (normal Goat Serum e 50% DAPI) por 2h, sendo então lavados e suspensos em PBS para contagem dos núcleos de neurônios em microscópio de fluorescência.

Após homogeneização mecânica uma alíquota de 10µl foi pipetada na câmara de Neubauer, aguardou-se por 5' e então fez-se a leitura, contando-se primeiramente os núcleos marcados com DAPI. Em realizou-se a contagem dos núcleos de neurônios NeuN+ pelo Cy3 .

Foi contado o número de células contidos em pelo menos 40 $\mu$ l e multiplicado pelo volume total da solução.

### **3.3 Análise Estatística**

Quando foram avaliadas duas variáveis com dispersão paramétrica utilizando o teste t de Student. Quando avaliadas mais de duas variáveis com distribuição paramétrica foi realizada a Análise de Variância (ANOVA) com *post-hoc* de Bonferroni. Quando a distribuição dos dados não assumiu uma curva normal foi utilizado o teste estatístico de Kruskal-Wallis. O nível crítico definido foi de 5% ( $p < 0,05$ ). Os resultados estão expressos em Média  $\pm$  Desvio Padrão da Média. *Outliers* foram identificados e eliminados da análise quando extrapolaram para mais ou para menos 2x o Desvio Padrão em relação à Média. O software para análise dos dados e confecção dos gráficos foi o GraphPad Prism v5.00



## 4. RESULTADOS

### 4.1 Artigo 1

#### EARLY MORPHOLOGICAL AND NEUROCHEMICAL CHANGES OF THE BED NUCLEUS OF STRIA TERMINALIS (BNST) IN GESTATIONAL PROTEIN-RESTRICTED OFFSPRING

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#### **Abstract**

The bed nucleus of the stria terminalis (BNST) is a structure involved in the stress-response and it is associated with the modulation of anxiety-like behavior and fear. Due to its plasticity, its behaviors can be affected by prenatal events as protein

restriction *in utero*, where the fetus is exposed to high maternal glucocorticoids levels, leading to a deregulation of HPA axis and exacerbated stress answer at adult life. Through severe maternal low protein diet with 6% casein we analyzed males offspring with 7 and 14 days post natal and found low birth weight until the 14<sup>th</sup> day. It was not significant effect on levels of basal corticosterone, however we found expressive changes on BNST morphology with reduction of length of dendrites, of volume of dorsal anterior region even as decreased at total numbers of cells, glial cells and BNST neurons. These changes may be involved with the expressed depletion of BDNF receptors that is essential for plasticity and neural survival. Beyond that, the gestational protein restriction led to reduction on GR and MR receptors expression. Considering that MR is involved in the basal activity maintenance, it's may be an indicative of HPA axis deregulation. Besides that, this axis has strong influence on the catecholaminergic and serotonergic activity, which was affected by the significant rise of norepinephrine and 5HIAA levels and reduction of dopamine turnover, showing that gestational protein restriction exerts a stressor effect on BNST trough the activation of these systems. Our results reveal that gestational protein restriction alters BNST morphology and neurochemistry in the beginning of postnatal life and may be involved with the anxiety processes in the adult life.

**Keywords:** BNST, Fetal programming, glucocorticoids, CRF, BDNF, Catecholamine.

### ***Introduction***

Maternal malnutrition is even nowadays one major public health issue worldwide (WHO 2013). Inadequate levels of nutrients during pregnancy can lead to permanent alterations in fetal tissues and to an increased risk of developing metabolic and endocrine dysfunctions and cardiovascular diseases in adulthood (Ashton, 2000). Researchers have shown that both nutritional and psychological stress during pregnancy can lead to a low weight at birth and are risk factors for the development of psychiatry disorders (Burd et al, 1999).

One of the mechanisms involved in the development of these disorders is the elevated exposure of the fetus to maternal glucocorticoids (GC) due to a low concentration and decreased activity of the placental enzyme 11 $\beta$ -hydroxysteroid dehydrogenase (11 $\beta$ -HSD) type 2 (Benediktsson et al, 1993; Stewart et al, 1995; Langley-Evans et al, 1996; Langley-Evans, 1997). These prenatal alterations may lead to a chronic increase in GC as well as exacerbated response to stressful stimuli in adulthood (for review see Welberg & Seckl, 2001). There is a wide range of evidence that shows that the postnatal activity of the hypothalamic-pituitary-adrenal (HPA) axis, the main mechanism of stress-response in the brain, can be altered by prenatal events (Barker, 1995).

In particular, the brain is extremely sensitive to fetal programming and the intra-uterus exposure to GC can deeply influence the structure of the nervous system and synaptic formation (Ding et al, 2016). Among the different brain areas involved in the stress-response the bed nucleus of the stria terminalis (BNST) seems particularly sensitive to stress and early-life exposure to glucocorticoids (Oliveira et al, 2012) showing elevated plasticity (Walker et al, 2003; Pêgo et al, 2008). The BNST is positioned in a privileged location for the regulation of the stress-response and, in particular, the BNST has been strongly associated with the modulation of anxiety-like behavior. The sensitivity of the BNST to stress can be, at least in part, explained by the presence of corticosteroid receptors in this brain region. In fact, GC gluco- (GR) (Kream et al, 1983) and mineralocorticoid receptors (MR) (for review Gomez-Sanchez, 2011) within the BNST have been associated respectively with in anxiogenic and anxiolytic roles.

The BNST is rich in corticotrophin releasing factor (CRF) neurons. CRF is a neuropeptide that has long been associated with the stress-response and with anxiety-behavior. CRF can act through two different receptors: CRF<sub>1</sub> and CRF<sub>2</sub>. While the role of CRF<sub>2</sub> is still not completely understood the activation CRF<sub>1</sub> has been strongly associated with an increase in anxiety (Davis, 1992; Merali et al, 2004; Muller, 2003). In the brain, in response to stress CRF is also released by neurons in the paraventricular nucleus of the hypothalamus (PVN) leading to the activation of the HPA axis. Considering the elevated connectivity of both BNST and

PVN neurons with projections to other areas in the central nervous system (CNS) such as the central nucleus of the amygdala (CeA), hippocampus, prefrontal cortex and ventral tegmental area (VTA), the release of CRF in these areas can influence the activity of many others regions (for review see Corominas et al, 2010).

As mentioned before in response to stressful stimuli CRF is able to regulate the activity of the HPA axis and this will also influence other neurotransmitter systems such as serotonin (5-HT) (Millan, 2005; Holsboer, 2003; Nestler et al, 2002; Leonard, 2005; Holmes et al, 2003). The serotonergic system is highly sensitive to both stress and corticosteroids; acute stress leads to the release of 5-HT by cells in the raphe nucleus but chronic stress can lead to the depletion of 5-HT in this area (Fontenot et al, 2005). Many other factors are also important for the stress response and among those the brain derived neurotropic factor (BDNF) seems to be extremely sensitive to stress. BDNF is a neurotropic factor that has a key role for physiology and pathology development in the CNS and for brain plasticity mechanisms and has been implicated in both fear and anxiety-behaviors (Hammack et al, 2009).

We found an impoverished dendritic arborization in BNST neurons, in parallel-enhanced anxiety-like behavior and elevated plasmatic corticosterone levels, in adult rats submitted to gestational protein restriction (submitted article). Thus, BNST is “programmed” by gestational protein restriction, resulting in fine structural changes and neurochemical adaptations of these brain regions that constitute potential underlying causes of the altered behavioral states.

In the current study, we aim to analyze the early effects of gestational protein restriction on BNST morphology and neurochemistry.

## ***Materials and methods***

***Animals and treatments*** - The experiments were conducted on age-matched female offspring of sibling-mated Wistar Hannover rats (250-300g). The experiments were done in accordance with the general guidelines established by the Brazilian College of Animal Experimentation (COBEA) and approved by the

Institutional Ethics Committee (CEE/UNICAMP #3908-1) and National Institutes of Health guidelines on animal care and experimentation and approved by Director General Veterinary (DGV; the Portuguese National Institute of Veterinary 023-432/08.30.2013). A part of our site colonies originated from the breeding stock supplied by Charles River Laboratories, Barcelona, Spain. Another part was originated from a breeding stock supplied by CEMIB/Unicamp, Campinas, SP, Brazil. The animals were housed in pairs under standard laboratory conditions (lights on from 8 a.m. to 8 p.m.) and had access to food and water *ad libitum* and followed up to 12 weeks of age. The rats were placed to mate and the day that sperm were seen in the vaginal smear was designated as day 1 of pregnancy. The dams were divided in two groups: one maintained on isocaloric standard rodent laboratory chow with normal protein content [NP] (17% protein) and other received a diet with low protein content [LP] (6% protein) *ad libitum* throughout the entire pregnancy. Food consumption was determined every day (subsequently normalized for body weight). All groups returned to the NP chow intake after delivery. On the day of birth the male pups were weighted and they were kept only 8 pups per female.

***Radioimmunoassay (RIA) method*** - Basal corticosterone was determined on blood samples that were collected from animals at postnatal day 7 and 14, between 8 and 9 am. After collection the blood was centrifuged (10-min at 13000 rpm) and the supernatant was quickly removed and stored in a freezer at -80°C. The dosage was performed by RIA using a commercial kit of R&D corticosterone Systems™ Biotechne the brand following the manufacturer's instructions.

***Morphological analyses methods*** - At 7<sup>th</sup> and 14<sup>th</sup> postnatal day male rats from NP (n=27/per age from 10 different mothers) and LP (n=27/per age from 10 different mothers) groups were deeply anesthetized with a mixture of ketamine and xylazine (75 and 10mg/kg respectively, i.p.) and monitoring the corneal reflex controlled the level of anesthesia. For stereology and isotropic fractioning the rats were transcidentally perfused with saline containing heparin (5%, for 15 min, under

constant pressure) followed by fixative solution: 0.1M phosphates buffer (PB; pH 7.4) containing 4% (w/v) paraformaldehyde. For 3D reconstruction we used only saline for perfusion.

**Stereology** - After perfusion the brains were removed and placed on the same fixative solution for 2 weeks, dehydrated and included in Tecnovit 7100 (Heraeus Kulzer. Gmbh). Coronal sections (30 $\mu$ m thick) were collected and stained with Giemsa 20%. The anterodorsal and anteroventral BNST volume and cell number were determined using the software Stereo Investigator (MicroBrightField. Williston VT USA) and a motorized microscope (Axioplan 2 Carl Zeiss Hamburg Germany) attached to a camera (DXC- 390. Sony Corporation. Tokyo. Japan). Cavalier's principle was used to evaluate the volume of each region. Average cell number was estimated using the optical fractionator method. Coefficients of error were calculated based on previously published formulas for cell number and for volume estimates (Oliveira et al, 2012).

**Isotropic fractionator method** - For total cells and neuron quantification we used the technique described by Herculano-Houzel & Lent (2005). The brains of 14-day-old rats (3 NP and 3LP, from different mothers) were removed and BNST was dissected. A suspension of nuclei was obtained through mechanical dissociation in a standard solution (40mM sodium citrate and 1% Triton X-100), using a 40-ml glass Tenbroeck tissue homogenizer. After washed several times with dissociation solution and centrifuged (10-min at 4000 g) pelleted nuclei were suspended in phosphate-buffered saline (PBS) containing 1% 4', 6-diamidino-2-phenylindole (DAPI, Molecular Probes, Eugene, OR, USA). DAPI-stained nuclei were counted under a fluorescence microscope at 400 $\times$ objective. The nuclear density in the suspension was determined by averaging over at least eight samples the total number of cells in the original tissue was estimated by multiplying mean nuclear density by total suspension volume. Neuron number was estimated after immunohistochemistry using anti-NeuN antibody (1:300 in PBS; Chemicon Temecula, CA, USA) and CY3 goat anti-mouse secondary antibody (1:400 in 40%

PBS, 10% goat serum and 50% DAPI; Accurate Chemicals, Westbury, NY, USA) under the fluorescence microscope (Scabora et al., 2013). Total number of other BNST cells nuclei is calculated by subtracting the number of NeuN containing nuclei from the total number of nuclei.

***Neuronal 3D reconstruction and dendritic tree analysis*** - Brain from 14 days old animals (4 NP and 4 LP from different mothers) were Golgi–Cox staining according to a published protocol (Gibb R & Kolb B, 1998). Briefly brains were removed and immersed in Golgi–Cox solution (Glaser & Van der Loos, 1981) for 14 days; brains were then transferred to a 30% sucrose solution (3-days) before being cut on a vibratome. Coronal sections (200  $\mu\text{m}$  thick) were collected in 6% sucrose and blotted dry onto gelatin-coated microscope slides. They were subsequently alkalized in 18.7% ammonia developed in Dektol (Kodak), fixed in Kodak Rapid Fix, dehydrated and cleared in xylene before being mounted and coverslipped. Slides were coded before morphometric analysis in both sets. For dendritic tree analyze the criteria used to select neurons for reconstruction were as follows: (i) full impregnation of the neurons along the entire length of the dendritic tree; (ii) dendrites without significant truncation of branches; (iii) relative isolation from neighboring impregnated neurons to avoid interference with the analysis; and (iv) no morphological changes attributable to incomplete dendritic impregnation of Golgi–Cox stain (Pêgo et al. 2008). Accordingly we chose neurons with bipolar conformation confined to the anteromedial area (BNSTam) for dendritic analysis using the following criteria: (i) presence of transverse anterior commissure; (ii) rostral location to the stria terminalis main bundle; and (iii) selection of neurons adjacent to the anterior commissure. These landmarks correspond to the rostral portion of the medial division described by McDonald (1983). For each selected neuron all branches of the dendritic tree were reconstructed at 600x magnification using a motorized microscope (Carl Zeiss Axioplan 2) attached to a camera (DXC-390; Sony Co, Japan) and Neurolucida software (Micro Bright Field, VT, USA). Three-dimensional analysis of the reconstructed neurons was performed using

NeuroExplorer software (MicroBrightField). For the dendritic analysis were reconstructed 91 neurons.

**Neurochemical analyses methods** - Male rats at 7 and 14 days were decapitated and the heads were snap-frozen in liquid nitrogen. The BNST was dissected and stored at -80°C until further analysis.

**Western blot (WB)** - The tissue was homogenized in extraction buffer RIPA (Radio Immune Precipitation Assay Buffer) and then 10% triton x 100 and 10% SDS were added to the homogenate. The tissue extracts were centrifuged (1300 rpm at 4°C for 40 min) and the supernatants used as a sample. Protein quantification was performed using the Bradford method. The samples were treated with a Laemmle buffer containing 100-mmol/l dithiothreitol (DTT) heated in a boiling water bath for 4 min and subjected to 8% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) in a Bio-Rad mini-gel apparatus (Mini-Protean Bio-Rad). Electrotransfer of proteins from the gel to the nitrocellulose membranes was performed for 90 min at 120 V (constant) in a Bio-Rad miniature transfer apparatus (Mini-Protean). The non-specific protein binding to the nitrocellulose was reduced by preincubating the filter for 2 h at 22°C in a blocking buffer (BSA 5%, 10mmol/l Tris, 150mmol/l NaCl and 0.02% Tween 20). The nitrocellulose blots were incubated at 4°C overnight with antibodies against GR (H-300-Santa Cruz 8992), MR (H-300-Santa Cruz), BDNF (Abcam; ab46176), 5HT1A (ab101914), 5HT2A (ab160228), CRF (S-19- Santa Cruz 1761), CRF1 (ab 59023) and Alfa tubulin (DSHB; AA4.3) diluted in a blocking buffer (2.5% BSA, 10mmol/l Tris, 150mmol/l NaCl and 0.02% Tween 20). Immunoreactivity bands were detected using the enhanced chemiluminescence method (RPN 2108 ECL Western blotting analysis system; Amersham Biosciences) and were detected by a ChemiDoc XRS system (Biorad; 170870). The band intensities were quantitated by optical densitometry (*TINA* software) of the developed autoradiographs that were used at exposures in the linear range.



***High performance liquid chromatography combined with electrochemical detection (HPLC/CE)***

The level of catecholamine and serotonin in the BNST was assessed by HPLC/CE using a Gilson instrument (Golson, Middleton, WI, USA) equipped with an analytical column (Supleco Supelcosil LC-18, 3mM, Bellefonte, PA, USA, flow rate: 1.0ml/min). Perchloric acid (200ul) was added to each sample that were incubated for 30 min in ice and then sonicated and centrifuged (13000rpm, 10min, 4°C). Supernatant was collected to a 1.5ml tube and then centrifuged for 8min (10000rpm, 4°C) and the resulting pellet was saved for later use. The supernatant was then filtered through an HPLC Spin-X column (Costar, Lowell, MA, USA) to remove debris and 150ul aliquots were injected into the HPLC system using a mobile phase of 0.7 M aqueous potassium phosphate (pH 3.0) in 10% methanol 1-heptanesulfonic acid (222mg l<sup>-1</sup>) and Na-EDTA (40mg l<sup>-1</sup>). A standard curve (Sigma H-7752) with known concentrations of each catecholamine was run each day for 5HIAA (Sigma H-8876), Dopamine (Sigma H-8502), DOPAC (Sigma D-9128), HVA (Sigma H-1252), Epinephrine (Sigma E-4375) and Norepinephrine (Sigma 74460).

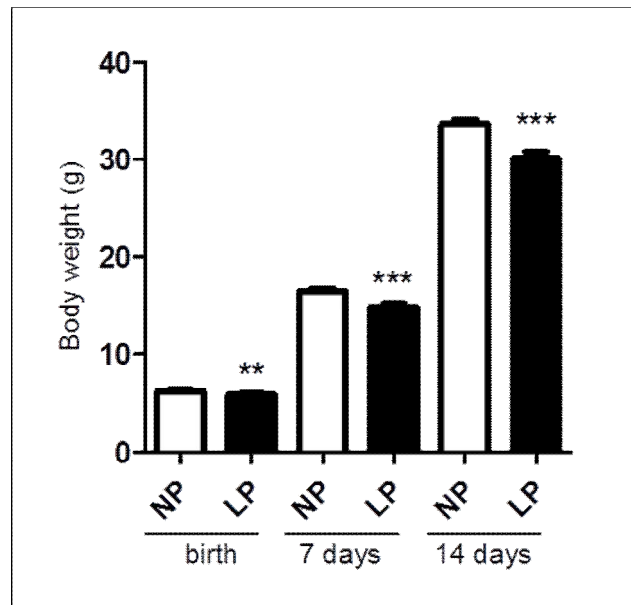
***Data presentation and statistical analysis***

All data is reported as mean ± SD. Data obtained over time was analyzed using appropriate two-way analysis of variance (two-way ANOVA). Post hoc comparisons between selected means were made by Bonferroni's contrast test when initial two-way ANOVA indicated statistical differences between experimental groups. Comparison involving only two samples of independent observations tends within or between groups was made using a Student's test. The band intensities were quantitated by optical densitometry (*software TINA*). The Tukey–Kramer test for multiple comparisons was used for analysis. The level of significance was set at  $P \leq 0.05$ .

**RESULTS**

We have not observed differences in food intake and body weight when compared NP and LP dams during pregnancy. However, the male pups body weight was

significantly reduced from birth to the 14th day of life in LP offspring when compared to NP male pups (Fig 1).



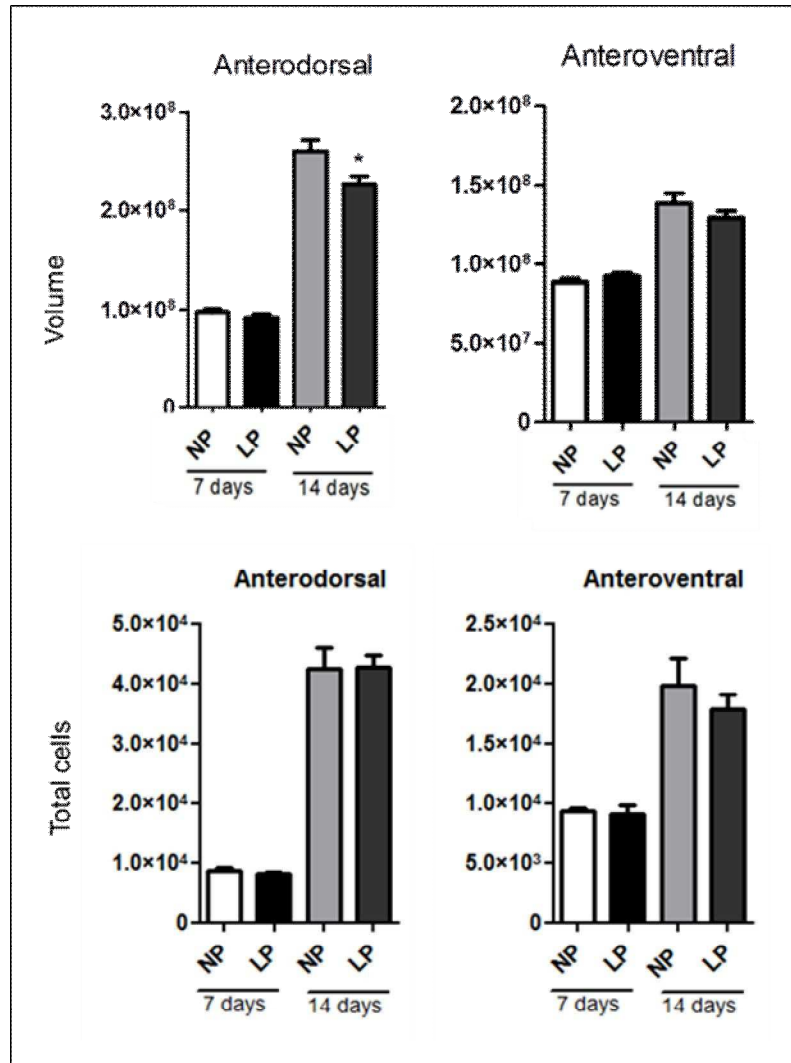
**Fig. 1.** Body weight at birth and with 7 and 14 days of life (\*\* $p=0.002$ , \*\*\* $p\leq 0.0001$ ).

### ***Corticosterone serum levels***

The corticosterone basal levels were not affected by gestational diet treatment in either postnatal day 7 (NP:  $1.1 \pm 0.02$ ,  $n=7$  vs. LP:  $1.1 \pm 0.03$ ,  $n=7$ ,  $p=0.8$ ) or 14 (NP:  $1.36 \pm 0.01$ ,  $n=7$  vs. LP:  $1.37 \pm 0.01$ ,  $n=8$ ,  $p = 0.5$ ) in both experimental groups.

### ***Stereology***

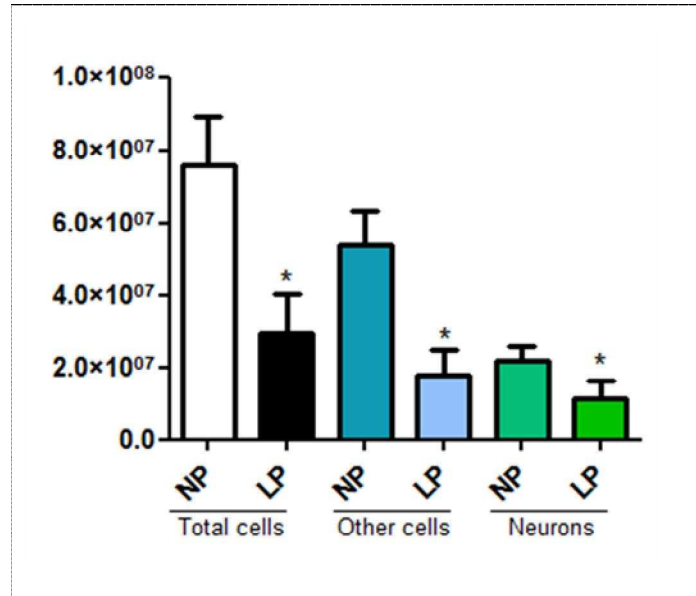
The volume of anterodorsal and anteroventral BNST divisions was not altered in 7 day-old offspring. Otherwise, 14 day-old LP rats showed 13% decreased anterodorsal BNST division volume when compared with NP offspring (Fig. 2). The total cells number was not altered in both divisions and time points.



**Fig 2.** Volume and cells number in the anterodorsal and anteroventral divisions of the BNST (\* $p= 0.04$ ).

### ***BNST total cells and neurons quantification***

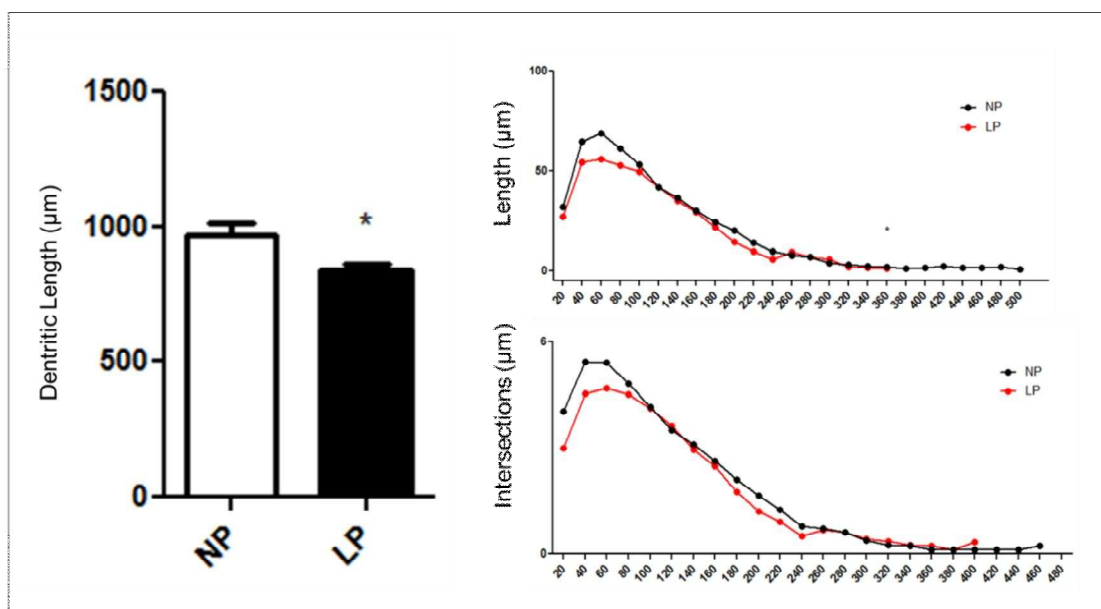
On the 14<sup>th</sup> postnatal day the BNST total cells number was 61% reduced in LP when compared to that found in age-matched NP group ( $p = 0.03$ ). The calculus of BNST neurons and non-neurons cells showed a reduction of 57,6% in the number of neurons ( $p=0.04$ ) and 66% in other cells ( $p=0.02$ ) (Fig.3).



**Fig 3.** Number of neurons and other cells in the BNST of 14 days old animals.

### ***Dendritic tree analysis***

We found significant reduction of 13,5% in dendritic length of BNST neurons from 14 days-old LP rats when compared with age-matched NP ( $p=0.04$ ). Sholl analysis revealed a reduction of 140 $\mu\text{m}$  at the length of dendrites and 13% reduced ramifications in the neurons of the LP offspring compared to NP rats (Fig.4).

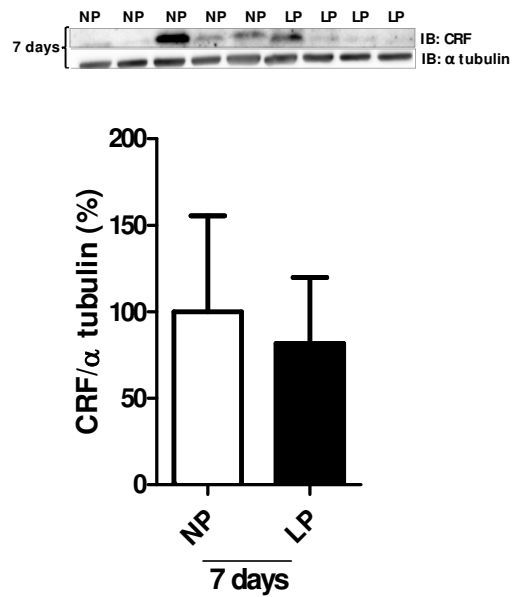
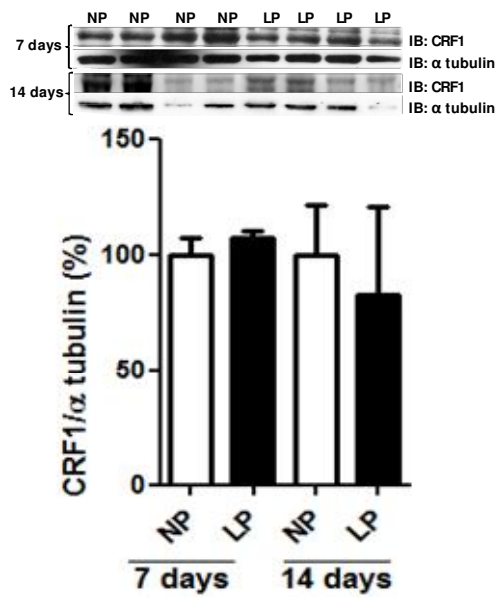
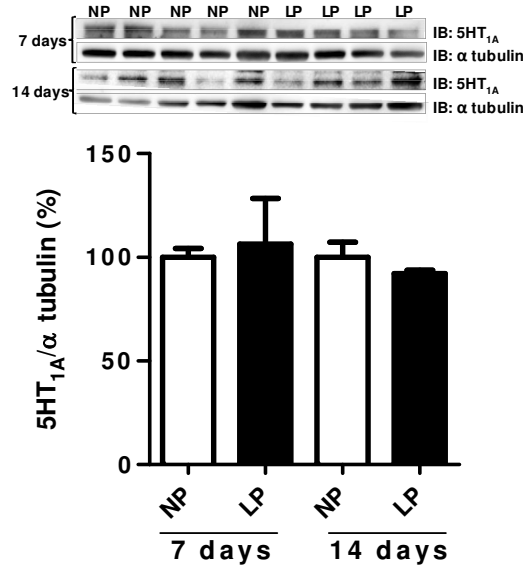
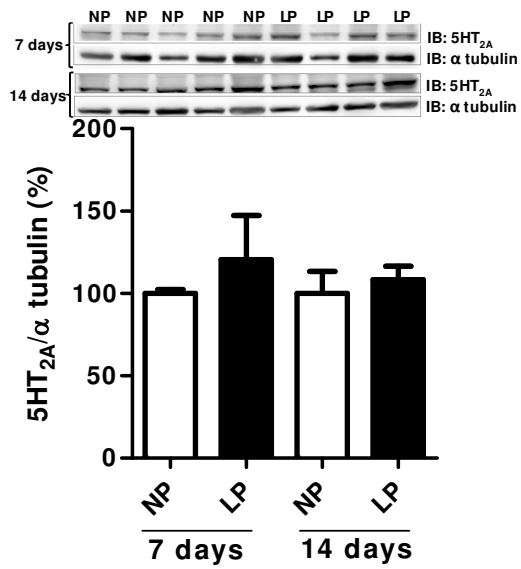
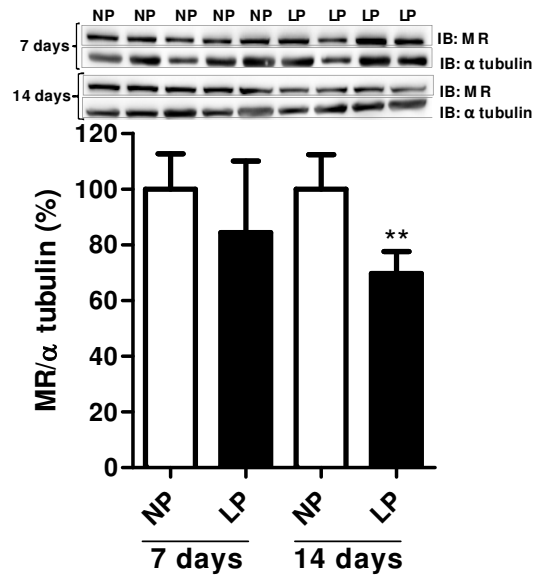
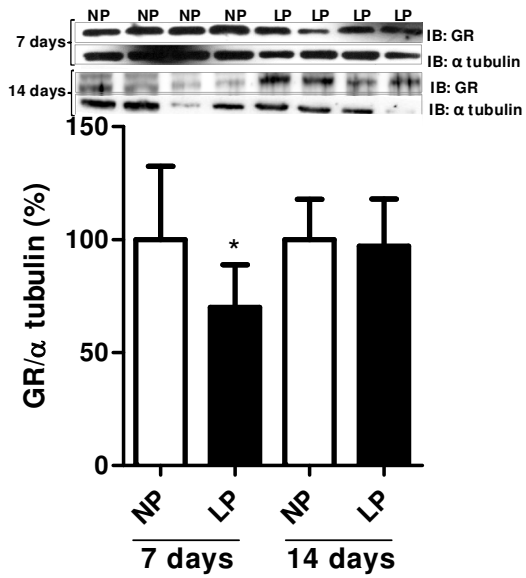


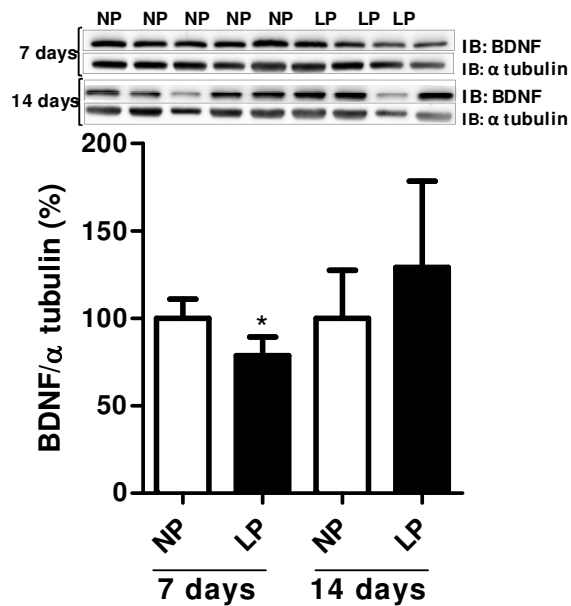
**Fig. 4.** The gestational protein restriction led to a decrease in the length of BNST dendrites after 14 days of life. By Sholl analysis we can observe the distance of dendrites from pericaria and dendrites bifurcations in LP and NP groups.

### ***Western blot analysis***

Western blot comparative analysis in the BNST of 7 day-old offspring from NP and LP groups revealed that the levels of GR and BDNF from LP group were significantly reduced in 31% (n=8,  $p=0.03$ ) and 22% (n=5,  $p=0.01$ ), respectively. The expression of CRF1 was significantly enhanced (7%, n=4,  $p=0.05$ ). In BNST was observed a reduction of MR (16%, n=4,  $p=0.16$ ) and CRF expression (19%, n=4,  $p=0.22$ ) associated with an enhance of 5HT1A (6%, n=4,  $p=0.29$ ) and 5HT2A (20%, n=4,  $p=0.09$ ) in LP offspring when compared to NP age-matched group, although expressions has not achieved statistical significance.

The comparative analysis in the 14 d-old offspring showed that the levels of MR and 5HT1A from LP group were significantly reduced in 31% (n=9,  $p=0.003$ ) and 8% (n=4,  $p=0.03$ ), respectively. Although do not statistically significant, the present study shows a reduced GR (3%, n=9,  $p=0.38$ ) and CRF1 (18%, n=9,  $p=0.14$ ) expressions and, enhanced 5HT2A (8%, n=8,  $p=0.77$ ) and BDNF (29% (n=5,  $p=0.15$ ) in LP when compared to NP age-matched group. (Fig.5).





**Fig 5.** Effects of maternal protein restriction on expression of BNST proteins. Protein levels for 5HT2A, 5HT1A, GR, MR, BDNF, CRF and CRF1 in the BNST of NP and LP animals at postnatal day 7 and 14. The results of scanning densitometry were expressed as relative to NP, assigning a value of 100% to the control rats. Columns and bars represent the mean  $\pm$  SD. \* $P \leq 0.05$ , NP versus LP.

### ***High-Performance Liquid Chromatography (HPLC)***

The HPLC analysis shows a significant increase in norepinephrine BNST concentration in 7 d-old and increased 5 HIAA BHST concentrations in 14 d-old LP offspring compared to NP age-matched group. Additionally, 14 day-old LP rats present a significant decreased of dopamine *turnover* when compared to control offspring (Table 1).

**Table 1: Determination of the concentration of neurotransmitters by HPLC analysis of neurochemical or BNST:** Values represent the mean  $\pm$  standard deviation. Statistical comparisons were made with the Student t test and statistical significance for  $p \leq 0.05$ .

	BNST					
	7 days			14 days		
	NP	LP	p	NP	LP	p
						0.07
<b>Norepinephrine</b>	83.5 ± 38.7 N=4	244 ± 26.7 N=4	0.007**	123 ± 13 N=5	162.5 ± 22 N=3	0.34
<b>Epinephrine</b>	5 ± 0.9 N=4	5.5 ± 0.66 N=4	0.4	12 ± 1.4 N=5	13 ± 1.1 N=3	0.204
<b>Dopamine</b>	23.8 ± 6 N=3	22.4 ± 2.3 N=3	0.42	28 ± 5.5 N=5	40.6 ± 16 N=3	0.04*
<b>DOPAC</b>	14.9 ± 4 N=3	22.7 ± 4.6 N=4	0.14	36 ± 4.431 N=5	21 ± 6 N=3	0.0006***
<b>5HIAA</b>	6. ± 0.9 N=4	7 ± 0.9 N=4	0.21	8 ± 0.46 N=4	20 ± 2 N=3	0.45
<b>5HT</b>	6.8 ± 0.87 N=4	7.7 ± 0.2 N=4	0.17	12 ± 1 N=3	12.7 ± 2 N=4	0.1
<b>TURNOVER 5HT</b>	1 ± 0.3 N=4	0.8 ± 0.03 N=3	0.31	1 ± 0.1 N=3	1.57 ± 0.36 N=3	0.01*
<b>TURNOVER DOPAMINE</b>	1.14 ± 0.1 N=3	1.25 ± 0.1 N=4	0.27	1.47 ± 0.09 N=4	1.1 ± 0.07 N=4	

## DISCUSSION

According to 2013 data from the WHO maternal malnutrition is still a worldwide public health issue and it is directly correlated with low weight at birth and to several health issues in adulthood. The concept of fetal programming defined by David Baker (Baker et al, 1989) establishes that insults occurring during gestation can lead to several diseases in adulthood such as metabolic and cardiovascular disorders. In the current study, we hypothesize an association between gestational protein restriction and BNST morphological and neurochemistry changes proposing that, the BNST is a CNS area in which permanent changes underlie, at least in part, the development of behavioral disorders in this experimental model. Nutritional undernutrition during critical periods of development may lead to several metabolic, morphological and neurochemical disorders. This study confirms previous studies of our laboratory (Mesquita et al, 2010; Lima et al, 2012; Scabora et al, 2013) showing a significant reduction of offspring birthweight in gestational protein-restricted group accompanied by renal, cardiac and neuronal dysfunctions when compared to NP offspring. These studies demonstrate that effect was



associated with a significant enhance in arterial blood pressure beyond the 7-week of age in LP offspring (Mesquita et al., 2010).

Otherwise, BNST cell quantification and stereology studies show a significant reduction of the total cells number, decreased ratio between neurons and non-neuron cells associated with reduced volume of the anterodorsal BNST division in 14 d-old LP offspring when compared to that found in age-matched NP group. Also, in the present study, Sholl analysis revealed a reduction of the length of dendrites and reduced ramifications in the neurons of the LP offspring. Several factors may be involved in fetal programming but the hypothesis more strongly considered involve a fetal high exposure to GC. Maternal undernutrition or protein restriction leads to increased and persistent fetal exposure to maternal GC, which promotes disruption of the HPA axis balance and consequently, faster fetal tissues and organs maturation (Drake, 2007). Recently, we and other authors have demonstrated that gestational protein malnutrition leads to CNS developmental changes particularly, by reducing dendritic arborization (Lopes et al, 2013; Kim Sung-Yon et al, 2013), the number of synaptic ends and, in the neural myelination (Lima & Voigt, 1999). Supporting the present data, we priory demonstrate that gestational protein restriction cause a significant reduction of the BNST dendritical length and in the anterodorsal BNST division volume in 16 wk-old protein-restricted offspring (submitted article). Since the BNST is highly plastic, studies have demonstrated that encephalic region during fetal development, is extremely vulnerable to environmental stresses and exposure to endogen and exogenous corticosterone high levels, which in turn lead to several morphological and functional disorders (for review see Hammack et al, 2010). In fact, exposure to unpredictable chronic psychological stress is associated with an increase in the volume and dendritic length in the BNST (Pêgo et al, 2008) and chronic immobilization paradigms increases dendritic arborization in BNST neurons (Vyas et al, 2002; 2003). The BNST is considered the main integrator nucleus of excitatory and inhibitory inputs that regulate the HPA axis (Forray & Gysling, 2004). Studies have shown that the gestational exposure to psychological stress (Weinstock et al, 1992) or by administration of stress hormones (Fameli et al,

1994) presents an increased corticosterone plasma level in the offspring. The activation of HPA axis promoted by stress stimulus ends with the release of corticosteroids by the adrenals (Ieraci et al, 2016). Corticosteroids are the main hormones for the maturation in the final days of gestation (Wood 2016) and the lack of balance in the expression of their receptors (MR and GR) can increase the vulnerability of the CNS to adverse effects (Sousa, et al 2008). In this work we observed a decrease in the expression of GR and MR both 7 and 14 postnatal days. Nevertheless the expression of MR was decreased in 28% in 14 days old animals. Endogenous GCs in basal conditions have higher affinity for MR (De Kloet et al, 1998) and the activation of MR seems to be involved with survival actions and is primarily involved in the maintenance of basal activity. Chronic stress can lead to the continuous activation of both receptors causing dendritic atrophy and deficits in synaptic plasticity (Sousa et al, 2008). Taking in account this results and once corticosteroids modulate structural alterations in the CNS that include changes in cellularity, structural volume and also, in synaptic and dendritic branching and morphology (Leão et al, 2007), we suppose that present finding may also be explained by this phenomenon.

Furthermore, the present work demonstrates by western blotting studies, a decreased expression of gluco- and mineralocorticoid receptors in 7 and 14-d old accompanied, by fall in BDNF and enhanced CRF1 receptor expression in the BNST of the 7-d old LP offspring when compared to NP control rats, despite of unchanged corticosterone plasma level. The basal levels of corticosterone may be used as an indication of higher or lower stress levels (Ventura-Silva et al, 2012). Several studies have demonstrated that the gestational exposure to stress (Weinstock et al, 1992) or the administration of stress hormones (Fameli et al, 1994) during gestation lead to an increase in the plasma concentration of corticosterone in the offspring. Prenatal stress is associated with alterations in the HPA axis in the offspring (for review see Charil et al, 2010). Previous studies from our lab (submitted article) have shown that animals that suffered gestational protein restriction have higher levels of plasmatic corticosterone at 16 weeks of age. In the current study, however, 7 and 14 d-old LP offspring compared to NP

group did not present any difference in the basal corticosterone levels. This results may be explained by well-characterized stress hyporesponsive period (SHRP) associated to decreased activity of HPA axis, that may last until the 14<sup>th</sup> postnatal-day (Sapolsky & Meane 1986). At this time, there is a reduced secretion of corticosterone, which remains low until the second postnatal week (Sapolsky & Meane 1986). The SHRP may assume as a protective mechanism ensuring low levels of glucocorticoids during early postnatal development (Mesquita et al, 2007). BDNF is strongly connected with the serotonergic system and both are involved with memory processes and mood (Van Donkelaar et al, 2009). The two systems may act together to regulate neuronal plasticity and survival of new neurons (Mattson, 2004). Our study shows a significant decrease in the expression of BDNF in 7-d old LP offspring. The current study shows a significant decrease in the neurons and non-nuron cells and in volume of anterodorsal portion of BNST of 14-d old LP offspring. We may state that this reduction may be associated with the decrease in the expression of BDNF since this factor acts directly in glial and neuronal progenitor cells (Rial, 2016). Also, sustained our study in this model, previous results have been showed that stress exposure decreases the level of BDNF in brain regions associated with depression (Duman, 2004; Barrientos et al, 2003).

In addition, the 14 d-old LP offspring BNST presents a reduced 5HT<sub>1A</sub> receptor subtype levels, reciprocally accompanied, by increased 5HT<sub>2A</sub> receptors compared to age-matched NP offspring. It is also known that different ways of exposure to stress can alter the serotonergic system which is involved in emotional behavior and anxiety disorders (Ressler & Nemeroff, 2000). Both chronic stress and treatment with anxiogenic drugs have been shown to activate a subset of serotonergic neurons in raphe nucleus that has main targets limbic areas such as the BNST (Grahn et al, 1999a; Lowry et al, 2000; Singewald et al, 2000). 5HT<sub>1A</sub> and 5HT<sub>2A</sub> are expressed widely throughout the central nervous system including neocortex, hippocampus, septum, amygdala, raphe nucleus, basal ganglia, thalamus and the olfactory tubercle. Especially high concentrations of these receptors on the apical dendrites of pyramidal cells in layer V of the cortex

may modulate cognitive processes, working memory and attention (Ciranna, 2006). The mammalian 5HT<sub>1A</sub> and 5HT<sub>2A</sub> are subtypes of the serotonin receptor G protein-coupled receptor (GPCR). 5-HT<sub>1A</sub> receptor agonists are involved in neuromodulation of behavioral activity, learning and memory in rodents. Activation of central 5-HT<sub>1A</sub> receptors triggers the inhibition of norepinephrine and enhances acetylcholine release, depending on species and areas of the brain. Also, 5-HT<sub>1A</sub> receptor activation has been shown to increase dopamine release in the medial prefrontal cortex, striatum, and hippocampus, and may be useful for improving the symptoms of schizophrenia and Parkinson's disease. 5-HT<sub>1A</sub> receptor agonists relieve the anxiety and depression mainly by synaptic serotonin increasing concentration. The 5HT<sub>2A</sub> is the main excitatory receptor subtype among the GPCRs for serotonin (5-HT). Study has describes that overdensity of post-synaptic 5HT<sub>2A</sub> receptor is involved in the pathogenesis of depression. Interestingly stressors that activate the BNST also activate central serotonergic systems (Dilts & Boadle-Biber, 1995. Grahn et al, 1999b. Takase al, 2004). Additionally, several studies have implicated the serotonergic system in the modulation of fear and anxiety-like behavior (Graeff et al, 1996; Handley et al, 1993; Handley, 1995; Lowry et al, 2005). In the present study, we may hypothesized that reduced BNST 5HT<sub>1A</sub> and elevated 5HT<sub>2A</sub> expression may be related to anxiogenic and fear behavior observed for us, in previous study (submitted article).

Additionally, the BNST of 7-d old LP rats presents an enhanced level of norepinephrine compared with the NP age-matched offspring. This phenomenon was accompanied by decreased BNST dopamine turnover and DOPAC level, a metabolite of the neurotransmitter dopamine in 14 day-old protein-restricted offspring. As mentioned above, it is widely known that catecholamine play an important role in the neurochemistry of the brain and are involved in a series of brain functions among them the response to fear and anxiety. Sympathetic nervous terminals as well as chromaffin cells in adrenals are the main sources of circulating catecholamines (Lympelopoulos et al, 2016). In the CNS catecholamines work as neurotransmitters in the synaptic cleft and are a crucial part of the maintenance of

homeostasis quickly responding to any stressor that threatens the homeostasis of the organism (Andreis, 2016; Riedemann et al, 2010).

Our findings show a significant increase in the BNST norepinephrine levels of as well as serotonin precursor (5HIAA) in 7-d old LP offspring. The BNST is a structure that receives projections from noradrenergic receptors from the brain stem and for this plays a series of responses to stress. Nociceptive stimuli or immobilization stress in rats cause an increase in the BNST norepinephrine suggesting that an aversive stimulus activates noradrenergic projections to the BNST (Onaka, 1998; Pacak et al, 1995).

Finally, the present study shows a reduced dopamine turnover and 3,4-Dihydroxyphenylacetic acid (DOPAC), a metabolite of the neurotransmitter dopamine, BNST concentration, just in 14 d-old LP offspring. Here, these results suggest that at 14 d-old LP animals present a decreased neuronal release and degradation of dopamine. Oliveira et al., (2012) have demonstrated that prenatal administration of dexamethasone does not alter the concentration of dopamine, DOPAC and HVA in the BNST of the adult offspring. Nevertheless there were alterations in the HPA axis (Oliveira et al, 2006), which can act as a modulator of dopaminergic circuits (Piazza & Le Moal. 1996).

Thus, the current study, as far as we know is the first description of the modulation of dendritic plasticity, morphology and neurochemistry of the BNST in the early life, by protein restriction during developmental period. These interesting findings may represent the adaptation during embryonic development to exposure to elevated maternal corticosteroids as a consequence of nutritional stress. It is important to consider that different stimuli as well as intensities and time can lead to different responses in the CNS morphology (Oliveira et al, 2012). Also, additional studies must be done, but the present study suggests strongly that morphological and neurochemical results may be associated with the development of psychiatric disorders in adulthood.

## REFERECES

ANDREIS DT; SINGER M. Catecholamines for inflammatory shock: a Jekyll-and-Hyde conundrum. **Intensive Care Med.** 2016 Feb 12. [Epub ahead of print]

ASHTON, N. Perinatal development and adult blood pressure. **Braz J Med Biol Res** 2000; 33:731-40.

BARKER, DJ. The fetal and infant origins of disease. **Eur J Clin Invest.** 1995; 25:457-63.

BARKER, DJP.; OSMOND, C.; GOLDING, J.; KUH, D.; WADSWORTH, MEJ. Growth in utero, blood pressure in childhood and adult life, and mortality from cardiovascular disease. **BMJ.** 1989; 298:564-567.

BARRIENTOS RM; SPRUNGER DB; CAMPEAU S; HIGGINS EA; WATKINS LR; RUDY JW; MAIER SF. Brain-derived neurotrophic factor mRNA downregulation produced by social isolation is blocked by intrahippocampal interleukin-1 receptor antagonist. **Neuroscience.** 2003;121(4):847-53.

BENEDIKTSSON, R.; LINDSAY, RS.; NOBLE, J.; SECKL, JR.; EDWARDS, C.R. Glucocorticoid exposure in utero: new model for adult hypertension. **Lancet.** 1993; 34: 339-41

BURD, L.; SEVERUD, R.; KERBESHIAN, J.; KLUG, MG. Prenatal and perinatal risk factors for autism. **J Perinat Med** 1999; 27: 441-450.

CHARIL A; LAPLANTE DP; VAILLANCOURT C; KING S. Prenatal stress and brain development. **Brain Res Rev.** 2010 Oct 5;65(1):56-79. doi: 10.1016/j.brainresrev.2010.06.002. Epub 2010 Jun 13.

CIRANNA L. Serotonin as a modulator of glutamate- and GABA-mediated neurotransmission: implications in physiological functions and in pathology. **Curr Neuropharmacol.** 2006 Apr;4(2):101-14.

COROMINAS M.; RONCERO C.; CASAS M. Corticotropin releasing factor and neuroplasticity in cocaine addiction. **Life Sci.** 2010; 86: 1–9.

DAVIS, M. The role of the amygdala in fear and anxiety. **Ann. Rev. Neurosci.** 1992; 15:353–375.

DE KLOET ER; VREUGDENHIL E; OITZL MS; JOËLS M. Brain corticosteroid receptor balance in health and disease. **Endocr Rev.** 1998 Jun;19(3):269-301.

DILTS RP; BOADLE-BIBER MC. Differential activation of the 5-hydroxytryptamine-containing neurons of the midbrain raphe of the rat in response to randomly presented inescapable sound. **Neurosci Lett.** 1995 Oct 13;199(1):78-80.

DING, Y X; SHI, Y HAN, W J; CUI, H. Regulation of glucocorticoid related genes and receptors/regulatory enzyme expression in intrauterine growth restriction filial rats. **Life Sci.** 2016 Feb 23. pii: S0024-3205(16)30128-X. doi: 10.1016/j.lfs.2016.02.079. [Epub ahead of print].

DRAKE AJ; TANG JI; NYIRENDA MJ. Mechanisms underlying the role of glucocorticoids in the early life programming of adult disease. *Clin Sci (Lond)*. 2007 Sep;113(5):219-32.

DUMAN RS. Role of neurotrophic factors in the etiology and treatment of mood disorders. **Neuromolecular Med.** 2004;5(1):11-25.

FAMELI M; KITRAKI E; STYLIANOPOULOU F. Effects of hyperactivity of the maternal hypothalamic-pituitary-adrenal (HPA) axis during pregnancy on the

development of the HPA axis and brain monoamines of the offspring. **Int J Dev Neurosci.** 1994 Nov;12(7):651-9.

FONTENOT, M.B.; KAPLAN, J.R.; MANUCK, S.B.; ARANGO, V.; MANN, J.J. Long-term effects of chronic social stress on serotonergic indices in the prefrontal cortex of adult male cynomolgus macaques. **Brain Res.** 1995; 705:105-108.

FORRAY MI; GYSLING K. Role of noradrenergic projections to the bed nucleus of the stria terminalis in the regulation of the hypothalamic-pituitary-adrenal axis. **Brain Res Brain Res Rev.** 2004 Dec;47(1-3):145-60.

GIBB, R.; KOLB, B. A method for vibratome sectioning of Golgi-Cox stained whole rat brain. **J Neurosci Methods** 1998; 79: 1–4.

GLASER, E.M.; VAN, D.E.R.; LOOS, H. Analysis of thick brain sections by obverse-reverse computer microscopy: application of a new, high clarity Golgi-Nissl stain. **J Neurosci Methods** 1981; 4:117–125.

GOMEZ-SANCHEZ, EP. Mineralocorticoid receptors in the brain and cardiovascular regulation: minority rule? **Trends Endocrinol Metab.** 2011 May;22(5):179-87. Epub 2011 Mar 21.

GRAEFF, F.G.; GUIMARÃES, F.S.; DE ANDRADE, T.G.; DEAKIN, J.F. Role of 5-HT in stress, anxiety, and depression. **Pharmacol Biochem Behav.** 1996; 54(1):129-41.

GRAHN RE; MASWOOD S; MCQUEEN MB; WATKINS LR; MAIER SF. Opioid-dependent effects of inescapable shock on escape behavior and conditioned fear responding are mediated by the dorsal raphe nucleus. **Behav Brain Res.** 1999a Mar;99(2):153-67.



GRAHN RE; WILL MJ; HAMMACK SE; MASWOOD S; MCQUEEN MB; WATKINS LR; MAIER SF. Activation of serotonin-immunoreactive cells in the dorsal raphe nucleus in rats exposed to an uncontrollable stressor. **Brain Res.** 1999b Apr 24;826(1):35-43.

HAMMACK, SE.; ROMAN, CW.; LEZAK, KR.; KOCHO-SHELLENBERG, M. GRIMMIG, B.; FALLS, WA.; BRAAS, K; MAY, V. Roles for pituitary adenylate cyclase-activating peptide (PACAP) expression and signaling in the bed nucleus of the stria terminalis (BNST) in mediating the behavioral consequences of chronic stress. **J Mol Neurosci.** 2010;42(3):327-40. Epub 2010 Apr 20.

HAMMACK, S.E.; GUO, J.D.; HAZRA, R.; DABROWSKA, J.; MYERS, K.M.; RAINNIE, D.G. The response of neurons in the bed nucleus of the stria terminalis to serotonin: implications for anxiety. **Prog Neuropsychopharmacol Biol Psychiatry.** 2009; 13;33(8):1309-20. Epub 2009 May 23.

HANDLEY, S.L. 5-Hydroxytryptamine pathways in anxiety and its treatment. **Pharmacol Ther** 1995; 66:103–48.

HANDLEY, S.L.; MCBLANE, J.W.; CRITCHLEY, M.A.; NJUNG'E, K. Multiple serotonin mechanisms in animal models of anxiety: environmental, emotional and cognitive factors. **Behav Brain Res.** 1993; 20;58(1-2):203-10.

HERCULANO-HOUZEL S; LENT R. Isotropic fractionator: a simple, rapid method for the quantification of total cell and neuron numbers in the brain. **J Neurosci.** 2005 Mar 9;25(10):2518-21.

HOLMES, A.; HEILIG, M.; RUPNIAK, N.M.; STECKLER, T.; GRIEBEL, G. Neuropeptide systems as novel therapeutic targets for depression and anxiety disorders. **Trends Pharmacol. Sci.** 2003; 24:580–588.

HOLSBOER F. Corticotropin-releasing hormone modulators and depression. *Curr. Opin. Investig. Drugs.* 2003; 4:46–50.

KIM SY; ADHIKARI A; LEE SY; MARSHEL JH; KIM CK; MALLORY CS; LO M, PAK S; MATTIS J; LIM BK; MALENKA RC; WARDEN MR; NEVE R,; TYE KM; DEISSEROTH K.  
Diverging neural pathways assemble a behavioural state from separable features in anxiety. *Nature.* 2013 Apr 11;496(7444):219-23. doi: 10.1038/nature12018. Epub 2013 Mar 20.

KREAM, J.; MULAY, S.; FUKUSHIMA, DK.; SOLOMON, S. Determination of plasma dexamethasone in the mother and the newborn after administration of the hormone in a clinical trial. *J Clin Endocrinol Metab.* 1983; 56(1):127-33.

LANGLEY-EVANS, SC. Intrauterine programming of hypertension by glucocorticoids. *Life Sci.* 1997; 60:1213-21.

LANGLEY-EVANS, SC.; WELHAM, SJ.; SHERMAN, RC.; JACKSON, AA. Weanling rats exposed to maternal low-protein diets during discrete periods of gestation exhibit differing severity of hypertension. *Clin Sci (Lond).* 1996; 91:607-15.

LEÃO P; SOUSA JC; OLIVEIRA M; SILVA R; ALMEIDA OF; SOUSA N. Programming effects of antenatal dexamethasone in the developing mesolimbic pathways. *Synapse.* 2007 Jan;61(1):40-9.

LEONARD BE. The HPA and immune axes in stress: the involvement of the serotonergic system. *Eur. Psychiatry.* 2005; 20:302–306.

LIMA AD; VOIGT T. Astroglia inhibit the proliferation of neocortical cells and prevent the generation of small GABAergic neurons invitro. *Eur J Neurosci.* 1999 Nov;11(11):3845-56.

LIMA MC; SCABORA JE; LOPES A; MESQUITA FF; TORRES D; BOER PA; GONTIJO JAR. Early changes of hypothalamic angiotensin II receptors expression in gestational protein-restricted offspring: effect on water intake, blood pressure and renal sodium handling. **J Renin Angiotensin Aldosterone Syst.** 2013 Sep;14(3):271-82. doi: 10.1177/1470320312456328. Epub 2012 Aug 30.

LINGAS, R.; DEAN, F.; MATTHEWS, S.G. Maternal nutrient restriction (48h) modifies brain corticosteroid receptor expression and endocrine function in the fetal guinea pig. **Brain Res**, 1999; 846: 236-242.

LOPES A; TORRES DB; RODRIGUES AJ; CERQUEIRA JJ; PÊGO JM; SOUSA N; GONTIJO JA; BOER PA. Gestational protein restriction induces CA3 dendritic atrophy in dorsal hippocampal neurons but does not alter learning and memory performance in adult offspring. **Int J Dev Neurosci.** 2013 May;31(3):151-6. doi: 10.1016/j.ijdevneu.2012.12.003. Epub 2012 Dec 30

LOWRY CA; RODDA JE; LIGHTMAN SL; INGRAM CD. Corticotropin-releasing factor increases in vitro firing rates of serotonergic neurons in the rat dorsal raphe nucleus: evidence for activation of a topographically organized mesolimbocortical serotonergic system. **J Neurosci.** 2000 Oct 15;20(20):7728-36.

LOWRY, C.A.; JOHNSON, P.L.; HAY-SCHMIDT, A.; MIKKELSEN, J.; SHEKHAR A. Modulation of anxiety circuits by serotonergic systems. **Stress** 2005; 8:233–46.

LYMPEROPOULOS A; BRILL A; MCCRINK KA. GPCRs of adrenal chromaffin cells & catecholamines: The plot thickens. **Int J Biochem Cell Biol.** 2016 Feb 3. pii: S1357-2725(16)30021-8. doi: 10.1016/j.biocel.2016.02.003. [Epub ahead of print]

MATTSON MP; MAUDSLEY S; MARTIN B. BDNF and 5-HT: a dynamic duo in age-related neuronal plasticity and neurodegenerative disorders. **Trends Neurosci.** 2004 Oct;27(10):589-94.

MCDONALD AJ. Neurons of the bed nucleus of the stria terminalis: a golgi study in the rat. **Brain Res Bull.** 1983 Jan;10(1):111-20.

MERALI Z, KHAN S, MICHAUD, D.S.; SHINOSITOL, PHOSPATEPY SA, ANISMAN H. Does amygdaloid corticotropin-releasing hormone (CRF) mediate anxiety-like behaviors? Dissociation of anxiogenic effects and CRF release. **Eur. J. Neurosci.** 2004; 20:229–239.

MESQUITA AR; PÊGO JM; SUMMAVIELLE T; MACIEL P; ALMEIDA OF; SOUSA N. Neurodevelopment milestone abnormalities in rats exposed to stress in early life. **Neuroscience.** 2007 Jul 29;147(4):1022-33. Epub 2007 Jun 22.

MESQUITA FF, GONTIJO JAR, BOER PA. Maternal undernutrition and the offspring kidney: from fetal to adult life. **Braz J Med Biol Res.** 2010 Nov;43(11):1010-8. Epub 2010 Oct 29.

MILLAN MJ. Serotonin 5-HT<sub>2C</sub> receptors as a target for the treatment of depressive and anxious states: focus on novel therapeutic strategies. **Therapie.** 2005; 60:441–460.

MULLER MB. Limbic corticotropin-releasing hormone receptor 1 mediates anxiety-related behavior and hormonal adaptation to stress. **Nat. Neurosci.** 2003; 6:1100–1107.

NESTLER EJ.; BARROT, M.; DILEONE, RJ.; EISCH, AJ.; GOLD, SJ.; MONTEGGIA, LM. Neurobiology of depression. **Neuron.** 2002; 34:13–25.

OLIVEIRA M; BESSA JM; MESQUITA A; TAVARES H; CARVALHO A; SILVA R; PÊGO JM; CERQUEIRA JJ; PALHA JA; ALMEIDA OF; SOUSA N. Induction of a hyperanxious state by antenatal dexamethasone: a case for less detrimental natural corticosteroids. **Biol Psychiatry**. 2006 May 1;59(9):844-52. Epub 2005 Sep 28.

OLIVEIRA M; RODRIGUES AJ; LEÃO P; CARDONA D; PÊGO JM; SOUSA N. The bed nucleus of stria terminalis and the amygdala as targets of antenatal glucocorticoids: implications for fear and anxiety responses. **Psychopharmacology** (Berl). 2012 Apr;220(3):443-53. doi: 10.1007/s00213-011-2494-y. Epub 2011 Sep 21.

ONAKA, T.; YAGI, K. Role of noradrenergic projections to the bed nucleus of the stria terminalis in neuroendocrine and behavioral responses to fear-related stimuli in rats. **Brain Res**. 1998; 788:287– 293.

PACAK K; MCCARTY R; PALKOVITS M; KOPIN IJ; GOLDSTEIN DS. Effects of immobilization on in vivo release of norepinephrine in the bed nucleus of the stria terminalis in unconscious rats. **Brain Res**. 1995 Aug 7;688(1-2):242-6.

PÊGO, JM.; MORGADO, PLG.; CERQUEIRA, JJ., ALMEIDA, OFX.; SOUSA, N. Dissociation of the morphological correlates of stress-induced anxiety and fear. **European Journal of Neuroscience** 2008; 27:1503–1516.

PIAZZA PV; LE MOAL ML. Pathophysiological basis of vulnerability to drug abuse: role of an interaction between stress, glucocorticoids, and dopaminergic neurons. **Annu Rev Pharmacol Toxicol**. 1996;36:359-78.

RESSLER KJ; NEMEROFF CB. Role of serotonergic and noradrenergic systems in the pathophysiology of depression and anxiety disorders. **Depress Anxiety**. 2000;12 Suppl 1:2-19.

RIAL D; LEMOS C; PINHEIRO H; DUARTE JM; GONÇALVES FQ; REAL JI; PREDIGER RD; GONÇALVES N; GOMES CA; CANAS PM; AGOSTINHO P CUNHA RA. Depression as a Glial-Based Synaptic Dysfunction. **Front Cell Neurosci**. 2016 Jan 22;9:521. doi: 10.3389/fncel.2015.00521. eCollection 2015.

RIEDEMANN T; PATCHEV AV; CHO K; ALMEIDA OF. Corticosteroids: way upstream. **Mol Brain**. 2010 Jan 11;3:2. doi: 10.1186/1756-6606-3-2.

SAPOLSKY RM; MEANEY MJ. Maturation of the adrenocortical stress response: neuroendocrine control mechanisms and the stresshypo-responsive period. **Brain Res**. 1986 Mar;396(1):64-76.

SCABORA JE; LIMA MC; LOPES A; LIMA IP; MESQUITA FF; TORRES DB; BOER PA; GONTIJO JAR. Impact of taurine supplementation on blood pressure in gestational protein-restricted offspring: Effect on the medial solitary tract nucleus cell numbers, angiotensin receptors, and renal sodium handling. **J Renin Angiotensin Aldosterone Syst**. 2015 Mar;16(1):47-58. doi: 10.1177/1470320313481255. Epub 2013 Mar 6.

SINGEWALD N; KOUVELAS D; KAEHLER ST; SINNER C; PHILIPPU A. Peripheral chemoreceptor activation enhances 5-hydroxytryptamine release in the locus coeruleus of conscious rats. **Neurosci Lett**. 2000 Jul 28;289(1):17-20.

SOUSA N; CERQUEIRA JJ; ALMEIDA OF. Corticosteroid receptors and neuroplasticity. **Brain Res Rev**. 2008 Mar;57(2):561-70. Epub 2007 Jul 17.

STEWART, PM.; WHORWOOD, CB.; MASON, J.I. Type 2 11 beta-hydroxysteroid dehydrogenase in foetal and adult life. **J Steroid Biochem Mol Biol.** 1995; 55(5-6):465-71

TAKASE LF; NOGUEIRA MI; BARATTA M; BLAND ST; WATKINS LR; MAIER SF; FORNAL CA; JACOBS BL. Inescapable shock activates serotonergic neurons in all raphe nuclei of rat. **Behav Brain Res.** 2004 Aug 12;153(1):233-9.

VAN DONKELAAR EL; VAN DEN HOVE DL; BLOKLAND A; STEINBUSCH HW; PRICKAERTS J. Stress-mediated decreases in brain-derived neurotrophic factor as potential confounding factor for acutetryptophan depletion-induced neurochemical effects. **Eur Neuropsychopharmacol.** 2009 Nov;19(11):812-21. doi: 10.1016/j.euroneuro.2009.06.012. Epub 2009 Jul 28.

VENTURA-SILVA AP; PÊGO JM; SOUSA JC; MARQUES AR; RODRIGUES AJ; MARQUES F; CERQUEIRA JJ; ALMEIDA OF; SOUSA N. Stress shifts the response of the bed nucleus of the stria terminalis to an anxiogenic mode. **Eur J Neurosci.** 2012 Nov;36(10):3396-406. doi: 10.1111/j.1460-9568.2012.08262.x. Epub 2012 Aug 29.

VYAS, A.; BERNAL, S.; CHATTARJI, S. Effects of chronic stress on dendritic arborization in the central and extended amygdala. **Brain Res.** 2003; 7;965(12):290-4.

VYAS, A.; MITRA, R.; SHANKARANARAYANA, R.B.S.; CHATTARJI S. Chronic stress induces contrasting patterns of dendritic remodeling in hippocampal and amygdaloid neurons. **Neurosci.** 2002 1:22(15)6810-8.

WALKER, D.L.; TOUFEXIS, D.J.; DAVIS, M. Role of the bed nucleus of the stria terminalis versus the amygdala in fear, stress, and anxiety. **European Journal of Pharmacology** 2003; 463:199– 216

WALKER, DL.; TOUFEXIS, DJ.; DAVIS, M. Role of the bed nucleus of the stria terminalis versus the amygdale in fear, stress, and anxiety. **European Journal of Pharmacology** 2003; 463:199– 216

WEINSTOCK M; MATLINA E; MAOR GI; ROSEN H; MCEWEN BS. Prenatal stress selectively alters the reactivity of the hypothalamic-pituitary adrenal system in the female rat. **Brain Res.** 1992 Nov 13;595(2):195-200

WELBERG, L.A.; SECKL, J.R. Prenatal stress, glucocorticoids and the programming of the brain. **J Neuroendocrinol** 2001; 13: 113-128.

WOOD CE; KELLER-WOOD M. The critical importance of the fetal hypothalamus-pituitary-adrenal axis. **F1000Res.** 2016 Jan 28;5. pii: F1000 Faculty Rev-115. doi: 10.12688/f1000research.7224.1. e Collection 2016.

WORLD HEALTH ORGANIZATION. Global targets 2025 to improve maternal, infant and young child nutrition. Available from: [http://www.who.int/nutrition/topics/nutrition\\_globaltargets2025/en/index.html](http://www.who.int/nutrition/topics/nutrition_globaltargets2025/en/index.html) ( cited 30 May 2013).



## 4.2 Artigo 2

### **GESTATIONAL PROTEIN RESTRICTION ALTERS EARLY AMYGDALA NEUROCHEMISTRY IN MALE OFFSPRING**

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#### ***Abstract***

The amygdala's cytology and neurochemical composition has been poorly documented in gestational protein-restricted offspring, therefore, in the present

study we investigate the effects of gestational protein restriction on whole amygdala neurochemical compound in parallel with cytological content and neuron structure (basolateral amygdala) in male offspring, in key moments of post-natal neural development. The current study shows a significant decrease in body birthweight that remain up to 14 day of age, in gestational protein-restricted offspring. Additionally, this study also confirms several previous that have shown that protein restriction intake did not alter the brain weight in youth or adults offspring. In the current study, the amygdala neuronal 3D dendritic analysis of dendrites length and dendritic ramifications by Sholl analysis, was not altered in 14 d-old NP compared with age-matched gestational protein-restricted offspring. Also, the amygdala neurons and non-neuronal cells number was similar in 14 day-old LP and NP offspring. In the present study we also investigate the effects of gestational protein restriction on whole amygdala neurochemical compound in male offspring, in key moments of post-natal neural development. Here, we demonstrate significant decrease in the amygdala content of norepinephrine, epinephrine, dopamine, CRF and BDNF in 7 day-old rats, as well as reduction in the expression GR and MR and CRF in 14 day-old LP offspring. In conclusion, the amygdala neurochemical changes observed in this study may contribute to behavioral alterations induced by gestational protein restriction and may also be a primer for alterations in other brain regions. Also, these findings may represent the adaptation during embryonic development to exposure to elevated maternal corticosteroids as a consequence of nutritional stress.

**Keywords:** Amygdala, gestational protein-restricted intake, neuron morphology, neurochemistry, dendritic analysis.

### ***Introduction***

The concept of “fetal programming” suggests that the fetus may be programmed during intrauterine development (Barker, 1995; Lucas, 1991). Stressful stimuli during this period may be risk factors for the development of

neuropsychiatric disorders in adulthood (Borges, 2014). Gestational protein restriction promotes an increased fetal exposure to maternal glucocorticoids by decreased placental concentration and activity of the enzyme type-2 11  $\beta$ -HSD2, which promotes faster maturation of fetal tissues and organs leading to a decreased birthweight offspring and, increasing risk of behavioral disorders (Lopes, 2013; Mesquita, 2010; Scabora, 2012, Burd et al., 1999).

One of the mechanisms involved in the development of these disorders is the elevated exposure of the fetus to maternal glucocorticoids (GC) due to a low concentration and decreased activity of the placental enzyme 11beta-hydroxysteroid dehydrogenase (11 $\beta$ -HSD) type 2 (Benediktsson et al, 1993; Stewart et al, 1995; Langley-Evans et al, 1996; Langley-Evans, 1997). These prenatal alterations may lead to a chronic increase in GC as well as exacerbated response to stressful stimuli in adulthood (for review see Welberg & Seckl, 2001). There is a wide range of evidence that shows that the postnatal activity of the hypothalamic-pituitary-adrenal (HPA) axis, the main mechanism of stress-response in the brain, can be altered by prenatal events (Barker, 1995). Recently, we have demonstrated hyperanxious phenotype associated with atrophy of the dendritic neurons arborization from bed nucleus of the stria terminalis (BNST) in gestational protein-restricted adult offspring (unpublished data). No work has shed light on the amygdala morphology and neurochemical compounds of gestational protein restriction programmed animals.

The amygdala plays a crucial role coordinating behavioral, autonomic, and neuroendocrine stress responses, via mostly excitatory influences on the hypothalamus and brainstem (Aggleton, 2000). Anxiety-like behavior and fear-enhancing effect are enhanced by amygdala steroids stimulation (Venkova et al 2010) through local activation of high density of MR and GR (Herman et al 1989; Oitzl et al 2001). In the central amygdaloid nucleus (CeA), MR and GR are expressed in corticotrophin-releasing factor (CRF) neurons, a key modulator of stress-related anxiety and the role of the amygdala in the modulation of the HPA axis, suggesting a regulatory effect of glucocorticoid on this nucleus (Honkaniemi et al., 1992; Davis, 1992; Merali et al., 2004; Muller et al., 2003). Also, studies have

demonstrate that type 1 CRF receptor could modulates a number of neurotransmitter systems such as serotonin (5HT) and dopamine (Millan, 2005; Holsboer, 2003; Nestler et al., 2002; Leonard, 2005; Holmes et al, 2003). In addition, they may be interacting with brain derived neurotrophic factor (BDNF). This network is deeply involved in fear and anxiety (Hammack et al, 2009; Duman, 2004; Kumari, et al 2016).

The amygdala's cytology and neurochemical composition has been poorly documented in gestational protein-restricted offspring, therefore, in ten present study we investigate the effects of gestational protein restriction on whole amygdala neurochemical compound in parallel with cytological content and neuron structure (basolateral amygdala) in male offspring, in key moments of post-natal neural development.

### ***Materials and methods***

***Animals and treatments*** - The experiments were conducted on age-matched female offspring of sibling-mated Wistar Hannover rats (250-300g). The experiments were done in accordance with the general guidelines established by the Brazilian College of Animal Experimentation (COBEA) and approved by the Institutional Ethics Committee (CEEA/UNICAMP #3908-1) and National Institutes of Health guidelines on animal care and experimentation and approved by Director General Veterinary (DGV; the Portuguese National Institute of Veterinary 023-432/08.30.2013). A part of our site colonies originated from the breeding stock supplied by Charles River Laboratories, Barcelona, Spain. Another part was originated from a breeding stock supplied by CEMIB/Unicamp, Campinas, SP, Brazil. The animals were housed in pairs under standard laboratory conditions (lights on from 8 a.m. to 8 p.m.) and had access to food and water ad libitum and followed up to 12 weeks of age. The rats were placed to mate and the day that sperm were seen in the vaginal smear was designated as day 1 of pregnancy. The dams were divided in two groups: one maintained on isocaloric standard rodent laboratory chow with normal protein content [NP] (17% protein) and other received a diet with low protein content [LP] (6% protein) ad libitum throughout the entire pregnancy. Food consumption was determined every day (subsequently

normalized for body weight). All groups returned to the NP chow intake after delivery. On the day of birth the male pups were weighted and they were kept only 8 pups per female. At 7<sup>th</sup> and 14<sup>th</sup> postnatal day the brain, thymus and adrenals were weighted.

***Morphological analyses methods*** - At 7<sup>th</sup> and 14<sup>th</sup> postnatal day male rats from NP (n=27/per age from 10 different mothers) and LP (n=27/per age from 10 different mothers) groups were deeply anesthetized with a mixture of ketamine and xylazine (75 and 10mg/kg respectively, i.p.) and monitoring the corneal reflex controlled the level of anesthesia. For isotropic fractioning the rats were transcardially perfused with saline containing heparin (5%, for 15 min, under constant pressure) followed by fixative solution: 0.1M phosphates buffer (PB; pH 7.4) containing 4% (w/v) paraformaldehyde. For 3D reconstruction we used only saline for perfusion.

***Isotropic fractionator method*** - For total cells and neuron quantification we used the technique described by Herculano-Houzel & Lent (2005). The brains of 14-day-old rats (3 NP and 3 LP, from different mothers) were removed and amygdala was dissected. A suspension of nuclei was obtained through mechanical dissociation in a standard solution (40mM sodium citrate and 1% Triton X-100), using a 40-ml glass Tenbroeck tissue homogenizer. After washed several times with dissociation solution and centrifuged (10-min at 4000 g) pelleted nuclei were suspended in phosphate-buffered saline (PBS) containing 1% 4,6-diamidino-2-phenylindole (DAPI, Molecular Probes, Eugene, OR, USA). DAPI-stained nuclei were counted under a fluorescence microscope at 400×objective. The nuclear density in the suspension was determined by averaging over at least eight samples the total number of cells in the original tissue was estimated by multiplying mean nuclear density by total suspension volume. Neuron number was estimated after immunohistochemistry using anti-NeuN antibody (1:300 in PBS; Chemicon Temecula, CA, USA) and CY3 goat anti-mouse secondary antibody (1:400 in 40% PBS, 10% goat serum and 50% DAPI; Accurate Chemicals, Westbury, NY, USA) under the fluorescence microscope (Scabora et al., 2013). Total number of other

amygdala cells nuclei is calculated by subtracting the number of NeuN containing nuclei from the total number of nuclei.

***Neuronal 3D reconstruction and dendritic tree analysis*** - The amygdala from 14 days old offspring (4 NP and 4 LP from different mothers) were Golgi–Cox staining according to a published protocol (Gibb R & Kolb B, 1998). Briefly brains were removed and immersed in Golgi–Cox solution (Glaser & Van der Loos, 1981) for 14 days; brains were then transferred to a 30% sucrose solution (3-days) before being cut on a vibratome. Coronal sections (200  $\mu\text{m}$  thick) were collected in 6% sucrose and blotted dry onto gelatin-coated microscope slides. They were subsequently alkalized in 18.7% ammonia developed in Dektol (Kodak), fixed in Kodak Rapid Fix, dehydrated and cleared in xylene before being mounted and coverslipped. Slides were coded before morphometric analysis in both sets. The basolateral amygdala, strongly associated with fear and anxiety behavior was selected for neuron reconstruction. For dendritic tree analyze the criteria used to select neurons for reconstruction were as follows: (i) full impregnation of the neurons along the entire length of the dendritic tree; (ii) dendrites without significant truncation of branches; (iii) relative isolation from neighboring impregnated neurons to avoid interference with the analysis; and (iv) no morphological changes attributable to incomplete dendritic impregnation of Golgi–Cox stain (Pêgo et al. 2008). For each selected neuron all branches of the dendritic tree were reconstructed at 600x magnification using a motorized microscope (Carl Zeiss Axioplan 2) attached to a camera (DXC-390; Sony Co, Japan) and NeuroLucida software (Micro Bright Field, VT, USA). Three-dimensional analysis of the reconstructed neurons was performed using NeuroExplorer software (MicroBrightField). For the dendritic analysis were reconstructed 91 neurons.

***Neurochemical analyses methods*** - Male rats at 7 and 14 days were decapitated and the heads were snap-frozen in liquid nitrogen. The amygdala was dissected and stored at  $-80\text{ }^{\circ}\text{C}$  until further analysis.

**Western blot (WB)** - The tissue was homogenized in extraction buffer RIPA (Radio Immune Precipitation Assay Buffer) and then 10% triton x 100 and 10% SDS were added to the homogenate. The tissue extracts were centrifuged (1300 rpm at 4°C for 40 min) and the supernatants used as a sample. Protein quantification was performed using the Bradford method. The samples were treated with a Laemmle buffer containing 100-mmol/l dithiothreitol (DTT) heated in a boiling water bath for 4 min and subjected to 8% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) in a Bio-Rad mini-gel apparatus (Mini-Protean Bio-Rad). Electrotransfer of proteins from the gel to the nitrocellulose membranes was performed for 90 min at 120 V (constant) in a Bio-Rad miniature transfer apparatus (Mini-Protean). The non-specific protein binding to the nitrocellulose was reduced by preincubating the filter for 2 h at 22°C in a blocking buffer (BSA 5%, 10mmol/l Tris, 150mmol/l NaCl and 0.02% Tween 20). The nitrocellulose blots were incubated at 4°C overnight with antibodies against GR (H-300-Santa Cruz 8992), MR (H-300-Santa Cruz), BDNF (Abcam; ab46176), 5HT1A (ab101914), 5HT2A (ab160228), CRF (S-19- Santa Cruz 1761), CRF1 (ab 59023) and Alfa tubulin (DSHB; AA4.3) diluted in a blocking buffer (2.5% BSA, 10mmol/l Tris, 150mmol/l NaCl and 0.02% Tween 20). Immunoreactivity bands were detected using the enhanced chemiluminescence method (RPN 2108 ECL Western blotting analysis system; Amersham Biosciences) and were detected by a ChemiDoc XRS system (Biorad; 170870). The band intensities were quantitated by optical densitometry (*TINA* software) of the developed autoradiographs that were used at exposures in the linear range.

**High performance liquid chromatography combined with electrochemical detection (HPLC/CE)** - The level of catecholamine and serotonin in the amygdala was assessed by HPLC/CE using a Gilson instrument (Golson, Middleton, WI, USA) equipped with an analytical column (Supleco Supelcosil LC-18, 3mM, Bellefonte, PA, USA, flow rate: 1.0ml/min). Perchloric acid (200ul) was added to each sample that were incubated for 30 min in ice and then sonicated and centrifuged (13000rpm, 10min, 4°C). Supernatant was collected to a 1.5ml tube

and then centrifuged for 8min (10000rpm, 4°C) and the resulting pellet was saved for later use. The supernatant was then filtered through an HPLC Spin-X column (Costar, Lowell, MA, USA) to remove debris and 150ul aliquots were injected into the HPLC system using a mobile phase of 0.7 M aqueous potassium phosphate (pH 3.0) in 10% methanol 1-heptanesulfonic acid (222mg l<sup>-1</sup>) and Na-EDTA (40mg l<sup>-1</sup>). A standard curve (Sigma H-7752) with known concentrations of each catecholamine was run each day for 5HIAA (Sigma H-8876), Dopamine (Sigma H-8502), DOPAC (Sigma D-9128), HVA (Sigma H-1252), Epinephrine (Sigma E-4375) and Norepinephrine (Sigma 74460).

**Data presentation and statistical analysis** - All data is reported as mean ± SD. Data obtained over time was analyzed using appropriate two-way analysis of variance (two-way ANOVA). Post hoc comparisons between selected means were made by Bonferroni's contrast test when initial two-way ANOVA indicated statistical differences between experimental groups. Comparison involving only two samples of independent observations tends within or between groups was made using a Student's test. The band intensities were quantitated by optical densitometry (*software TINA*). The Tukey–Kramer test for multiple comparisons was used for analysis. The level of significance was set at  $P \leq 0.05$ .

## **Results**

The birthweight of LP offspring was significantly decreased when compared with that obtained in NP group. This reduction was also verified at LP 7 and 14 day-old offspring when compared to age-matched NP rats. The brain, thymus and adrenals weight were similar in both groups, when normalized for body weight (Table 1).

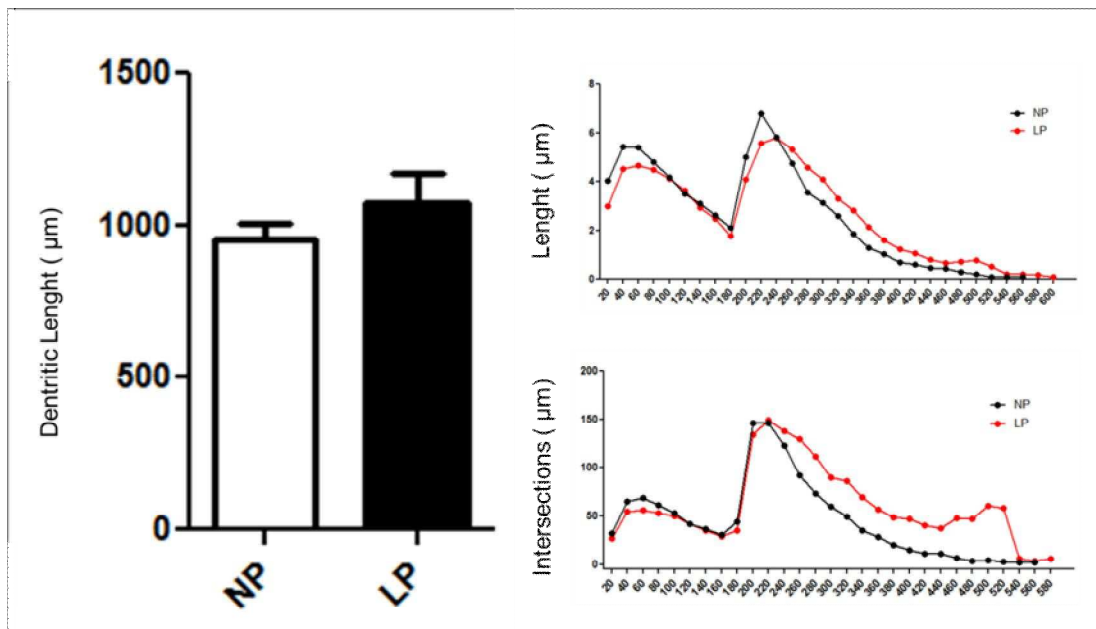
**Table 1.** Body birthweight, brain, thymus and adrenal weight of offspring from NP and LP groups. Mean ± SEM. Statistically difference accepted when  $p < 0.05$ .



	Weight (g)			Organ/body weight (g/g)		
	NP (n=20)	LP (n=20)	p	NP (n=20)	LP (n=20)	p
Birth weight	6.251 ± 0.06074	5.910 ± 0.09273	0.0023**			
Body weight 7 <sup>o</sup> day	16.52 ± 0.2652	14.93 ± 0.3020	0.0002***			
Body weight 14 <sup>o</sup> day	33.64 ± 0.4805	30.12 ± 0.6916	0.0001***			
Brain 7 <sup>o</sup> day	0.7231 ± 0.005922	0.6841 ± 0.008642	0.0004***	14.22 ± 1.747	15.63 ± 1.478	0.5366
Brain 14 <sup>o</sup> day	1.270 ± 0.01091	1.206 ± 0.01471	0.0008***	13.10 ± 2.530	9.318 ± 2.529	0.2979
Adrenal 7 <sup>o</sup> day	0.003396 ± 0.0001811	0.003359 ± 0.0001936	0.8916	0.0002064 ± 0.00001017	0.0002338 ± 0.00001362	0.1211
Adrenal 14 <sup>o</sup> day	0.008333 ± 0.0005236	0.008433 ± 0.0003930	0.8944	0.0002529 ± 0.00001533	0.0002905 ± 0.00001900	0.1413
Thymus 7 <sup>o</sup> day	0.04669 ± 0.002280	0.04754 ± 0.001781	0.7714	0.002816 ± 0.0001341	0.003260 ± 0.0001065	0.0125*
Thymus 14 <sup>o</sup> day	0.1105 ± 0.005273	0.09426 ± 0.01102	0.1484	0.003354 ± 0.0001451	0.003084 ± 0.0002097	0.2889

### Neuronal 3D reconstruction and dendritic tree analysis

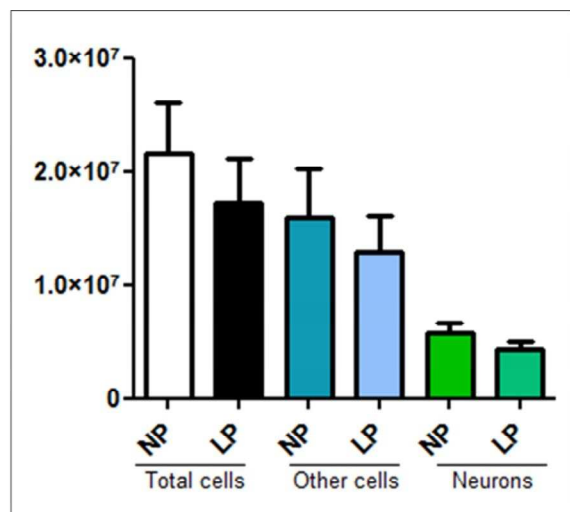
The neuronal 3D dendritic analysis studying dendrites length and dendritic ramifications by Sholl analysis, in neurons from amygdala was not different in 14-d-old NP and LP offspring (Fig. 1).



**Fig 1.** The gestational protein restriction has not effects in the length and ramification of amygdala neuronal dendrites.

**Amygdala total cells and neurons quantification:**

The results show a statistically non-significant reduction of neurons and non-neurons cells number in the amygdala of LP offspring when compared to NP rats (Fig. 2).

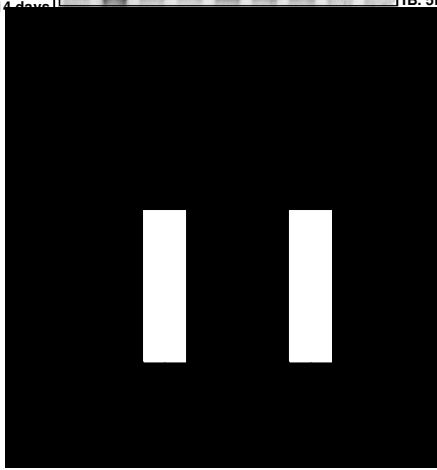


**Fig 2.** Neurons and other cells number in the amygdala of 14 day-old animals.

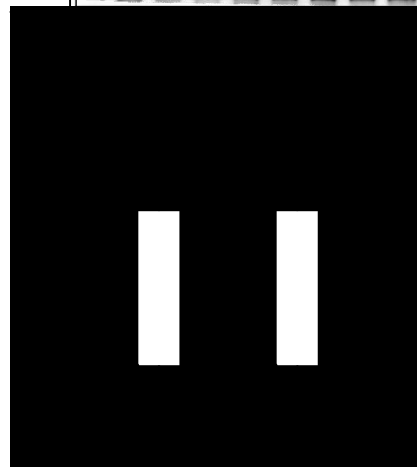
**Western blot analysis**

The western-blotting analyses show a significant reduction of GR expression (24%) in 14 day-old LP when we compared to NP offspring. Also, MR expression was reduced at 7 and 14 day-old LP respectively, 20% ( $p=0.07$ ) and 45% ( $p=0.01$ ) compared to age-matched NP animals. The CRF expression was significantly lower in both 7<sup>th</sup> (36,5%) and 14<sup>th</sup> (69%) day of life in LP group. Additionally, BDNF expression was 12,5% ( $p=0.03$ ) and 25% ( $p=0.08$ ) reduced in LP group respectively at 7 and 14 day-old offspring. The study did not show statistical difference in both experimental groups to 5HT<sub>1A</sub>, 5HT<sub>2A</sub> and CRF1 receptors expression (Fig. 3).

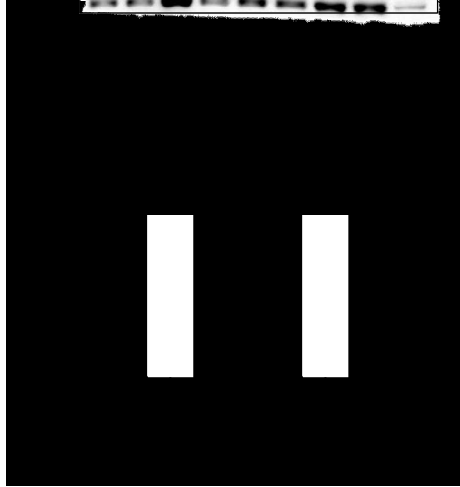
NP NP NP NP NP LP LP LP LP  
 7 days IB: 5HT<sub>1A</sub>  
 IB:  $\alpha$  tubulin  
 14 days IB: 5HT<sub>1A</sub>  
 IB:  $\alpha$  tubulin



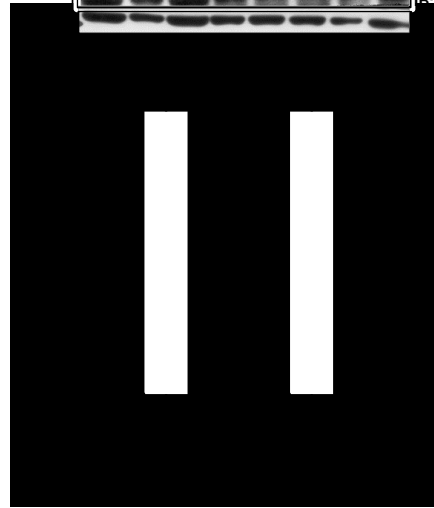
NP NP NP NP NP LP LP LP LP  
 7 days IB: 5HT<sub>2A</sub>  
 IB:  $\alpha$  tubulin  
 IB: 5HT<sub>2A</sub>  
 IB:  $\alpha$  tubulin



NP NP NP NP NP LP LP LP LP  
 7 days IB: M R  
 IB:  $\alpha$  tubulin  
 14 days IB: M R  
 IB:  $\alpha$  tubulin



NP NP NP NP LP LP LP LP  
 7 days IB: GR  
 IB:  $\alpha$  tubulin  
 IB: GR  
 IB:  $\alpha$  tubulin

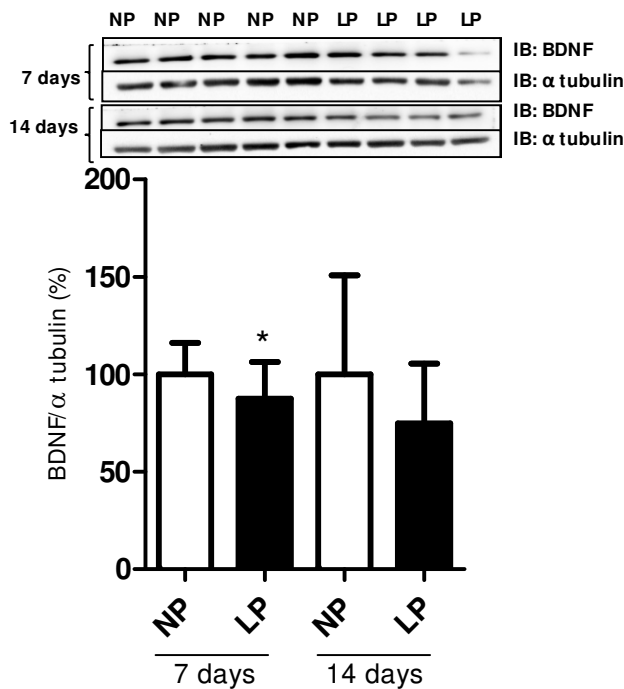


NP NP NP NP LP LP LP LP  
 7 days IB: CRF  
 IB:  $\alpha$  tubulin  
 14 days IB: CRF  
 IB:  $\alpha$  tubulin



NP NP NP NP LP LP LP LP  
 7 days IB: CRF1  
 IB:  $\alpha$  tubulin  
 14 days IB: CRF1  
 IB:  $\alpha$  tubulin





**Fig 3.** Western blot analysis of the 5HT2A, 5HT1A, GR, MR, BDNF, CRF and CRF1 expression in the amygdala of animals from NP and LP groups with either 7 or 14 days old. Results of scanning densitometry are presented as relative to NP, assigning a value of 100% to NP rats. Columns and bars represent the mean  $\pm$  SD. \* $P < 0.05$ , NP versus LP.

### High-Performance Liquid Chromatography (HPLC)

The concentrations of norepinephrine and DOPA, assessed by HPLC were significantly decreased in 7 day-old LP animals when compared to age-matched NP offspring. No differences were found in the level of other neurotransmitters analyzed.

### Table 2: Determination of the concentration of neurotransmitters by HPLC:

Values represent the mean  $\pm$  standard deviation. Statistical comparisons were made with the Student t test and statistical significance was accepted for  $p \leq 0.05$ .

	Amygdala					
	7 days		p	14 days		p
	NP	LP		NP	LP	
Norepinephrine	35.50 $\pm$ 4.41 N=3	17.95 $\pm$ 1.06 N=3	0.009**	44.63 $\pm$ 13.32 N=5	20.91 $\pm$ 4.54 N=4	0.09
Epinephrine	6.53 $\pm$ 0.95 N=3	4.22 $\pm$ 0.42 N=3	0.04*	9.24 $\pm$ 2.12 N=5	7.46 $\pm$ 0.8 N=3	0.28
Dopamine	14.89 $\pm$ 2.69 N=4	7.02 $\pm$ 1.59 N=4	0.02 *	5.21 $\pm$ 1.18 N=4	3.8 $\pm$ 0.75 N=4	0.18
DOPAC	10.39 $\pm$ 1.55 N=4	6.89 $\pm$ 1.83 N=3	0.1	8.2 $\pm$ 2.05 N=5	4.72 $\pm$ 1.16 N=4	0.11
5HIAA	8.9 $\pm$ 2.76 N=3	6.65 $\pm$ 1.73 N=4	0.25	8.94 $\pm$ 2.04 N=5	5.74 $\pm$ 1.71 N=4	0.14
5HT	11.72 $\pm$ 1.25 N=4	11.90 $\pm$ 2.24 N=3	0.47	24.12 $\pm$ 7.8 N=5	9.33 $\pm$ 2.81 N=3	0.11
5HT TURNOVER	0.42 $\pm$ 0.09 N=4	0.61 $\pm$ 0.08 N=3	0.09	0.43 $\pm$ 0.09 N=6	0.35 $\pm$ 0.12 N=4	0.3
DOPAMINE TURNOVER	1.16 $\pm$ 0.1365 N=5	1.25 $\pm$ 0.13 N=4	0.32	1.33 $\pm$ 0.07 N=5	1.2 $\pm$ 0.09 N=4	0.14

## Discussion

In accordance with previous reports, in the current study, we also observe a significant decrease in body birth weight that remains up to 14 day of age, in gestational protein-restricted offspring. Additionally, this study also confirms several previous that have shown that protein restriction intake did not alters the brain weight in adults offspring (Lingas et al, 1999). Recent study in our laboratory has demonstrated that 16 week-old male offspring from gestational protein-restricted dam, also, did not present any changes in the brain mass. In this study, we also do not show significant difference in the brain mass at 7 and 14 day-old LP rats. In this sense, our data add support to the “selfish brain” theory, which proposes that intrauterine adverse events such as emotional stress, either undernutrition, restriction of specific nutrients or hypoxia, may program the brain to maintain stability in its own energy sources. Particularly, alterations in early life, program the developing neuronal substrate to decrease neuroendocrine activity to, in turn, decrease the somatic growing rate, guarantying an adequate nutrition to the developing brain (Lumbers, et al, 2001). This theory can explain the reduced body weight of LP animals without affecting the brain mass.

The present study also evaluated the post-mortem adrenal and thymus weight. At 7 day-old LP offspring the results show an increased thymus weight that not remain in 14 day-old offspring. This thymus change may be related to steroidal plasma levels that could be associated with alteration in lymphoid tissues proliferation of protein-restricted animals. In fact, it has been strongly suggested that neuronal plasticity may be modulated by the immune system (Li et al; 2016). Here surprisingly, the adrenal mass was unchanged in both age-matched groups. This finding, despite fact that stress in adults rats induces an increased adrenal mass (Ventura-Silva et al., 2012), in the current study, it seems that the gestational protein restriction does not affect this organ in the times studied.

In the current study, the amygdala neuronal 3D dendritic analysis of dendrites length and dendritic ramifications by Sholl analysis, not show alteration in 14 d-old NP compared with age-matched gestational protein-restricted offspring. Also, the amygdala neurons and non-neuronal cells number was similar in 14 day-old LP

and NP offspring. It is noteworthy that both dendrites' volume and length was evaluated in specific region on basolateral amygdala while the cells' number was determined on the whole amygdala. The dendritical length and arborization was determined in the basolateral amygdala, once this nucleus is the main responsible to modulate signals coming from several regions of CNS and, furthermore, is strongly associated with fear and anxiety behavior (Izquierdo et al, 2016; Li et al, 2016).

By the way, Pêgo et al., (2008) did not report any morphological alterations in the amygdala in animals that were exposed to chronic unpredictable stress while chronic immobilization stress leads to an hypertrophy of dendrites in this brain structure (Mitra & Sapolsky , 2008). The intensity and duration of stress exposure are therefore, seems to be key factors for the development of morphological and neurochemical alterations in the amygdala (Wilson et al, 2015). Thus, studies showing that chronic immobilization stress increases dendritic length, the number of ramifications and the number of spines in neurons of the basolateral amygdala (Mitra et al, 2005; Vyas et al, 2002). Additionally, these morphological changes are related with anxiety-like behavior in chronic stressed-animals (Vyas & Chattarji, 2004).

It is broadly confirmed that gestational protein restriction may induce fetal programming leading to modification in the brain structure and behavior of offspring that remain up to adulthood (Ashton, 2000; Procter & Campbell, 2014, Barker, 1995; Forsdahl, 1967). Animal studies have shown that early-life undernutrition has a deep impact on fetal tissues and organs development, causing permanent changes in a wide range of morphological and physiological functions, including brain function (Alamy & Bengelloun, 2012). However, the amygdala's functional neurochemical modulation has been poorly documented in gestational protein-restricted offspring, therefore, in the present study we investigate the effects of gestational protein restriction on whole amygdala neurochemical compound in male offspring, in key moments of post-natal neural development. Here, we demonstrate significant decrease in the norepinephrine, epinephrine and dopamine levels, as well as reduction in the expression GR and MR steroid

receptors and CRF in the amygdala in 7 and 14 day-old LP offspring compared to control group.

The amygdala plays a crucial role coordinating behavioral, autonomic, and neuroendocrine stress responses, via mostly excitatory influences on the hypothalamus and brainstem (Aggleton, 2000). It stimulates the HPA axis through indirect projections to the hypothalamic paraventricular nucleus (PVN), including a disinhibitory pathway via the bed nucleus of the stria terminalis (Feldman et al., 1990; Herman et al., 2003, Freese & Amaral, 2009). PVN stimulation releases corticotrophin-releasing factor (CRF) into the portal vasculature, which binds to CRF receptors in the anterior pituitary stimulating the release of adrenocorticotrophic hormone (ACTH), which stimulates the synthesis and release of glucocorticoids by the adrenals (Ulrich-Lai and Herman, 2009). Glucocorticoids inhibit their own release via negative feedback through binding to glucocorticoid receptors (GRs) in the pituitary, PVN, and extrahypothalamic brain regions (Myers et al., 2012). In the present study, the reduced expression of amygdala GR and MR may have a key role on excitatory HPA axis response, priority recorded, in LP offspring.

The amygdala's excitatory influence on the HPA axis stress response has been primarily demonstrated in adult animals, with electrical stimulation increasing secretion of glucocorticoids (Redgate & Fahringer, 1973; Ehle et al., 1977) and lesions resulting in blunted HPA axis stress responses (Kalin et al., 2004; Machado & Bachevalier, 2008). Recent study has demonstrated administration of corticosteroid into amygdala nucleus promotes an anxiety-like behavior (Venkova et al 2010) by MR and GR stimuli (Herman et al 1989). Also, reporters suggested that both receptors are implicated in glucocorticoids fear-enhancing effect (Oitzl et al 2001). In the central amygdala nucleus (CeA) MR and GR are expressed in CRF neurons, suggesting direct glucocorticoid regulation of CRF expression in this nucleus (Honkaniemi et al., 1992). The amygdala is also rich in corticotrophin-releasing factor (CRF), a specific modulator of stress-related anxiety and the role of the amygdala in the modulation of the HPA axis may be through the activation of CRF receptor (CRF1) (Davis, 1992; Merali et al., 2004; Muller et al., 2003).

However, recent data from Raper et al (2014) has demonstrate that early amygdala damage (decreasing GR/MR and CRF amygdala levels) alters the typical development of the primate HPA axis resulting in increased rather than decreased activity, presumably via alterations in central CRF and GR systems in neural structures that control its activity. Thus, in contrast to evidence that the amygdala stimulates both CRF and HPA axis systems in the adult, our data suggest an opposite inhibitory role of the amygdala on the HPA axis during early development, which fits with decreased GR, MR and CRF levels observed in the present study. The CRF receptors activation may also induces alterations in other neurotransmitter systems such as serotonin (5HT), dopamine and norepinephrine (Millan, 2005; Holsboer, 2003; Nestler et al., 2002; Leonard, 2005; Holmes et al, 2003) and the amygdala CRF reduction in protein-restricted offspring, may affect this neurotransmitter release. In addition, they may be interacting with brain derived neurotrophic factor (BDNF), network deeply involved in fear and anxiety (Hammack et al, 2009; Duman, 2004; Kumari, et al 2016).

In the current work, the animals submitted to gestational nutritional restrictions presented a decrease in amygdala norepinephrine, epinephrine and dopamine levels. It is well known that stress exposure leads to a cascade of neuroendocrine response that promotes release of corticosteroids and catecholamines (Wolf, 2015). Nevertheless, authors report a decrease in dopaminergic and/or noradrenergic release in the depression pathophysiology (Guiard et al, 2008). *In vitro* studies suggest that the excitability of neurons in the amygdala is modulated by dopamine (Kroner et al, 2005) and that there is a strong relation between catecholamines and HPA axis alterations (Leonard, 2001). On the other hand, norepinephrine activates CRF, which, in turn, can induce the release of norepinephrine, leading to a cycle (Koob, 1999). This modulatory system may be blunted in the current study. These results may be explained by well-characterized stress hyporesponsive period (SHRP) associated to decreased activity of HPA axis, that may last until the 14<sup>th</sup> postnatal-day (Sapolsky & Meane 1986). At this time, there is a reduced secretion of corticosterone, which remains low until the second postnatal week (Sapolsky & Meane 1986). The SHRP may assume as a



protective mechanism ensuring low levels of glucocorticoids during early postnatal development (Sapolsky et al, 1986; Mesquita et al, 2007). The decrease in the expression of catecholamines in gestational protein-restricted model, may be one of mechanisms that contribute to the lower activity of the HPA axis during this period of life.

In conclusion, the amygdala's role on behavioral, autonomic, and neuroendocrine stress responses has been poorly studied in programmed models, with scarce studies reporting the morphological and neurochemistry characteristics and the relationship with the key role of the amygdala in the stress response (Herman & Cullinan 1997; Van de Kar & Blair, 1999). The amygdala neurochemical changes observed in this study may contribute to behavioral alterations induced by gestational protein restriction and may also be a primer for alterations in other brain regions. Also, these findings may represent the adaptation during embryonic development to exposure to elevated maternal corticosteroids as a consequence of nutritional stress. Additional studies must be done, but the present study suggests strongly that amygdala morphological and neurochemical disorders may be associated with the development of psychiatric disorders in adulthood.

## REFERENCES

- AGGLETON JP. The amygdala. A functional analysis. (2000) Ed 2. New York: **Oxford Universit Press.**
- ALAMY M; BENGELLOUN WA. Malnutrition and brain development: an analysis of the effects of inadequate diet during different stages of life in rat. **Neurosci Biobehav Rev.** 2012 Jul;36(6):1463-80. doi: 10.1016/j.neubiorev.2012.03.009. Epub 2012 Apr 3.
- ASHTON, N. Perinatal development and adult blood pressure. **Braz J Med Biol Res** 2000; 33:731-40.

BARKER, D.J. The fetal and infant origins of disease. **Eur J Clin Invest.** 1995; 25:457-63.

BORGES S; COIMBRA B; SOARES-CUNHA C; VENTURA-SILVA AP; PINTO L; CARVALHO MM; PÊGO JM; RODRIGUES AJ; SOUSA N. Glucocorticoid programming of the mesopontine cholinergic system. **Front Endocrinol (Lausanne).** 2013 Dec 13;4:190. doi: 10.3389/fendo.2013.00190. eCollection 2013.

BURD, L.; SEVERUD, R.; KERBESHIAN, J.; KLUG, M.G. Prenatal and perinatal risk factors for autism. **J Perinat Med** 1999; 27: 441-450.

DAVIS, M. The role of the amygdala in fear and anxiety. **Ann. Rev. Neurosci.** 1992; 15:353–375.

DING, Y X; SHI, Y HAN, W J; CUI, H. Regulation of glucocorticoid related genes and receptors/regulatory enzyme expression in intrauterine growth restriction filial rats. **Life Sci.** 2016 Feb 23. pii: S0024-3205(16)30128-X. doi: 10.1016/j.lfs.2016.02.079. [Epub ahead of print].

DUMAN RS. Role of neurotrophic factors in the etiology and treatment of mood disorders. **Neuromolecular Med.** 2004;5(1):11-25.

EHLE AL, MASON JW, PENNINGTON LL Plasma growth hormone and cortisol changes following limbic stimulation in conscious monkeys. **Neuroendocrinology** (1977) 23:52– 60.

FELDMAN S, CONFORTI N, SAPHIERD. The preoptic area and bed nucleus of the stria terminalis are involved in the effects of the amygdala on adrenocortical secretion. **Neuroscience** (1990) 37:775–779.

FORSDAHL, S. Are poor living conditions in childhood and adolescence an important risk factor for arteriosclerotic heart disease? **British journal of Preventive and Social Medicine,** 1967; 31: 91-95.

FREESE JL, AMARAL DG Neuroanatomy of the primate amygdala. In: The human amygdala (Whalen PJ, Phelps EA, eds), (2009) pp 1–42. New York: Guilford. Furay AR, Bruest

GIBB, R.; KOLB, B. A method for vibratome sectioning of Golgi-Cox stained whole rat brain. **J Neurosci Methods** 1998; 79: 1–4.

GLASER, E.M.; VAN, D.E.R.; LOOS, H. Analysis of thick brain sections by obverse-reverse computer microscopy: application of a new, high clarity Golgi-Nissl stain. **J Neurosci Methods** 1981; 4:117–125.

GUIARD BP; EL MANSARI M; BLIER P. Cross-talk between dopaminergic and noradrenergic systems in the rat ventral tegmental area, locus ceruleus, and dorsal hippocampus. **Mol Pharmacol.** 2008 Nov;74(5):1463-75. doi: 10.1124/mol.108.048033. Epub 2008 Aug 14.

HAMMACK, S.E.; ROMAN, C.W.; LEZAK, K.R.; KOCHO-SHELLENBERG, M. GRIMMIG, B.; FALLS, W.A.; BRAAS, K.; MAY, V. Roles for pituitary adenylate cyclase-activating peptide (PACAP) expression and signaling in the bed nucleus of the stria terminalis (BNST) in mediating the behavioral consequences of chronic stress. **J Mol Neurosci.** 2010;42(3):327-40. Epub 2010 Apr 20.

HAMMACK, S.E.; GUO, J.D.; HAZRA, R.; DABROWSKA, J.; MYERS, K.M.; RAINNIE, D.G. The response of neurons in the bed nucleus of the stria terminalis to serotonin: implications for anxiety. **Prog Neuropsychopharmacol Biol Psychiatry.** 2009; 13;33(8):1309-20. Epub 2009 May 23.

HERCULANO-HOUZEL S; LENT R. Isotropic fractionator: a simple, rapid method for the quantification of total cell and neuron numbers in the brain. **J Neurosci.** 2005 Mar 9;25(10):2518-21.

HERMAN JP, CULLINAN WE. Neurocircuitry of stress: central control of the hypothalamo–pituitary–adrenocortical axis. **Trends Neurosci** 1997; **20**: 78–84.

HERMAN JP, FIGUEIREDO H, MUELLER NK, ULRICH-LAI Y, OSTRANDER MM, CHOI DC, CULLINAN WE Central mechanisms of stress integration: hierarchical circuitry controlling hypothalamo-pituitary-adrenocortical responsiveness. *Front Neuroendocrinol* (2003)24:151–180.

HERMAN JP, PATEL PD, AKIL H, WATSON SJ. Localization and regulation of glucocorticoid and mineralocorticoid receptor messenger RNAs in the hippocampal formation of the rat. *Mol Endocrinol* 1989, 3:1886–1894.

HOLMES, A.; HEILIG, M.; RUPNIAK, N.M.; STECKLER, T.; GRIEBEL, G. Neuropeptide systems as novel therapeutic targets for depression and anxiety disorders. *Trends Pharmacol. Sci.* 2003; 24:580–588.

HOLSBOER, F. Corticotrophin-releasing hormone modulators and depression. *Curr. Opin. Investig. Drugs.* 2003; 4:46–50.

HONKANIEMI J., M. PELTO-HUIKKO, L. RECHARDT, J. ISOLA, A. LAMMI, K. FUXE, J. GUSTAFSSON, A. WIKSTROM, T. HOKFELT. Colocalization of peptide and glucocorticoid receptor immunoreactivities in rat central amygdaloid nucleus, *Neuroendocrinology* 55 1992 451–459.

IZQUIERDO I, FURINI CR, MYSKIW JC. Fear Memory. *Physiol Rev.* 2016 Apr;96(2):695-750. doi: 10.1152/physrev.00018.2015.

JOTHIE RICHARD E; ILLURI R; BETHAPUDI B; ANANDHAKUMAR S; BHASKAR A; CHINAMPUDUR VELUSAMI C MUNDKI; NAJEDDU D; AGARWAL A. Anti-stress Activity of *Ocimum sanctum*: Possible Effects on Hypothalamic-Pituitary-Adrenal Axis. *Phytother Res.* 2016 Feb 22. doi: 10.1002/ptr.5584. [Epub ahead of print]

KALIN NH, SHELTON SE, DAVIDSON RJ The role of the central nucleus of the amygdala in mediating fear and anxiety in the primate. *J Neurosci* (2004) 24:5506–5515.

KOOB GF. Corticotropin-releasing factor, norepinephrine, and stress. **Biological Psychiatry** Volume 46, Issue 9, 1 November 1999, Pages 1167–1180.

KRÖNER S, ROSENKRANZ JA, GRACE AA, BARRIONUEVO G. Dopamine modulates excitability of basolateral amygdala neurons in vitro. **J Neurophysiol**. 2005 Mar;93(3):1598-610. Epub 2004 Nov 10.

KRÖNER S, ROSENKRANZ JA, GRACE AA, BARRIONUEVO G. Dopamine modulates excitability of basolateral amygdala neurons in vitro. **J Neurophysiol**. 2005 Mar;93(3):1598-610. Epub 2004 Nov 10.

KUMARI A; SINGH P; BAGHEL MS; THAKUR MK. Social isolation mediated anxiety like behavior is associated with enhanced expression and regulation of BDNF in the female mouse brain. **Physiol Behav**. 2016 Feb 23;158:34-42. doi: 10.1016/j.physbeh.2016.02.032. [Epub ahead of print]

LEONARD BE. Stress, norepinephrine and depression. **J Psychiatry Neurosci**. 2001;26 Suppl:S11-6.

LEONARD BE. The HPA and immune axes in stress: the involvement of the serotonergic system. **Eur Psychiatry**. 2005; 20:302–306.

LI M; GUO K; ADACHI Y; IKEHARA S. Immune Dysfunction Associated with Abnormal Bone Marrow-Derived Mesenchymal Stroma Cells in Senescence Accelerated Mice. **Int J Mol Sci**. 2016 Jan 29;17(2). pii: E183. doi: 10.3390/ijms17020183.

LINGAS R; DEAN F; MATTHEWS SG. Maternal nutrient restriction (48 h) modifies brain corticosteroid receptor expression and endocrine function in the fetal guinea pig. **Brain Res**. 1999 Nov 6;846(2):236-42.

LOPES A; TORRES DB; RODRIGUES AJ; CERQUEIRA JJ; PÊGO JM; SOUSA N; GONTIJO JA; BOER PA. Gestational protein restriction induces CA3 dendritic atrophy in dorsal hippocampal neurons but does not alter learning and memory

performance in adult offspring. **Int J Dev Neurosci.** 2013 May;31(3):151-6. doi: 10.1016/j.ijdevneu.2012.12.003. Epub 2012 Dec 30.

LUCAS A. Programming by early nutrition in man. **Ciba Found Symp.** 1991;156:38-50; discussion 50-5.

LUMBERS ER, YU ZY, GIBSON KJ. The selfish brain and the barker hypothesis. **Clin Exp Pharmacol Physiol.** 2001 Nov;28(11):942-7.

LUMBERS ER; YU ZY; GIBSON KJ. THE SELFISH BRAIN AND THE BARKER HYPOTHESIS. **Clin exp pharmacol physiol.** 2001 NOV;28(11):942-7.

MACHADO CJ, BACHEVALIER J Behavioral and hormonal reactivity to threat: effects of selective amygdala, hippocampal, or orbital frontal lesions in monkeys. **Psychoneuroendocrinology** (2008) 33:926–941.

MERALI Z, KHAN S, MICHAUD, D.S.; SHINOSITOL, PHOSPHATEPY SA, ANISMAN H. Does amygdaloid corticotrophin-releasing hormone (CRF) mediate anxiety-like behaviors? Dissociation of anxiogenic effects and CRF release. **Eur. J. Neurosci.** 2004; 20:229–239.

MESQUITA FF, GONTIJO JAR, BOER PA. Maternal undernutrition and the offspring kidney: from fetal to adult life. **Braz J Med Biol Res.** 2010 Nov;43(11):1010-8. Epub 2010 Oct 29.

MILLAN, M.J. Serotonin 5-HT<sub>2C</sub> receptors as a target for the treatment of depressive and anxious states: focus on novel therapeutic strategies. **Therapies.** 2005; 60:441–460.

MITRA R, JADHAV S, MCEWEN BS, VYAS A, CHATTARJI S. Stress duration modulates the spatiotemporal patterns of spine formation in the basolateral amygdala. **Proc Natl Acad Sci USA.** 2005;102:9371–9376.[

MITRA R, SAPOLSKY RM. Acute corticosterone treatment is sufficient to induce anxiety and amygdaloid dend

itic hypertrophy. **Proc Natl Acad Sci U S A**. 2008 Apr 8;105(14):5573-8. doi: 10.1073/pnas.0705615105. Epub 2008 Apr 7.

MULLER, M.B. Limbic corticotrophin-releasing hormone receptor 1 mediates anxiety-related behavior and hormonal adaptation to stress. **Nat. Neurosci**. 2003; 6:1100–1107.

MYERS B, MCKLVEEN JM, HERMAN JP Neural regulation of the stress response: the many faces of feedback. **Cell Mol Neurobiol** (2012) 32:683– 694.

NESTLER, E.J.; BARROT, M.; DILEONE, R.J.; EISCH, A.J.; GOLD, S.J.; MONTEGGIA, L.M. Neurobiology of depression. **Neuron**. 2002; 34:13–25.

OITZL MS, REICHARDT HM, JOELS M, DE KLOET ER. Point mutation in the mouse glucocorticoid receptor preventing DNA binding impairs spatial memory. **Proc Natl Acad Sci USA** 2001, 98:12790–12795.

PÊGO, J. M.; MORGADO, P.L.G.; CERQUEIRA, J.J., ALMEIDA, O.F.X.; SOUSA, N. Dissociation of the morphological correlates of stress-induced anxiety and fear. **European Journal of Neuroscience** 2008; 27:1503–1516.

PROCTER, S B.; Campbell CG. Position of the Academy of Nutrition and Dietetics: Nutrition and Lifestyle for a Healthy Pregnancy Outcome. **J Acad Nutr Diet**. 2014; 114(7):1099-103. doi: 10.1016/j.jand.2014.05.005.

RAPER J, SHANNON B.Z. STEPHENS, AMY HENRY, TRINA VILLARREAL, JOCELYNE BACHEVALIER, KIM WALLEN AND MAR M. SANCHEZ. Neonatal Amygdala Lesions Lead to Increased Activity of Brain CRF Systems and Hypothalamic-Pituitary-Adrenal Axis of Juvenile Rhesus Monkeys **The Journal of Neuroscience** 2014 • 34(34):11452–11460

REDGATE ES, FAHRINGER EE A comparison of the pituitary adrenal activity elicited by electrical stimulation of preoptic, amygdaloid and hypothalamic sites in the rat brain. **Neuroendocrinology** (1973) 12:334 –343.

SCABORA JE, LIMA MC, LOPES A, LIMA IP, MESQUITA FF, TORRES DB, BOER PA, GONTIJO JAR. Impact of taurine supplementation on blood pressure in gestational proteinrestricted offspring: Effect on the medial solitary tract nucleus cell numbers, angiotensin receptors, and renal sodium handling. **J Renin Angiotensin Aldosterone Syst.** 2015 Mar;16(1):47-58. doi: 10.1177/1470320313481255. Epub 2013 Mar 6.

SHOLL, D.A. The measurable parameters of the cerebral cortex and their significance in its organization. **Prog Neurobiol** . 1956; 2:324–333.

SOUSA N; CERQUEIRA JJ; ALMEIDA OF. Corticosteroid receptors and neuroplasticity. **Brain Res Rev.** 2008 Mar;57(2):561-70. Epub 2007 Jul 17.

ULRICH-LAI YM, HERMAN JP Neural regulation of endocrine and autonomic stress responses. **Nat Rev Neurosci** (2009) 10:397– 409. 2009 Jun;10(6):397-409. doi: 10.1038/nrn2647.

UYLINGS, H.B.M.; VAN, P.J. Measures for quantifying dendritic arborizations. **Network: comput neural syst** 2002 13:397–414

VAN DE KAR LD, BLAIR ML. Forebrain pathways mediating stress-induced hormone secretion. **Front Neuroendocrinol.** 1999 Jan;20(1):1-48.

VENKOVA K, ANTHONY C, MYERS JB, GREENWOOD-VAN MEERVELD B. Exposure of the amygdala to elevated levels of corticosterone alters colonic motility in response to acute psychological stress. **Neuropharmacology** 2010;58:1161–1167.

VENTURA-SILVA AP; PÊGO JM; SOUSA JC; MARQUES AR; RODRIGUES AJ; MARQUES F; CERQUEIRA JJ; ALMEIDA OF; SOUSA N. Stress shifts the response of the bed nucleus of the stria terminalis to an anxiogenic mode. **Eur J Neurosci.** 2012 Nov;36(10):3396-406. doi: 10.1111/j.1460-9568.2012.08262.x. Epub 2012 Aug 29.



VYAS A, CHATTARJI S. Modulation of different states of anxiety-like behavior by chronic stress. **Behav Neurosci.** 2004;118:1450–1454.

VYAS A, MITRA R, SHANKARANARAYANA RAO BS, CHATTARJI S. Chronic stress induces contrasting patterns of dendritic remodeling in hippocampal and amygdaloid neurons. **J Neurosci.** 2002;22:6810–6818.

WELBERG, L.A.; SECKL, J.R. Prenatal stress, glucocorticoids and the programming of the brain. **J Neuroendocrinol** 2001; 13: 113-128.

WILSON MA; GRILLO CA; FADEL JR; REAGAN LP. Stress as a one-armed bandit: Differential effects of stress paradigms on the morphology, neurochemistry and behavior in the rodent amygdala. **Neurobiol Stress.** 2015 Jun 9;1:195-208. doi: 10.1016/j.ynstr.2015.06.001. eCollection 2015.

WOLF OT, ATSAK P, DE QUERVAIN DJ, ROOZENDAAL B, WINGENFELD K. Stress and memory: A selective review on recent developments in the understanding of stress hormone effects on memory and their clinical relevance. **J Neuroendocrinol.** 2015 Dec 28. doi: 10.1111/jne.12353. [Epub ahead of print]

YANG YL; CHAO PK; LU KT. Systemic and intra-amygdala administration of glucocorticoid agonist and antagonist modulate extinction of conditioned fear. **Neuropsychopharmacology.** 2006 May;31(5):912-24.

## 5. CONSIDERAÇÕES GERAIS

A amígdala estimula o eixo HPA através de projeções indiretas para o PVN, incluindo uma via excitatória através do BNST (Feldman et al, 1990; Herman et al, 2003 Freese & Amaral, 2009). O estímulo ao PVN promove a liberação de CRF nos vasos portais, que se liga aos receptores de CRF da hipófise estimulando a secreção de ACTH, que por sua vez estimula a síntese e liberação de GC pelas glândulas suprarrenais (Ulrich-Lai e Herman, 2009). Os GC inibem a sua própria liberação através de alças de retroalimentação negativas, pela ligação a GR na hipófise, no PVN e em regiões extra-hipotalâmicas do cérebro, tais como hipocampo e núcleos da amígdala (Myers et al., 2012).

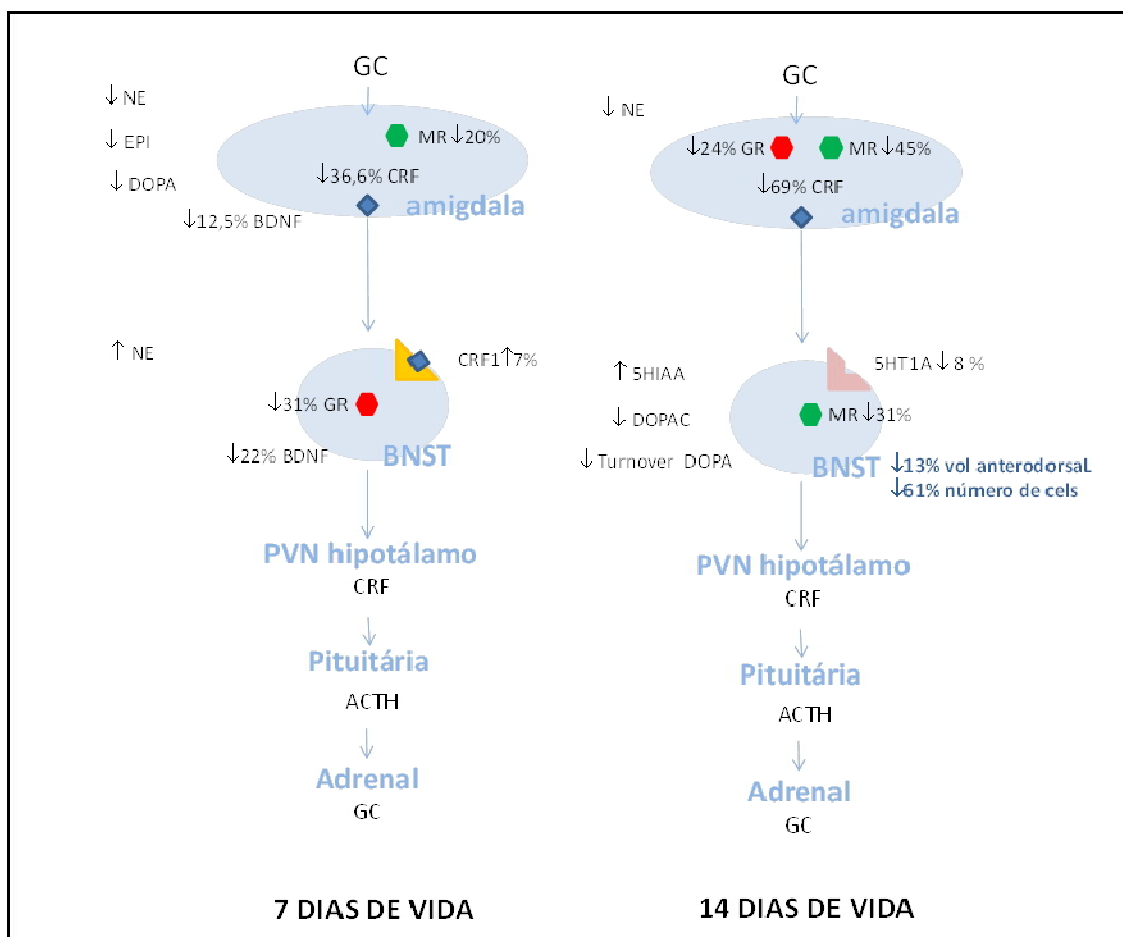
A influência excitatória da amígdala sobre a resposta do eixo HPA foi demonstrada primeiramente em animais adultos, pela estimulação eléctrica desta causando um aumento na secreção de GC (Mason, 1959; Redgate & Fahringer, 1973; Ehle et al, 1977) e, através de lesões eletrolíticas em núcleos amigdaloides que resultaram na redução na atividade do eixo HPA associada a um estado de estresse (Beaulieu et al, 1986; Feldman et al., 1994; Kalin et al, 2004).

Estudos em roedores mostram que a amígdala encontra-se funcionalmente em estado latente antes do desmame, quando os filhotes são mais dependentes de cuidados maternos, e acredita-se que este estado seja para inibir reações de medo da mãe. Posteriormente, ocorre a ativação dos núcleos da amígdala para que seja possível a ocorrência de reações de medo/defesa coincidindo com o aumento dos níveis séricos e liquorícos de corticosterona durante fase de transição do desenvolvimento (pós-natal imediato) para fase de independência do filhote, após 16º dia de vida (Rincón-Cortés & Sullivan, 2014).

Raper e colaboradores (2014) sugerem que durante o desenvolvimento, a amígdala tem influência inibitória na atividade basal do eixo HPA, tanto em primatas quanto em roedores, mudando mais tarde, para um estado excitatório na adolescência e idade adulta. Várias doenças psiquiátricas (p.e. autismo, esquizofrenia dentre outras) envolvem alterações no desenvolvimento da amígdala

e desregulação do controle da síntese e dos níveis líquóricos e séricos de CRF e de suas ações sobre o eixo HPA (Schumann et al., 2011; Tottenham, 2014)

Desta forma, a redução do CRF (36,6 e 69% respectivamente no 7º e 14º dia de vida) observada e, dos receptores de GC na amígdala e BNST de animais oriundos de mães submetidas á restrição proteica gestacional, pode atenuar a ação inibitória destas estruturas sobre o eixo HPA (Figura 1). Assim, nossos resultados confirmam prévios estudos mostrando que modificações morfológicas e/ou neuroquímicas da amígdala durante o desenvolvimento podem comprometer a ação destes núcleos cerebrais na modulação de respostas ao estresse e ao medo (Rincón-Cortés & Sullivan, 2014; Raper et al, 2014).



**Figura 2.** Compilação das alterações neuroquímicas e estruturais observadas em animais do grupo LP.

## 6. CONCLUSÃO

Os resultados do presente estudo mostram que a restrição proteica gestacional leva a alterações neuroquímicas na amígdala e neuroquímicas e morfológicas no BNST do sétimo ao décimo quarto dias de vida da prole. Estes achados sugerem a interferências do estresse nutricional durante a gestação, sobre o desenvolvimento de vias de sinalização dos glicocorticoides e do CRF que implica diferentes estruturas do sistema nervoso central resultando em hiperatividade destas e, conseqüentemente do eixo HPA, associada à elevação do corticosteroide plasmático e do estado de ansiedade, já observado na idade adulta nestes animais.

## 7. REFERÊNCIAS BIBLIOGRÁFICAS

ALHEID, G.F.; BELTRAMINO, C.A.; OLMOS J.S.; HEIMER, L. Supracapsular portions of the rat extended amygdala, **European Neuroscience Meeting Amsterdam** 1995; 104.

ALHEID, G.F.; BETRAMINO, C.A.; OLMOS, J.S.; FORBES, M.S.; SWANSON D.J.; HEIMER, L. The neuronal organization of the supracapsular part of the stria terminalis in the rat: the dorsal component of the extended amygdale. **Neuroscience** 1998; 84:967-996.

ALHEID, G.F.; HEIMER, L. New perspectives in basal forebrain organization of special relevance for neuropsychiatric disorders: the striatopallidal, amygdaloid, and corticopetal components of substantia innominat. **Neuroscience**. 1988; 27(1):1-39.

ASHTON, N. Perinatal development and adult blood pressure. **Braz J Med Biol Res** 2000; 33:731-40.

BARKER, D.J. In utero programming of chronic disease. **Clin Sci (Lond)**. 1998; 95:115-28.

BARKER, D.J. Intra-uterine programming of the adult cardiovascular system. **Curr Opin Nephrol Hypertens**. 1997;6:106-10.

BARKER, D.J. The fetal and infant origins of disease. **Eur J Clin Invest**. 1995; 25:457-63.

BARKER, D.J.P.; OSMOND, C.; GOLDING, J.; KUH, D.; WADSWORTH, M.E.J. Growth in utero, blood pressure in childhood and adult life, and mortality from cardiovascular disease. **BMJ**. 1989; 298:564-567.

BARKER, D.J.P.; OSMOND, C.; RODIN, I.; FALL, C.H.D.; WINTER, P.D. Low weight gain in infancy and suicide in adult life. **Br Med J** 1995; 311: 1203.

BEAULIEU S, DI PAOLO T, BARDEN N. Control of ACTH secretion by the central nucleus of the amygdala: implication of the serotonergic system and its relevance to the glucocorticoid delayed negative feedback mechanism. **Neuroendocrinology**. 1986;44(2):247-54.

BENEDIKTSSON, R.; LINDSAY, R.S.; NOBLE, J.; SECKL, J.R.; EDWARDS, C.R. Glucocorticoid exposure in utero: new model for adult hypertension. **Lancet**. 1993; 34: 339-41

BURD, L.; SEVERUD, R.; KERBESHIAN, J.; KLUG, M.G. Prenatal and perinatal risk factors for autism. **J Perinat Med** 1999; 27: 441-450.

COROMINAS, M.; RONCERO, C.; CASAS, M. Corticotropin releasing factor and neuroplasticity in cocaine addiction. **Life Sci**. 2010; 86: 1–9.

CULLINAN, W.E.; HERMAN, J.P.; WATSON, S.J. Ventral subicular interaction with the hypothalamic paraventricular nucleus: evidence for a relay in the bed nucleus of the stria terminalis. **J. Comp. Neurol**. 1993; 332:1 –20.

DAVIS, M. The role of the amygdala in fear and anxiety. **Ann. Rev. Neurosci**. 1992a; 15:353–375.

DAVIS, M.; WALKER, D.L.; MILES, L.; GRILLON, C. Phasic vs sustained fear in rats and humans: role of the extended amygdala in fear vs anxiety. **Neuropsychopharmacology**. 2010; 35(1):105-35.

DING, Y X; SHI, Y HAN, W J; CUI, H. Regulation of glucocorticoid related genes and receptors/regulatory enzyme expression in intrauterine growth restriction filial rats. **Life Sci**. 2016 Feb 23. pii: S0024-3205(16)30128-X. doi: 10.1016/j.lfs.2016.02.079. [Epub ahead of print].

DONG, H.W.; PETROVICH, G.D.; WATTS, A.G.; SWANSON, L.W. Basic organization of projections from the oval and fusiform nuclei of the bed nuclei of the stria terminalis in adult rat brain. **J. Comp. Neurol**. 2001 436, 430–455.

FELDMAN S, CONFORTI N, ITZIK A, WEIDENFELD J. Differential effect of amygdaloid lesions on CRF-41, ACTH and corticosterone responses following neural stimuli. **Brain Res**. 1994 Sep 26;658(1-2):21-6.

FELDMAN, S.; CONFORTI, N.; SAPHIER, D. The preoptic area and bed nucleus of the stria terminalis are involved in the effects of the amygdale on adrenocortical secretion. **Neuroscience** 1990; 37:775– 779.

FONTENOT, M.B.; KAPLAN, J.R.; MANUCK, S.B.; ARANGO, V.; MANN, J.J. Long-term effects of chronic social stress on serotonergic indices in the prefrontal cortex of adult male cynomolgus macaques. **Brain Res**. 1995; 705:105-108.

FORSDAHL, S. Are poor living conditions in childhood and adolescence an important risk factor for arteriosclerotic heart disease? **British journal of Preventive and Social Medicine**, 1967; 31: 91-95.

GENRO JP; KIELING C; ROHDE LA; HUTZ MH. Attention-deficit/hyperactivity disorder and the dopaminergic hypotheses. **Expert Rev Neurother**. 2010 Apr;10(4):587-601. doi: 10.1586/ern.10.17.

GIBB, R.; KOLB, B. A method for vibratome sectioning of Golgi-Cox stained whole rat brain. **J Neurosci Methods** 1998; 79: 1–4.

GLASER, E.M.; VAN, D.E.R.; LOOS, H. Analysis of thick brain sections by obverse-reverse computer microscopy: application of a new, high clarity Golgi-Nissl stain. **J Neurosci Methods** 1981; 4:117–125.

GODFREY, K.; ROBINSON, S.; BARKER, D.J, OSMOND C, COX V. Maternal nutrition in early and late pregnancy in relation to placental and fetal growth. **BMJ**. 1996;17;312 (7028):410-4.

GOMEZ-SANCHEZ, E.P. Mineralocorticoid receptors in the brain and cardiovascular regulation: minority rule? **Trends Endocrinol Metab**. 2011 May;22(5):179-87. Epub 2011 Mar 21.

GRAEFF, F.G.; GUIMARÃES, F.S.; DE ANDRADE, T.G.; DEAKIN, J.F. Role of 5-HT in stress, anxiety, and depression. **Pharmacol Biochem Behav**. 1996; 54(1):129-41.

GRAY, T.S.; PIECHOWSKI, R.A.; YRACHETA, J.M.; RITTENHOUSE, P.A, BETHEA, C.L.; VAN.; DE KAR, L.D. Ibotenic acid lesions in the bed nucleus of the stria terminalis attenuate conditioned stress-induced increases in prolactin, ACTH and corticosterone. **Neuroendocrinology** 1993; 57:517–524.

HALES CN, BARKER DJP. Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis. **International Journal of Epidemiology** 2013;42:1215–1222.

HAMMACK, S.E.; ROMAN, C.W.; LEZAK, K.R.; KOCHO-SHELLENBERG, M. GRIMMIG, B.; FALLS, W.A.; BRAAS, K.; MAY, V. Roles for pituitary adenylate cyclase-activating peptide (PACAP) expression and signaling in the bed nucleus of the stria terminalis (BNST) in mediating the behavioral consequences of chronic stress. **J Mol Neurosci**. 2010;42(3):327-40. Epub 2010 Apr 20.

HAMMACK, SE; ROMAN, CW; LEZAK, KR; KOCHO-SHELLENBERG, M; GRIMMIG, B, FALLS, WA; BRAAS, K; MAY, V. Roles for pituitary adenylate cyclase activating peptide (PACAP) expression and signaling in the bed nucleus of the stria terminalis (BNST) in mediating the behavioral consequences of chronic stress. **J Mol Neurosci**. 2010 Nov;42(3):327-40. doi: 10.1007/s12031-010-9364-7. Epub 2010 Apr 20.

HAMMACK, S.E.; GUO, J.D.; HAZRA, R.; DABROWSKA, J.; MYERS, K.M.; RAINNIE, D.G. The response of neurons in the bed nucleus of the stria terminalis to serotonin: implications for anxiety. **Prog Neuropsychopharmacol Biol Psychiatry**. 2009; 13;33(8):1309-20. Epub 2009 May 23.

HANDLEY, S.L. 5-Hydroxytryptamine pathways in anxiety and its treatment. **Pharmacol Ther** 1995; 66:103–48.

HANDLEY, S.L.; MCBLANE, J.W.; CRITCHLEY, M.A.; NJUNG'E, K. Multiple serotonin mechanisms in animal models of anxiety: environmental, emotional and cognitive factors. **Behav Brain Res**. 1993; 20;58(1-2):203-10.

HATALSKI, C.G.; GUIRGUIS, C.; BARAM, T.Z. Corticotropin releasing factor mRNA expression in the hypothalamic paraventricular nucleus and the central nucleus of the amygdala is modulated by repeated acute stress in the immature rat. **J Neuroendocrinol** 1998; 10: 663-669.

HERCULANO-HOUZEL S; LENT R. Isotropic fractionator: a simple, rapid method for the quantification of total cell and neuron numbers in the brain. **J Neurosci**. 2005 Mar 9;25(10):2518-21.

HERMAN JP, FIGUEIREDO H, MUELLER NK, ULRICH-LAI Y, OSTRANDER MM, CHOI DC, CULLINAN WE.  
Central mechanisms of stress integration: hierarchical circuitry controlling hypothal



amo-pituitary-adrenocorticalresponsiveness. **Front Neuroendocrinol.** 2003 Jul;24(3):151-80.

HERMAN, J.P.; CULLINAN, W.E.; WATSON, S.J. Involvement of the bed nucleus of the stria terminalis in tonic regulation of paraventricular hypothalamic CRH and AVP mRNA expression. **J. Neuroendocrinol.** 1994; 6:433– 442.

HOLMES, A.; HEILIG, M.; RUPNIAK, N.M.; STECKLER, T.; GRIEBEL, G. Neuropeptide systems as novel therapeutic targets for depression and anxiety disorders. **Trends Pharmacol. Sci.** 2003; 24:580–588.

HOLSBOER, F. Corticotropin-releasing hormone modulators and depression. **Curr. Opin. Investig. Drugs.** 2003; 4:46–50.

HOWES OD; KAPUR S. The dopamine hypothesis of schizophrenia: version III-- the final common pathway. **Schizophr Bull.** 2009 May;35(3):549-62. doi: 10.1093/schbul/sbp006. Epub 2009 Mar 26.

HSU, D.T.; CHEN, F.L.; TAKAHASHI, L.K.; KALIN, N.H. Rapid stress-induced elevations in corticotropin-releasing hormone mRNA in rat central amygdala nucleus and hypothalamic paraventricular nucleus: an in situ hybridization analysis. **Brain Res** 1998; 788: 305-310.

JOHNSTON, J.B. Further contributions to the study of the evolution of the forebrain. **Anatomical Laboratory, University of Minnesota** 1923.

KREAM, J.; MULAY, S.; FUKUSHIMA, D.K.; SOLOMON, S. Determination of plasma dexamethasone in the mother and the newborn after administration of the hormone in a clinical trial. **J Clin Endocrinol Metab.** 1983; 56(1):127-33.

LAARIS, N.; HAJDAHMANE, S.; HAMON, M.; LANFUMEY, L. Glucocorticoid receptor-mediated inhibition by corticosterone of 5-HT1A autoreceptor functioning in the rat dorsal raphe nucleus. **Neuropharmacology.** 1995; 34:1201-121

LANGLEY-EVANS, S.C. Intrauterine programming of hypertension by glucocorticoids. **Life Sci.** 1997; 60:1213-21.

LANGLEY-EVANS, S.C.; JACKSON, A.A. Captopril normalizes systolic blood pressure in rats with hypertension induced by fetal exposure to maternal low protein diets. **Comp Biochem Physiol A Physiol.** 1995; 110:223-28.

LANGLEY-EVANS, S.C.; WELHAM, S.J.; SHERMAN, R.C.; JACKSON, A.A. Weanling rats exposed to maternal low-protein diets during discrete periods of gestation exhibit differing severity of hypertension. **Clin Sci (Lond).** 1996; 91:607-15.

LANGLEY-EVANS.; S.C.; JACKSON, A.A. Increased systolic blood pressure in adult rats induced by fetal exposure to maternal low protein diets. **Clin Sci (Lond)**, 1994; 86:217-22,

LEONARD, B.E. The HPA and immune axes in stress: the involvement of the serotonergic system. **Eur. Psychiatry.** 2005; 20:302–306.

LEVITSKY DA; BARNES RH. Nutritional and environmental interactions in the behavioral development of the rat: long-term effects. **Science.** 1972 Apr 7;176(4030):68-71.

LIMA JG, OLIVEIRA LM, LACHAT JJ, DAL-BO CM, ALMEIDA SS. Comparison of the effects of lab chow and casein diets based on body and brain development of rats. **Braz J Med Biol Res.** 1993 Oct;26(10):1069-76.

LINGAS, R.; DEAN, F.; MATTHEWS, S.G. Maternal nutrient restriction (48h) modifies brain corticosteroid receptor expression and endocrine function in the fetal guinea pig. **Brain Res**, 1999; 846: 236-242.

LOWRY, C.A.; HALE, M.W.; EVANS, A.K.; HEERKENS, J.; STAUB, D.R.; GASSER, P.J. Serotonergic systems, anxiety, and affective disorder: focus on the

dorsomedial part of the dorsal raphe nucleus. **Ann NY Acad Sci** 2008; 1148:86–94.

LOWRY, C.A.; JOHNSON, P.L.; HAY-SCHMIDT, A.; MIKKELSEN, J.; SHEKHAR A. Modulation of anxiety circuits by serotonergic systems. **Stress** 2005; 8:233–46.

LUCAS A. Programming by early nutrition in man. **Ciba Found Symp.** 1991;156:38-50; discussion 50-5.

MACEY D.J.; SMITH, H.R.; NADER, M.A.; PORRINO, L.J. Chronic cocaine self-administration upregulates the norepinephrine transporter and alters functional activity in the bed nucleus of the stria terminalis of the rhesus monkey. **J Neurosci.** 2003; 1,23(1):12-6.

MAKINO, S.; GOLD, P.;W.; SCHULKIN, J. Corticosterone effects on corticotropin-releasing hormone messenger-RNA in the central nucleus of the amygdala and the parvocellular region of the paraventricular nucleus of the hypothalamus. **Brain Res** 1994; 640: 105-112.

MASON WA, HARLOW HF, RUEPING RR. The development of manipulatory responsiveness in the infant rhesus monkey. **J Comp Physiol Psychol.** 1959 Oct;52:555-8.

MATSUMOTO, A.; ARAI, Y. Sexual differentiation of neuronal circuitry in the neuroendocrine hypothalamus. **Biomed Rev** 1997; 7: 5-15.

MCDONALD AJ. Neurons of the bed nucleus of the stria terminalis: a golgi study in the rat. **Brain Res Bull.** 1983 Jan;10(1):111-20.

MCEWEN, B.S. Glucocorticoid±biogenic amine interactions in relation to mood and behavior. **Biochem Pharmacol** 1987; 36: 1755-1763.

MELIS, M; SPIGA, S; DIANA, M. The Dopamine Hypothesis of Drug Addiction: Hypodopaminergic State. **Int Rev Neurobiol.** 2005;63:101-54.

MERALI Z, KHAN S, MICHAUD, D.S.; SHINOSITOL, PHOSPHATEPY SA, ANISMAN H. Does amygdaloid corticotropin-releasing hormone (CRF) mediate anxiety-like behaviors? Dissociation of anxiogenic effects and CRF release. *Eur. J. Neurosci.* 2004; 20:229–239.

MILLAN, M.J. Serotonin 5-HT<sub>2C</sub> receptors as a target for the treatment of depressive and anxious states: focus on novel therapeutic strategies. *Therapie.* 2005; 60:441–460.

MORGANE PJ, MOKLER DJ, GALLER JR. Effects of prenatal protein malnutrition on the hippocampal formation. *Neurosci Biobehav Rev.* 2002 Jun;26(4):471-83.

MULLER, M.B. Limbic corticotropin-releasing hormone receptor 1 mediates anxiety-related behavior and hormonal adaptation to stress. *Nat. Neurosci.* 2003; 6:1100–1107.

NESTLER, E.J.; BARROT, M.; DILEONE, R.J.; EISCH, A.J.; GOLD, S.J.; MONTEGGIA, L.M. Neurobiology of depression. *Neuron.* 2002; 34:13–25.

OADES, RD; SADILE, AG; SAGVOLDEN, T; VIGGIANO, D; ZUDDAS, A; DEVOTO, P; AASE, H; JOHANSEN, EB; RUOCCO, LA; RUSSELL VA. The control of responsiveness in ADHD by catecholamines: evidence for dopamin, noradrenergic and interactive roles. *Dev Sci.* 2005 Mar;8(2):122-31.

OLIVEIRA M; RODRIGUES AJ; LEÃO P; CARDONA D; PÊGO JM; SOUSA N. The bed nucleus of stria terminalis and the amygdala as targets of antenatal glucocorticoids: implications for fear and anxiety responses. *Psychopharmacology* (Berl). 2012 Apr;220(3):443-53. doi: 10.1007/s00213-011-2494-y. Epub 2011 Sep 21.

OLIVEIRA M; BESSA JM; MESQUITA A; TAVARES H; CARVALHO A; SILVA R; PÊGO JM; CERQUEIRA JJ; PALHA JA; ALMEIDA OF; SOUSA N. Induction of a hyperanxious state by antenatal dexamethasone: a case for less detrimental

natural corticosteroids. **Biol Psychiatry**. 2006 May 1;59(9):844-52. Epub 2005 Sep 28.

ONAKA, T.; YAGI, K. Role of noradrenergic projections to the bed nucleus of the stria terminalis in neuroendocrine and behavioral responses to fear-related stimuli in rats. **Brain Res**. 1998; 788:287– 293.

ORGANIZAÇÃO DAS NAÇÕES UNIDAS PARA A ALIMENTAÇÃO E A AGRICULTURA (FAO), Fundo Internacional para o Desenvolvimento Agrícola (IFAD) e Programa Alimentar Mundial (PAM). The State of Food Insecurity in the World. 2015. <http://www.fao.org/3/a-i4646e.pdf>

PAINTER RC, ROSEBOOM TJ, BLEKER OP. Prenatal exposure to the Dutch famine and disease in later life: An overview. **Reproductive Toxicology** 20 (2005) 345–352.

PÊGO, J. M.; MORGADO, P.L.G.; CERQUEIRA, J.J., ALMEIDA, O.F.X.; SOUSA, N. Dissociation of the morphological correlates of stress-induced anxiety and fear. **European Journal of Neuroscience** 2008; 27:1503–1516.

PERSSON, E.; JANSSON, T. Low birth weight is associated with elevated adult blood pressure in the chronically catheterized guinea-pig. **Acta Physiol Scand**. 1992; 145(2):195-6.

PIAZZA PV; LE MOAL ML. Pathophysiological basis of vulnerability to drug abuse: role of an interaction between stress, glucocorticoids, and dopaminergic neurons. **Annu Rev Pharmacol Toxicol**. 1996;36:359-78.

PRENTICE, A.M. Can maternal dietary supplements help in preventing infant malnutrition? **Acta Paediatr Scand**. 1991; Suppl.374:67-77.

PREWITT, C.M.F.; HERMAN, J.P. Anatomical interactions between the central amygdaloid nucleus and the hypothalamic paraventricular nucleus of the rat—a dual tract-tracing analysis. **J. Chem. Neuroanat**. 1998; 15:173– 185.

PROCTER, S B.; Campbell CG. Position of the Academy of Nutrition and Dietetics: Nutrition and Lifestyle for a Healthy Pregnancy Outcome. **J Acad Nutr Diet.** 2014; 114(7):1099-103. doi: 10.1016/j.jand.2014.05.005.

RAPER J, SHANNON B.Z. STEPHENS, AMY HENRY, TRINA VILLARREAL, JOCELYNE BACHEVALIER, KIM WALLEN AND MAR M. SANCHEZ. Neonatal Amygdala Lesions Lead to Increased Activity of Brain CRF Systems and Hypothalamic-Pituitary-Adrenal Axis of Juvenile Rhesus Monkeys **The Journal of Neuroscience** 2014 • 34(34):11452–11460

RINCÓN-CORTÉS M, SULLIVAN RM.  
Early life trauma and attachment: immediate and enduring effects on neurobehavioral and stress axis development. **Front Endocrinol (Lausanne).** 2014 Mar 21;5:33. doi: 10.3389/fendo.2014.00033. eCollection 2014.

ROSE G. Familial patterns in ischaemic heart disease. **Br J Prev Soc Med.** 1964 Apr;18:75-80.

ROSEBOOM T, ROOIJ S, PAINTER R. The Dutch famine and its long-term consequences for adult health. **Early Human Development** (2006) 82, 485—491.

ROSEBOOM TJ, VAN DER MEULEN JHP, RAVELLI ACJ, OSMOND C, BARKER DJP, BLEKER OP. Effects of prenatal exposure to the Dutch famine on adult disease in later life: an overview. **Molecular and Cellular Endocrinology** 185 (2001) 93–98.

SCHULKIN, J.; GOLD, P.W.; MCEWEN, B.S. Induction of corticotropin-releasing hormone gene expression by glucocorticoids: implication for understanding the states of fear and anxiety and allostatic load. *Psychoneuroendocrinology.* 1998; 23(3):219-43. **Review.**

SCHULKIN, J.; MCEWEN, B.S.; GOLD, P.W. Allostasis, amygdala, and anticipatory angst. **Neurosci Biobehav Rev** 1994; 18: 385-396.

SCHUMANN CM, BAUMAN MD, AMARAL DG. Abnormal structure or function of the amygdala is a common component of neurodevelopmental disorders. **Neuropsychologia** (2011) 49:745–759.

SECKL, J.R. Physiologic programming of the fetus, **Clin. Perinatol.** 1998; 25: 939–964.

SETH, S; LEWIS, A J; SAFFERY, R; LAPPAS, M; GALBALLY, M. Maternal Prenatal Mental Health and Placental 11 $\beta$ HSD2 Gene Expression: Initial Findings from the Mercy Pregnancy and Emotional Wellbeing Study. **Int J Mol Sci.** 2015 Nov 17;16 (11):27482-96. doi: 10.3390/ijms161126034. *Int J Mol Sci.* 2015 Nov; 16(11): 27482–27496.

SHOLL, D.A. The measurable parameters of the cerebral cortex and their significance in its organization. **Prog Neurobiol** . 1956; 2:324–333.

STEWART, P.M.; WHORWOOD, C.B.; MASON, J.I. Type 2 11 beta-hydroxysteroid dehydrogenase in foetal and adult life. **J Steroid Biochem Mol Biol.** 1995; 55(5-6):465-71

STOUT, S.C.; MORTAS, P.; OWENS, M.J.; NEMEROFF, C.B.; MOREAU, J. Increased corticotropin-releasing factor concentrations in the bed nucleus of the stria terminalis of anhedonic rats. **Eur J Pharmacol.** 2000; 28;401(1):39-46.

TORRES, GE; GAINETDINOV, RR; CARON, MG. Plasma membrane monoamine transporters: structure, regulation and function. **Nat Rev Neurosci.** 2003 Jan;4(1):13-25.

TOTTENHAM N, HERTZIG ME, GILLESPIE-LYNCH K, GILHOOLY T, MILLNER AJ, CASEY BJ. Elevated amygdala response to faces and gaze aversion in autism spectrum disorder. **Soc Cogn Affect Neurosci.** 2014 Jan;9(1):106-17. doi: 10.1093/scan/nst050. Epub 2013 Apr 16.

ULRICH-LAI YM, HERMAN JP.  
 Neural regulation of endocrine and autonomic stress responses. **Nat Rev Neurosci.** 2009 Jun;10(6):397-409. doi: 10.1038/nrn2647.

UYLINGS, H.B.M.; VAN, P.J. Measures for quantifying dendritic arborizations. **Network: comput neural syst** 2002 13:397–414

VYAS, A.; BERNAL, S.; CHATTARJI, S. Effects of chronic stress on dendritic arborization in the central and extended amygdala. **Brain Res.** 2003; 7;965(12):290-4.

VYAS, A.; MITRA, R.; SHANKARANARAYANA, R.B.S.; CHATTARJI S. Chronic stress induces contrasting patterns of dendritic remodeling in hippocampal and amygdaloid neurons. **Neurosci.** 2002 1:22(15)6810-8.

WALKER, D.L.; MILES, L.A.; DAVIS, M. Selective participation of the bed nucleus of the stria terminalis and CRF in sustained anxiety-like versus phasic fear-like responses. *Prog Neuropsychopharmacol Biol Psychiatry.* 2009 Nov 13;33(8):1291-308. **Epub** 2009; 10. Review.

WALKER, D.L.; TOUFEXIS, D.J.; DAVIS, M. Role of the bed nucleus of the stria terminalis versus the amygdale in fear, stress, and anxiety. **European Journal of Pharmacology** 2003; 463:199– 216

WATTS, A.G.; SANCHEZ-WATTS, G. Region-specific regulation of neuropeptide mRNAs in rat limbic forebrain neurones by aldosterone and corticosterone. **J Physiol.** 1995; 484:721-36.

WATTS, A.G.; SANCHEZ-WATTS, G. Region-specific regulation of neuropeptide mRNAs in rat limbic forebrain neurones by aldosterone and corticosterone. **J Physiol.** 1995; 484:721-36.

WELBERG, L.A.; SECKL, J.R. Prenatal stress, glucocorticoids and the programming of the brain. **J Neuroendocrinol** 2001; 13: 113-128.



WELBERG, L.A.M.; SECKL, J.R.; HOLMES, M.C. Inhibition of 11 $\beta$ -hydroxysteroid dehydrogenase, the feto-placental barrier to maternal glucocorticoids, permanently programs amygdala GR mRNA expression and anxietylike behaviour in the offspring. **Eur J Neurosci** 2000; 12:1047-1054.

WELBERG, L.A.M.; SECKL, J.R.; HOLMES, M.C. Inhibition of 11 $\beta$ -hydroxysteroid dehydrogenase, the feto-placental barrier to maternal glucocorticoids, permanently programs amygdala GR mRNA expression and anxietylike behaviour in the offspring. **Eur J Neurosci** 2000; 12:1047-1054.

WELLER, K.L.; SMITH, D.A. Afferent connections to the bed nucleus of the stria terminalis **Brain Research** 1982; 232: 255-270

WOODALL, S.M.; JOHNSTON, B.M.; BREIER, B.H.; GLUCKMAN, P.D. Chronic maternal undernutrition in the rat leads to delayed postnatal growth and elevated blood pressure of offspring. **Pediatr Res.** 1996; 40: 438-443.

WORLD HEALTH ORGANIZATION. Global targets 2025 to improve maternal, infant and young child nutrition. Available from: [http://www.who.int/nutrition/topics/nutrition\\_globaltargets2025/en/index.html](http://www.who.int/nutrition/topics/nutrition_globaltargets2025/en/index.html) ( cited 30 May 2013).

ZHU, W.; UMEGAKI, H.; SUZUKI, Y.; MIURA, H.; IGUCHI, A. Involvement of the bed nucleus of the stria terminalis in hippocampal cholinergic system-mediated activation of the hypothalamo – pituitary – adrenocortical axis in rats. **Brain Res.** 2001; 916:101– 106.

## 8. ANEXO

### 8.1 Certificado Comitê de Ética- Unicamp



CEUA/UNICAMP

#### CERTIFICADO

Certificamos que o projeto intitulado "Análise dos efeitos da restrição proteica in utero no BNST e na amígdala de ratos: Estudo da estrutura dendrítica neural, de parâmetros funcionais e moleculares", protocolo nº 3908-1, sob a responsabilidade de Dra. Patricia Aline Boer / Daniele Braz Torres, que envolve a produção, manutenção e/ou utilização de animais pertencentes ao filo *Chordata*, subfilo *Vertebrata* (exceto o homem) para fins de pesquisa científica ou ensino, encontra-se de acordo com os preceitos da **LEI Nº 11.794, DE 8 DE OUTUBRO DE 2008**, que estabelece procedimentos para o uso científico de animais e do **DECRETO Nº 6.899, DE 15 DE JULHO DE 2009**, e com as normas editadas pelo **Conselho Nacional de Controle da Experimentação Animal - CONCEA**, e foi aprovado pela **Comissão de Ética no Uso de Animais da Universidade Estadual de Campinas - CEUA/UNICAMP**, em 31 de julho de 2015.

**Vigência do projeto:** 08/2015-02/2017

**Espécie/Linhagem:** Rato heterogênico Wistar

**No. de animais:** 48

**Peso/Idade:** 04 semanas / 250gr

**Sexo:** 24 machos / 24 fêmeas

**Origem:** CEMIB/UNICAMP



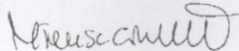
A aprovação pela CEUA/UNICAMP não dispensa autorização prévia junto ao **IBAMA**, **SISBIO** ou **CIBio**.

Campinas, 31 de julho de 2015.

Profa. Dra. Liana Maria Cardoso Verinaud  
Presidente

Fátima Alonso  
Secretária Executiva

## 8.2 Certificado Comitê de Ética-ICVS

 GOVERNO DE PORTUGAL MINISTÉRIO DA AGRICULTURA E DO MAR			
Ex <sup>ma</sup> Senhora <b>Doutora Magda João Castelhana Carlos</b> Escola de Ciências da saúde / Instituto de Investigação em Ciências da Vida e Saúde Campus de Gualtar 4710 – 057 BRAGA - P			
2013-08-30 023432	Nossa referência	Vossa referência	Vossa data
Assunto: <span style="border: 1px solid black; padding: 2px;">           PROTEÇÃO DOS ANIMAIS UTILIZADOS PARA FINS EXPERIMENTAIS E/OU            OUTROS FINS CIENTÍFICOS – PEDIDO DE AUTORIZAÇÃO PARA            REALIZAÇÃO DE PROJECTO DE EXPERIMENTAÇÃO ANIMAL         </span>			
<p>Na sequência do pedido efetuado por V. Ex<sup>a</sup> no sentido de poder ser autorizada a realização do projeto experimental designado “Efeitos programadores do stress”, tendo como investigadora responsável a <b>Doutora Ana João Rodrigues</b>, cabe-me informar que o mesmo foi levado à consideração dos membros da Comissão Consultiva prevista na alínea b) do n.º 49, da Portaria n.º 1005/92, de 23 de Outubro, sendo que os mesmos não levantaram qualquer objeção à solicitação supra referida.</p>			
<p>Mais se informa V. Ex<sup>a</sup> que esta Direção Geral, também, nada teve a opôr ao projeto apresentado, pelo que, o mesmo foi autorizado, ao abrigo do n.º 8.º do mesmo diploma legislativo.</p>			
<p>Com os melhores cumprimentos,</p>			
A Directora Geral			
			
As) Maria Teresa Villa de Brito			
DBEA/APM			
